

# Update on Non-immediate Drug Allergy Reactions: State and New Aspects

Enrique Gomez<sup>1\*</sup>, Natalia Blanca-Lopez<sup>2</sup>

<sup>1</sup>Pharmaceutical Science, Roche Pharma Research and Early Development, Roche Innovation Centre, Basel, F. Hoffmann-La Roche Ltd., Switzerland; <sup>2</sup>Department of Allergy, Infanta Leonor University Hospital, Madrid, Spain

## ABSTRACT

**Purpose of the review:** Among the Adverse Drug Reactions (ADR), around 10-15% are driven by an immunological mechanism and considered as allergic drug reactions. Within the Type IV of drug allergic reactions and in accordance with Gell and Coombs classification, the non-immediate drug hypersensitivity reactions (NI-DHR) relates to the most complex group of drug allergy, being cellular mediated responses and appearing from 1 hour to several weeks after drug/metabolites exposure. Current diagnosis protocols are limited and there is an unmet need to identify a diagnostic approach that mimic the pathological response and enhance the possibilities for a more accurate and realistic diagnosis.

**Recent findings:** Changes in gene patterns induced during the acute phase of the NI-DHR provide clues of the underlying immunological mechanisms, while the study and identification of specific HLA profiles in selected patients allows making inference about the risk of suffer a reaction.

**Conclusion:** Advances on the knowledge of NI-DHR, based on genetic and transcriptomic analysis, will provide better understanding of the biology behind, as well as more opportunities to diagnose and treat the patients

**Keywords:** Allergy; Drug; Non-immediate reactions; T-cells

**Abbreviations:** ADR: Adverse Drug Reactions; NI-DHR: Non-immediate Drug Hypersensitivity Reactions; NIR: Non-immediate Reactions; DHR: Drug Hypersensitivity Reactions; MPE: Maculopapular Exanthema; NI-U: Non-immediate Urticaria; AGEP: Acute Generalized Exanthematic Pustulosis; SJS: Stevens-Johnson Syndrome; TEN: Toxic Epidermal Necrolysis; DRESS: Drug Reaction with Eosinophilia and Systemic Symptoms; ICM: Iodinated Contrast Media; BLs: Betalactams; CYP: Cytochrome P; NSAIDs: Non-Steroidal Anti-inflammatory Drugs; IR: Immediate Reactions; APC: Antigen-presenting Cells; FQ: Fluoroquinolones; IS: Immune System; CS: Corticosteroids; CLA: Cutaneous LymphocyteA; DPT: Drug Provocation Test; PT: Patch Tests; ST: Skin Testing; IDT: Intradermal Test; LTT: Lymphocyte Transformation Test; CR: Cross-reactivity.

## INTRODUCTION

Among the Adverse Drug Reactions (ADR), around 10%-15% are driven by an immunological mechanism and considered as allergic drug reactions [1]. Categorized within Type IV of allergic drug reactions accordingly with Gel and Coombs classification [2], the Non-immediate Drug Hypersensitivity Reactions (NI-DHR) relates to the most complex group of drug allergy, being cellular mediated and with responses appearing more than 1 hour after drug/metabolites exposure. Current diagnosis protocols are limited and there is an unmet need to identify a diagnostic approach that mimic the pathological response and enhance the possibilities of an accurate diagnosis.

In this review, authors would like to update previous published work in this topic [3], include most recent discoveries on NI-DHR and providing their view on relevant aspects of the diagnosis.

## CLASSIFICATION OF T-CELL DRUG REACTIONS

The NI-DHR is T-cell mediated reactions classified in four categories, although recently a fifth one has been proposed. The clinical manifestations are wide, affecting different organs, with severity ranging from mild reactions like Maculopapular Exanthema (MPE) or Non-immediate Urticaria (NI-U), to life-threatening reactions like Acute Generalized Exanthematic Pustulosis (AGEP), bullous reactions (Stevens-Johnson Syndrome (SJS)/TEN) or DRESS (Table 1) [4-8].

**Correspondence to:** Enrique Gómez, Pharmaceutical Science, Roche Pharma Research and Early Development, Roche Innovation Centre, Basel F. Hoffmann-La Roche Ltd., Switzerland, Tel: +41 76 321 40 37; Email: enriquegomezalcaide@gmail.com

**Received:** June 14, 2021; **Accepted:** June 28, 2021; **Published:** July 05, 2021

**Citation:** Gomez E, Lopez NB (2021) Update on Non-immediate Drug Allergy Reactions: State and New Aspects. J Dermatitis. 6: 133.

**Copyright:** © 2021 Gomez E, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited..

**Table 1:** More relevant clinical entities and mechanisms involved in NIR.

Type of reaction	Mechanism implied	Clinical manifestations
Non systemic reactions	a) T cell specific/Toxic metabolites	1. Maculopapular Exanthema (MPE) 2. Contact Dermatitis (CD) 3. Photosensitivity contact dermatitis 4. Isolated Mucosal Involvement 5. Bullous/Desquamative exanthema 6. Fix Drug Eruption (FDE) 7. Non-Immediate Urticaria (NIU)
	b) Toxicological	1. Bile duct syndrome 2. Hepatitis 3. Meningitis 4. Pneumonitis 5. Nephritis 6. Pancreatitis 7. Other organ-specific clinical manifestations
Systemic reactions	a) T cell specific/heterologous immunity/HLA haplotypes	1. Serum Sickness Like Syndrome (SSLS) / accelerated urticaria 2. Severe cutaneous adverse reactions 3. Drug Reaction with Eosinophilia and Systemic Symptoms (DRESS) 4. Acute Generalized Exanthematic Pustulosis (AGEP) 5. Toxic Epidermal Necrolysis (TEN) 6. Stevens Johnson Syndrome (SJS)
	b) Unknown	Vasculitis
Located reactions	Vascular occlusion/T cell specific	Nicolau syndrome

## DRUGS INVOLVED

Any drug or its reactive metabolites can induce a NI-DHR [9]. Classical drug/drug-metabolites eliciting this type of reactions are anticonvulsants, Iodinated Contrast Media (ICM), Betalactams (BLs) or Non-steroidal Anti-Inflammatory Drugs (NSAIDs). However, other chemical compounds have shown the capacity of triggering them [3].

## ANTICONVULSANTS

Anticonvulsants are among the most frequent drugs eliciting NI-DHR. It is well demonstrated that for these compounds the bioactive metabolites generated during the drug-metabolism by different enzymes belonging to the cytochrome P450 (CYP) are responsible of the induced reaction, where sub-families CYP2C and CYP2E are involved [10-15].

## IODINATED CONTRAST MEDIA

ICM are widely involved in DHR, are inert chemical drugs that contain iodine atoms used for x-ray-based imaging. Based on their chemical structure, osmolarity, iodine content and ionization degree in solution they are classified in ionic or non-ionic, being the latest the most frequent eliciting NI-DHR [16-18]. Although most of reactions are specific to one CM compound, it has been shown an extensive cross-reactivity among them [19,20].

## BETALACTAMS

BLs is also involved in NIR, with some clinical entities like accelerated urticaria causing confusion [21]. In IR the specificity of the antibodies has been well proven, existing good correlation with clinical entities [22-24], but in NIR the final structure recognized is more complex to define [25]. All BLs bind spontaneously to proteins by nucleophilic attack of their amino groups. Amoxicillin forms penicilloyl adducts

with lysine residues on human serum albumin [26]. Other BLs like benzylpenicillin, aztreonam or piperacillin binds to similar lysine residues [27]. This also occurs with other BLs as clavulanic, although the metabolites generated are different and do not cross-react with classical BLs. Regarding cephalosporins, the lack of knowledge of the chemical structure of their antigenic determinants and the proteins involved in the sensitization makes difficult the understanding of the mechanisms involved [28]. However, whether differential binding occurs and if this influences the specificity of adducts is not well known. It has been shown that the hapten-specific IgG antibodies found in piperacillin-hypersensitive patients do not bind to other BL protein conjugates. Both hapten and carrier contribute to the formation of the antigen [24,29]. In the case of T-cell mediated responses the BL is recognized after the formation of a BL-peptide complex formed after the processing of the adduct by the antigen-presenting cells (APC). BLs happenizes extra and intracellular proteins. One study has found that heat shock protein 70 and enolase can be haptenized by amoxicillin [30]. These adducts can be incorporated into target cells transported by exosome.

## NON-STEROIDAL ANTI-INFLAMMATORY DRUGS

NSAIDs can also induce NIR, which are including in the phenotype of single NSAIDs-induced delayed hypersensitivity reactions. This comprises a wide spectrum of pathologies that goes from mild to severe reactions. Concerning the different NSAIDs, although some entities like photo contact dermatitis are more likely to be induced aryl-propionics or oximes, all of them can induce NIR [4,31-38].

Regarding the underlying mechanisms, drug-specific T-cell clones have not yet been developed. Concerning drug metabolites there have been identified for pyrazolones, although only few studies are focused on analysing their antigenicity [39,40].

## QUINOLONES, PARTICULARLY FLUORQUINOLONES (FQ)

Quinolones, particularly fluorquinolones are one of the main drugs involved in DHR worldwide [41-43] inducing IgE- and T-cell reactions. FQ have one pyridone and another aromatic ring. The structural differences within this group are derived from the number and position of nitrogen and fluoride atoms, and the side chains. All types of reactions (table), including photo-allergy, can be induced with differences depending on the FQ [44,45].

## SULFONAMIDES

Sulfonamide were the first group of drugs involved in DHR, and nowadays continue eliciting reactions, being the most common clinical entity elicited MPE, although urticaria may also occur. NIR to sulfonamides are mainly caused by metabolites, particularly with nitro-sulphometoxazol. Around 45%-70% of sulfonamides are acetylated in the liver by N-acetyltransferase and approximately 15% glucuronidated. Moreover, 10% is hydroxylated by the cytochrome CYP2C9 to sulfamethoxazole hydroxylamine. The metabolite nitroso-sulfamethoxazole is a model for T-cell activation. More details about the generation of metabolites are shown in references [46-48].

## MACROLIDS

Macrolids are other antibiotics that can be involved in NIR [49], but less is known about the metabolites generated and the recognition by the Immune System (IS) than with the antibiotics detailed above.

## OTHER DRUGS

### Abacavir

The elicits also NI-DHR, particularly associated with HIV infection [50]. The metabolism comprises phase II glucuronidation mediated by uridine diphosphate glucanoyltransferase, which yields an inactive glucuronide metabolite (abacavir-glucuronide), and phase I oxidation that yields a carboxylate (abacavir-carboxylate) [51].

### Oxipurinol

It is the main metabolite of allopurinol after transformation by xanthine oxidase. Excess production of this drug is assumed to lead to tissue damage, evoking an immunological response with development of antibodies against tissue-components [52].

### Corticosteroids

Inhibit the allergic response by suppressing the transcription of a variety of pro-inflammatory cytokines like IL-12 and IFN- $\gamma$ . However, they are more common involved in NIR than in IR. Although contact dermatitis has been the most frequently reported, reactions to parenterally administered drugs have also been published. The more frequently involved are betamethasone, dexamethasone and triamcinolone, although other like prednisone and metilprednisolone are also implicated. Though anecdotic, severe reactions like SJS/TEN or AGEP are published [53-60].

## DRUG-RECOGNITION AND IMMUNE RESPONSE

Drugs are molecules under 1Kda that are not visible to the IS, evolved to recognize antigens with a molecular weight above 1Kda.

There are 3 hypothesis explaining the molecular events behind the recognition of drugs by the IS [61].

I The hapten model: drug binds to a peptide/protein forming an adduct recognizable.

II. The danger hypothesis focuses on danger-signals released as consequence of a tissue damage created by the drug. These signals would activate APCs and eventually trigger the immune response [62].

III. The pharmacological interaction (p-i) model proposes interaction drug-immune receptors like drug-specific T-cell receptors or HLA molecules present on APCs. The reversible interaction drug: MHC-TTCR complexes would induce structural changes leading the response. However, this model was ineffective in mounting a primary response; therefore it is considered as complementary to the hapten model [63].

NI-DHR can be associated to certain HLA-I alleles. This modifies the antigen clef location allowing the recognition of new peptides otherwise not recognized. In the case of abacavir, the binding to HLA-B\*57:01 is able to re-shape the antigen-binding location and inducing CD8-mediated responses. Several alleles have been proposed for carbamazepine, with the strongest association reported with HLA-B\*15:02 and HLA-A\*31:01. Phenytoin have shown a genomic association with HLA-B\*15:02 HLA-B\*56:02, and HLA-B\*51:01, while lamotrigine is associated to HLA-A\*38:01, HLA-A\*24:02 or HLA-B\*15:02. Many other DHR have been associated with HLA-responses [64-72].

## MONITORING THE ACUTE RESPONSE

Monitoring acute phase of the reaction allows for instance the Identification of increased levels of circulating T-cells expressing the Cutaneous Lymphocyte Antigen (CLA) or the presence of skin chemokines like CTACK/CCL27, involved in the recruitment of

CCR10+ lymphocytes [73-77], provides valuable information about the mechanisms involved. Comparison between subjects developing IR versus NI-DHR shows a polarized immune response, with high expression of IL-12 and IFN- $\gamma$  and down-regulation of IL-4 (Th1 pattern), opposite to subjects with IR (Th2 pattern) [78]. Similarly, gene expression analysis of lymphocytes from NI-DHR patients showed high expression of TNF- $\alpha$ , perforin, Granzyme B and Fas-L, with higher levels in more severe responses [79], using microarray technology, demonstrated a differential expression of 85 genes during the acute phase, with overexpression of alarmins, suggesting that in severe reactions drugs can trigger this proteins [80].

## DIAGNOSTIC APPROACHES

Although for NIR the diagnosis relies mainly in the clinical history or in a DPT when indicated, positive intradermal, photocontact or Patch Tests (PT) have been reported [81]. *In vitro* tests, although useful, do not have appropriate sensitivity nor verified specificity [24,82].

The clinical history is always the first step; the more precise it is the more accurate will be the diagnosis. Important data to be collected are: Drugs involved, dose administered, duration of treatment, detailed description of symptoms presented, time interval between drug/drugs administration and reaction, treatment received and time to recovery after it. Important cofactors to be considered are underlying infections. Nevertheless, frequently data recorded are retrospective and imprecise, the clinical history is not always reliable. The European Academy Allergy Clinical Immunology-Drug Allergy interest Group/European Network for Drug Allergy (EAACI-DAIG/ENDA) has provided a detailed questionnaire useful for diagnosing DHR [83].

Concerning ST, the most used approaches are Intradermal Test (IDT) and/or PT with delayed readings. Details of how to perform them can be found elsewhere. The PT, applicable with non-soluble drugs, consists of the dilution of the drug in a vehicle for adequate skin absorption, being the photo-patch a modification recommended for photoallergic reactions. The IDT, recommended for soluble drugs, have higher sensitivity than PT, though in most studies published it is no higher than 50%-60%. Indications are available for BLs, CM, heparins and CS, amongst others. Recommendations for readings as well as the precautions to be taken to avoid risks are detailed elsewhere [84-94].

The cellular *in vitro* tests are considered for supporting the diagnosis of NI-DHR. The Lymphocyte Transformation Test (LTT) is based on flow cytometry technology that not only helps in measuring the proliferative capacity of drug-specific T-cells, but to identify which subpopulations are involved in the reaction, providing clues of the underlying mechanism. Conveniently, the LTT allows the evaluation in one single experiment of several compounds from the same drug-family/drug-metabolites that could induce cross-reactions [95-100]. A limitation of the test is its variable sensitivity, as demonstrated in studies with BLs with values from 62% to 74.4%. However, it must be kept in mind its complementary nature, since some results could lead to misdiagnosis. This is the case with penicilins, where has been published how some patients may respond exclusively to amoxicillin or BP, while others react to both compounds, which cannot be extrapolated to the response *in vivo*, where subjects with positive response to BP and negative to amoxicillin, can tolerate BP and react to amoxicillin. A 2nd generation of LTT has shown improvement of the sensitivity, based on the co-cultivation of T-cells with autologous monocyte-derive dendritic cells from patients in presence of the culprit drug [101-103]. However, the increased complexity limits its applicability.



DPT is the gold standard for the diagnosis, although it is only indicated in mild reactions. It is used to exclude drug allergy or to de-label cases diagnosed as drug allergy. If DPT is not performed a large number of cases can be overdiagnosed. It is particularly useful in children where the most frequent reactions are MPE. In fact, the number of cases finally confirmed after DPT is below 10%. Therefore, according to recent studies, it has been proposed to perform directly DPT without ST in cases with mild NIR [104-108].

Regarding the dosages and time intervals recommended, they depend on the drug, the route of administration and the severity of the reaction. Other conditions to consider are comorbidities and co-mediations [93]. For children doses are adjusted to weight and age. The procedure consist in administering escalating doses of the drug till reaching the full therapeutic dose, followed in negative cases by a prolonged challenge [96, 109-112].

## ASSESSMENT OF CROSS-REACTIVITY (CR)

A lot of information is available concerning CR with IgE antibodies between penicillins and cephalosporins, and within the penicillin group. Assessment is based on the knowledge of the chemical structure and no information has been provided concerning the protein carrier. Data published show high degree of CR between amoxicillin and ampicillin, although some cases are also positive to benzylpenicillin [109]. Regarding FQ three patterns of CR have been proposed: 1) T-cells reacting to the eliciting drug, 2) limited CR and 3) broad CR. Published studies conclude that CR is quite frequent.

Concerning sulfonamides, the structure of the functional group is unknown being the assessment of CR difficult. All these antibiotics contain structural characteristics that are absent from non-antibiotic ones. The CR appears to be due to a predisposition to allergic reactions rather than CR with sulfonamide-based drugs [111]. The metabolism of sulfonamide is specific to certain molecular substituents and stereospecific. Since it involves structures contained only in the sulfonamides, similar metabolites are not formed and CR must be regarded as highly unlikely.

Evaluation of CR among corticoids is also difficult. Frequently, individuals do not remember whether they have received CS previously. Within this group, it is difficult to confirm if we are dealing with CR or co-sensitization. Based on PT results and their chemical structure, classified CS in four groups: A (hydrocortisone type), B (triamcinolone acetonide type), C (betamethasone type), and D (hydrocortisone-17-butyrate type), with subdivision into groups (D1, D2) depending on the presence/absence of a C16-methyl substitution and/or halogenation on the C9 of the B-ring. It is known that high CR exists within each group as well as between groups D2, A and B, being CR with D1 quite low. This classification is especially useful with topically CS, although not accepted worldwide. Furthermore, other rings can be relevant for determining CR, showing substitutions at C6/9 and C16/17 sites important in inducing NI-DHR [113-116]. Regarding systemic reactions it is even more difficult to define patterns of CR. Succinate-ester seems to have more immunogenicity, probably because its capacity to bind to arginine groups from proteins similar to glyoxal derivatives.

## DISCUSSION AND CONCLUSION

NI-DHR comprises a heterogeneous group of clinical entities potentially induced by any drug. More studies are needed to decipher the interaction between immune and skin cells in NI-DHR. Likewise,

it is important to delve into the discovery of the drug metabolites and the mechanisms behind their generation on the skin, most likely by keratinocytes, with demonstrated expression of CYP-proteins that could act as APC.

There is an unmet for models to characterize the haptenome in cases of sensitization by the oral/parenteral route, as for contact dermatitis.

The LTT demonstrated its value in studying NI-DHR and a 3rd variation, integrating skin cells, would be desirable.

The monitorization of the acute phase and the integration of transcriptomics have proved its value in deciphering the underlying mechanisms in DHRs. The application of high-density expression platforms represents a more integrative way for providing a complete view of gene expression.

However, the lack of replication studies and the heterogeneity of the clinical entities have led to dispersed results and difficulties to stable genetic biomarkers related to DHRs. Further investigations are needed to identify genes and polymorphisms related to these disorders that could be useful for an accurate diagnosis

## REFERENCES

1. Romano A, WJ Pichler, Blanca M. Highlights of the 4th Drug Hypersensitivity Meeting-Rome, April 22-25, 2010. Preface. *J Allergy Clin Immunol*. 2011 ; 127(3): 59.
2. Gell PGH, RRA Coombs. Clinical aspects of immunology. *Brit Med J*. 1968 ; 1(55) : 597-602.
3. E Gómez, M Ruano, ML Somoza, J Fernández, N Blanca-López. Role of T cells in non-immediate drug allergy reactions. *Curr Opin Allergy Clin Immunol*. 2019; 19(4): 294-301.
4. Natalia Blanca-Lopez, Maria L Somoza-Alvarez, Teresa Bellon, Gemma Amo, Gabriela Canto, Miguel Blanca. NSAIDs hypersensitivity: questions not resolved. *Curr Opin Allergy Clin Immunol*. 2018 ; 18(4): 291-301.
5. Pichler WJ. Delayed drug hypersensitivity reactions. *Ann Intern Med*. 2003 ; 139(8): 683-693.
6. Markus Britschgi, Salome von Greyerz, Christoph Burkhart, Werner J Pichler. Molecular aspects of drug recognition by specific T cells. *Curr Drug Targets*. 2003 ; 4(1): 1-11.
7. Redwood A, J. Trubiano, E.J. Phillips. Prevention and Diagnosis of Severe T-Cell-Mediated Adverse Drug Reactions: Are We There Yet?. *J Allergy Clin Immunol Pract*. 2019; 7(1): 228-230.
8. R Cabanas, E Ramirez, E Sendagorta, R Alamar, R Barranco, N Blanca-López, et al., Spanish Guidelines for Diagnosis, Management, Treatment, and Prevention of DRESS Syndrome. *J Investig Allergol Clin Immunol*. 2020 ; 30(4): 229-253.
9. Andrew Sullivan, Andrew Gibson, Brian Kevin Park, Dean J Naisbitt. Are drug metabolites able to cause T-cell-mediated hypersensitivity reactions?. *Expert Opin Drug Metab Toxicol*. 2015; 11(3): 357-368.
10. Gwendolin S. Simper, Gia-Gia T. Hò, Alexander A. Celik, Trevor Huyton, Joachim Kuhn , Heike Kunze-Schumacher, et al., Carbamazepine-Mediated Adverse Drug Reactions: CBZ-10,11-epoxide but Not Carbamazepine Induces the Alteration of Peptides Presented by HLA-B \*15:02. *J Immunol Res*. 2018.
11. Ying Wu, Joseph P Sanderson, John Farrell, Nicola S Drummond, Anita Hanson, Elizabeth Bowkett, et al., Activation of T cells by carbamazepine and carbamazepine metabolites. *J Allergy Clin Immunol*. 2006; 118(1): 233-241.

12. Leeder JS. Mechanisms of idiosyncratic hypersensitivity reactions to antiepileptic drugs. *Epilepsia*. 1998 ; 39(7): 8-16.
13. Wolf HH, EA Swinyard, LS Goodman. Anticonvulsant properties of some N-substituted hydantoins. *J Pharm Sci*. 1962 ; 51 : 74-76.
14. Kumi Sakurada, Takeshi Kozaru, Kazuo Yamada, Masashi Nibuya, Kiyoshi Nagata, Eiji Suzuki. Allergy to chlorpromazine and valproic acid following carbamazepine hypersensitivity in a patient with an HLA-B\*4601 allele. *Neuropsychiatr Dis Treat*. 2018; 14: 1139-1142.
15. Francesca Mori, Natalia Blanca-Lopez, Jean-Christoph Caubert, Pascal Demoly, George Du Toit, Eva R Gomes, et al., Delayed hypersensitivity to antiepileptic drugs in children. *Pediatr Allergy Immunol*. 2021 ; 32(3): 425-436.
16. Francesco Lapi, Enrica Cecchi, Claudio Pedone, Francesco Attanasio, Grazia Banchelli, Alfredo Vannacci, et al., Safety aspects of iodinated contrast media related to their physicochemical properties: a pharmacoepidemiology study in two Tuscany hospitals. *Eur J Clin Pharmacol*. 2008; 64(7): 723-737.
17. Enrique Gómez, Adriana Ariza, Natalia Blanca-López, Maria J Torres. Nonimmediate hypersensitivity reactions to iodinated contrast media. *Curr Opin Allergy Clin Immunol*. 2013; 13(4): 345-353.
18. Brockow K. Immediate and delayed cutaneous reactions to radiocontrast media. *Chem Immunol Allergy*. 2012; 97: 180-90.
19. Marianne Lerch, Monika Keller, Markus Britschgi, Gisele Kanny, Valerie Tache, Daphne A Schmid, et al., Cross-reactivity patterns of T cells specific for iodinated contrast media. *J Allergy Clin Immunol*. 2007; 119(6): 1529-1536.
20. Angèle Soria, Emmanuelle Amsler, Claire Bernier, Brigitte Milpied, Florence Tétart, Cécile Morice, et al., DRESS and AGEP Reactions to Iodinated Contrast Media: A French Case Series. *J Allergy Clin Immunol Pract*. 2021; S2213-2198(21) : 318-314.
21. Enrique Gómez, Natalia Blanca-Lopez, Maria Salas, Gabriela Canto, Paloma Campo, Maria J Torres, et al., Induction of accelerated reactions to amoxicillin by T-cell effector mechanisms. *Ann Allergy Asthma Immunol*. 2013 ; 110(4): 267-273.
22. M Blanca, C Mayorga, MJ Torres, R Warrington, A Romano, P Demoly, et al., Side-chain-specific reactions to betalactams: 14 years later. *Clin Exp Allergy*. 2002; 32(2): 192-197.
23. F Moreno, M Blanca, C Mayorga, S Terrados, M Moya, E Pérez, et al., Studies of the specificities of IgE antibodies found in sera from subjects with allergic reactions to penicillins. *Int Arch Allergy Immunol*. 1995; 108(1): 74-81.
24. Cristobalina Mayorga, Didier G Ebo, David M Lang, Werner J Pichler, Vito Sabato, Miguel A Park, et al., Controversies in drug allergy: *In vitro* testing. *J Allergy Clin Immunol*. 2019; 143(1): 56-65.
25. M E Azoury, L Fili, R Bechara, N Scornet, L de Chaisemartin, R J Weaver, et al., Identification of T-cell epitopes from benzylpenicillin conjugated to human serum albumin and implication in penicillin allergy. *Allergy*. 2018; 73(8): 1662-1672.
26. Xiaoli Meng, Caroline Jane Earnshaw, Arun Tailor, Rosalind Jenkins, James C. Waddington, Paul Whitaker, et al., Amoxicillin and Clavulanate Form Chemically and Immunologically Distinct Multiple Haptenic Structures in Patients. *Chem Res Toxicol*. 2016; 29(10): 1762-1772.
27. Mohammed O Amali, Rosalind E Jenkins, Xiaoli Meng, Lee Faulkner, Paul Whitaker, Daniel Peckham, et al., Assessment of Antipiperacillin IgG Binding to Structurally Related Drug Protein Adducts. *Chem Res Toxicol*. 2017 ; 30(12): 2097-2099.
28. Fernandez J, TW Jimenez-Rodriguez, N Blanca-Lopez. Classifying cephalosporins: from generation to cross-reactivity. *Curr Opin Allergy Clin Immunol*. 2021.
29. FJ Sánchez-Gómez, JM González-Morena, Y Vida, E Pérez-Inestrosa, M Blanca, MJ Torres, et al., Amoxicillin haptens intracellular proteins that can be transported in exosomes to target cells. *Allergy*. 2017; 72(3): 385-396.
30. A Ariza, C Mayorga, TD Fernandez, N Barbero, A Martín-Serrano, D Pérez-Sala, et al., Hypersensitivity reactions to beta-lactams: relevance of hapten-protein conjugates. *J Investig Allergol Clin Immunol*. 2015; 25(1): 12-25.
31. M L Kowalski, R Asero, S Bavbek, M Blanca, N Blanca-Lopez, G Bochenek, et al., Classification and practical approach to the diagnosis and management of hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy*. 2013; 68(10): 1219-1232.
32. Blanca-López N, Pérez-Sánchez N, Agúndez JA, García-Martin E, Torres MJ, Cornejo-García JA, et al., Allergic Reactions to Metamizole: Immediate and Delayed Responses. *Int Arch Allergy Immunol*. 2016; 169(4): 223-230.
33. Koransky R, D Ferastraoraru, E. Jerschow. Single nonsteroidal anti-inflammatory drug induced serum sickness-like reaction to naproxen in a patient able to tolerate both aspirin and ibuprofen. *J Allergy Clin Immunol Pract*. 2016; 4(1): 160-161.
34. Lourdes Arochena, María Paz Zafra, Ma Carmen Fariña, Victoria Del Pozo, Mar Fernández-Nieto. Acute generalized exanthematic pustulosis due to ibuprofen. *Ann Allergy Asthma Immunol*. 2013; 110(5): 386-387.
35. Y Cherif, Moez Jallouli, M Mseddi, H Turki, Z Bahloul. Acute generalized exanthematous pustulosis induced by piroxicam: a case report. *Indian J Pharmaco*. 2014. 46(2): 232-233.
36. Nihira T, Y Hagiwara. Ketoprofen-induced photoallergic dermatitis. *Pediatr Int*. 2019; 61(6): 610-611.
37. Marie-Sara Marchand, Elisabeth Autret-Leca, Etienne Bourdais, Annie-Pierre Jonville-Béra. [Photoallergy to piroxicam: what should be contraindicates?]. *Thérapie*. 2013; 68(1): 57-58.
38. Satoshi Imai, Kenji Atarashi, Koichi Ikese, Katsuhiko Akiyama, Yoshiki Tokura. Establishment of murine model of allergic photocontact dermatitis to ketoprofen and characterization of pathogenic T cells. *J Dermatol Sci*. 2006; 41(2): 127-136.
39. Levy M, E Zylber-Katz, B Rosenkranz. Clinical pharmacokinetics of dipyrone and its metabolites. *Clin Pharmacokinet*. 1995; 28(3): 216-234.
40. C Herdeg, F Hilt, A Büchtemann, L Bianchi, R Klein. Allergic cholestatic hepatitis and exanthema induced by metamizole: verification by lymphocyte transformation test. *Liver*. 2002; 22(6): 507-513.
41. I Doña, E Barrionuevo, N Blanca-Lopez, M J Torres, T D Fernandez, C Mayorga, et al., Trends in hypersensitivity drug reactions: more drugs, more response patterns, more heterogeneity. *J Investig Allergol Clin Immunol*. 2014; 24(3): 143-153.
42. Blanca-Lopez N, I Andreu, MJ Torres Jaen. Hypersensitivity reactions to quinolones. *Curr Opin Allergy Clin Immunol*. 2011; 11(4): 285-291.
43. A Rosado Ingelmo, I Dona Diaz, R Cabanas Moreno, MC Moya Quesada, C García-Aviles, I Garcia Nunez, et al., Clinical Practice Guidelines for Diagnosis and Management of Hypersensitivity Reactions to Quinolones. *J Investig Allergol Clin Immunol*. 2021;

- 26(3) : 144-155.
44. Schmid DA, JP Depta, WJ Pichler. T cell-mediated hypersensitivity to quinolones: mechanisms and cross-reactivity. *Clin Exp Allergy*. 2006; 36(1): 59-69.
  45. Andreu I, C Mayorga, MA Miranda. Generation of reactive intermediates in photoallergic dermatitis. *Curr Opin Allergy Clin Immunol*. 2010; 10(4): 303-308.
  46. Brackett CC, H Singh, JH Block. Likelihood and mechanisms of cross-allergenicity between sulfonamide antibiotics and other drugs containing a sulfonamide functional group. *Pharmacotherapy*. 2004; 24(7): 856-870.
  47. Lee Faulkner, Andrew Gibson, Andrew Sullivan, Arun Tailor, Toru Usui, Ana Alfirevic, et al., Detection of Primary T Cell Responses to Drugs and Chemicals in HLA-Typed Volunteers: Implications for the Prediction of Drug Immunogenicity. *Toxicol Sci*. 2016; 154(2): 416-429.
  48. J Luis Castrejon, Neil Berry, Sabah El-Ghaiesh, Basil Gerber, Werner J Pichler, B Kevin Park, et al., Stimulation of human T cells with sulfonamides and sulfonamide metabolites. *J Allergy Clin Immunol*. 2010; 125(2): 411-418.
  49. M El Khoury, H Assier, G Gener, M Paul, C Haddad, O Chosidow, et al., Polysensitivity in delayed cutaneous adverse drug reactions to macrolides, clindamycin and pristinamycin: clinical history and patch testing. *Br J Dermatol*. 2018; 179(4): 978-979.
  50. Nádia M Grilo, Alexandra MM Antunes, Umbelina Caixas, Aline T Marinho, Catarina Charneira, M Conceicao Oliveira, et al., Monitoring abacavir bioactivation in humans: screening for an aldehyde metabolite. *Toxicol Lett*. 2013; 219(1): 59-64.
  51. Nádia M Grilo, Catarina Charneira, Sofia A Pereira, Emilia C Monteiro, M Matilde Marques, Alexandra MM Antunes. Bioactivation to an aldehyde metabolite-possible role in the onset of toxicity induced by the anti-HIV drug abacavir. *Toxicol Lett*. 2014; 224(3): 416-423.
  52. Rima Shalom, Sofia Rimbroth, Dganit Rozenman, Arie Markel. Allopurinol-induced recurrent DRESS syndrome: pathophysiology and treatment. *Ren Fail*. 2008; 30(3): 327-329.
  53. Butani L. Corticosteroid-induced hypersensitivity reactions. *Ann Allergy Asthma Immunol*. 2002; 89(5): 439-445.
  54. Joshua R Mann, Jana Mannan, Luis Antonio Quinones, Allyson A Palmer, Myriam Torres. Religion, spirituality, social support, and perceived stress in pregnant and postpartum Hispanic women. *J Obstet Gynecol Neonatal Nurs*. 2010; 39(6): 645-657.
  55. P Chaves, M J Torres, A Aranda, S Lopez, G Canto, M Blanca, et al., Natural killer-dendritic cell interaction in lymphocyte responses in hypersensitivity reactions to betalactams. *Allergy*. 2010; 65(12): 1600-1608.
  56. A Padial, S Posadas, J Alvarez, M-J Torres, J A Alvarez, C Mayorga, et al., Nonimmediate reactions to systemic corticosteroids suggest an immunological mechanism. *Allergy*. 2005; 60(5): 665-670.
  57. L Fernández de Corrés, I Urrutia, M Audicana, S Echechipia, G Gastaminza. Erythroderma after intravenous injection of methylprednisolone. *Contact Dermatitis*. 1991; 25(1): 68-70.
  58. Whitmore SE. Delayed systemic allergic reactions to corticosteroids. *Contact Dermatitis*. 1995; 32(4): 193-198.
  59. Andersen KE, N Hjorth, T Menne. The baboon syndrome: systemically-induced allergic contact dermatitis. *Contact Dermatitis*. 1984; 10(2): 97-100.
  60. MT Ventura, GF Calogiuri, L Muratore, E Di Leo, R Buquicchio, A Ferrannini. Cross-reactivity in cell-mediated and IgE-mediated hypersensitivity to glucocorticoids. *Curr Pharm Des*. 2006; 12(26): 3383-3391.
  61. Katie D White, Wen-Hung Chung, Shuen-Iu Hung, Simon Mallal, Elizabeth J Phillips. Evolving models of the immunopathogenesis of T cell-mediated drug allergy: The role of host, pathogens, and drug response. *J Allergy Clin Immunol*. 2015; 136(2): 219-234.
  62. Ferial Hacini-Rachinel, Mercedes Gomez de Agüero, Reem Kanjarawi, Ludovic Moro-Sibilot, Jean-Benoit Le Luduec, Claire Macari, et al., Intestinal dendritic cell licensing through Toll-like receptor 4 is required for oral tolerance in allergic contact dermatitis. *J Allergy Clin Immunol*. 2018; 141(1): 163-170.
  63. Pichler WJ. The p-i Concept: Pharmacological Interaction of Drugs With Immune Receptors. *World Allergy Organ J*. 2008; 1(6): 96-102.
  64. Patricia T Illing, Julian P Vivian, Nadine L Dudek, Lyudmila Kostenko, Zhenjun Chen, Mandvi Bharadwaj, et al., Immune self-reactivity triggered by drug-modified HLA-peptide repertoire. *Nature*. 2012; 486(7404): 554-558.
  65. Chaichon Lochareonkul, Jakrin Loplumlert, Chusak Limotai, Wiwat Korkij, Tayard Desudchit, Siraprapa Tongkobpetch, et al., Carbamazepine and phenytoin induced Stevens-Johnson syndrome is associated with HLA-B\*1502 allele in Thai population. *Epilepsia*. 2008; 49(12): 2087-2091.
  66. Mark Mc Cormack, Ana Alfirevic, Stephane Bourgeois, John J Farrell, Dalia Kasperaviciute, Mary Carrington, et al., HLA-A\*31:01 and carbamazepine-induced hypersensitivity reactions in Europeans. *N Engl J Med*. 2011; 364(12): 1134-1143.
  67. E Genin, DP Chen, S-I Hung, P Sekula, M Schumacher, PY Chang, et al., HLA-A\*31:01 and different types of carbamazepine-induced severe cutaneous adverse reactions: an international study and meta-analysis. *Pharmacogenomics J*. 2014; 14(3): 281-288.
  68. Elena Ramirez, Teresa Bellón, Hoi Y Tong, Alberto M Borobia, Francisco J de Abajo, Victoria Lerma, Miguel A Moreno Hidalgo, et al., Significant HLA class I type associations with aromatic antiepileptic drug (AED)-induced SJS/TEN are different from those found for the same AED-induced DRESS in the Spanish population. *Pharmacol Res*. 2017; 115: 168-178.
  69. Taisei Mushiroda, Yukitoshi Takahashi, Teiichi Onuma, Yoshiaki Yamamoto, Tetsumasa Kamei, Tohru Hoshida, et al., Association of HLA-A\*31:01 Screening With the Incidence of Carbamazepine-Induced Cutaneous Adverse Reactions in a Japanese Population. *JAMA Neurol*. 2018; 75(7): 842-849.
  70. Ryan J Schutte, Yonghu Sun, Danmeng Li, Furen Zhang, David A Ostrov, et al., Human Leukocyte Antigen Associations in Drug Hypersensitivity Reactions. *Clin Lab Med*. 2018; 38(4): 669-677.
  71. Phillips EJ. New Strategies to Predict and Prevent Serious Immunologically Mediated Adverse Drug Reactions. *Trans Am Clin Climatol Assoc*. 2018; 129: 74-87.
  72. Jason H Karnes, Matthew A. Miller, Katie D White, Katherine Konvinse, Rebecca Pavlos, Alec J Redwood, et al., Applications of Immunopharmacogenomics: Predicting, Preventing, and Understanding Immune-Mediated Adverse Drug Reactions. *Annu Rev Pharmacol Toxicol*. 2019; 59: 463-486.
  73. C Antúnez, MJ Torres, C Mayorga, JA Cornejo-García, LF Santamaría-Babi, M Blanca, et al., Different cytokine production and activation marker profiles in circulating cutaneous-lymphocyte-associated antigen T cells from patients with acute or chronic atopic dermatitis. *Clin Exp Allergy*. 2004; 34(4): 559-566.



74. Michael C Peters, Lando Ringel, Nathan Dyjack, Rachel Herrin, Prescott G Woodruff, Cydney Rios, et al., A Transcriptomic Method to Determine Airway Immune Dysfunction in T2-High and T2-Low Asthma. *Am J Respir Crit Care Med*. 2019; 199(4): 465-477.
75. Grégory Seumois, Jose Zapardiel-Gonzalo, Brandie White, Divya Singh, Veronique Schulten, Myles Dillon, et al., Transcriptional Profiling of Th2 Cells Identifies Pathogenic Features Associated with Asthma. *J Immunol*. 2016; 197(2): 655-664.
76. M Blanca, S Posadas, MJ Torres, L Leyva, C Mayorga, L Gonzalez, et al., Expression of the skin-homing receptor in peripheral blood lymphocytes from subjects with nonimmediate cutaneous allergic drug reactions. *Allergy*. 2000; 55(11): 998-1004.
77. Beatriz Tapia, Antonia Padial, Elena Sánchez-Sabaté, Javier Alvarez-Ferreira, Esther Morel, Miguel Blanca et al., Involvement of CCL27-CCR10 interactions in drug-induced cutaneous reactions. *J Allergy Clin Immunol*. 2004; 114(2): 335-340.
78. SJ Posadas, L Leyva, MJ Torres, JL Rodriguez, I Bravo, M Rosal, et al., Subjects with allergic reactions to drugs show in vivo polarized patterns of cytokine expression depending on the chronology of the clinical reaction. *J Allergy Clin Immunol*. 2000; 106(4): 769-776.
79. Sinforiano J Posadas, Antonia Padial, Maria J Torres, Cristobalina Mayorga, Laura Leyva, Elena Sanchez, et al., Delayed reactions to drugs show levels of perforin, granzyme B, and Fas-L to be related to disease severity. *J Allergy Clin Immunol*. 2002; 109(1): 155-161.
80. T Bellón, L Alvarez, C Mayorga, E Morel, MJ Torres, MA Martín-Díaz, et al., Differential gene expression in drug hypersensitivity reactions: induction of alarmins in severe bullous diseases. *Br J Dermatol*. 2010; 162(5):1014-1022.
81. Waheed A, T Hill, N. Dhawan. *Drug Allergy. Prim Care*. 2016; 43(3): 393-400.
82. Ruben Fernandez-Santamaria, Gador Bogas, Francisca Palomares, Maria Salas, Tahia D Fernandez, Isabel Jimenez, et al., Dendritic cells inclusion and cell-subset assessment improve flow-cytometry-based proliferation test in non-immediate drug hypersensitivity reactions. *Allergy*. 2021; 76(7): 2123-2134.
83. P Demoly, R Kropf, A Bircher, WJ Pichler. Drug hypersensitivity: questionnaire. EAACI interest group on drug hypersensitivity. *Allergy*. 1999; 54(9): 999-1003.
84. Elizabeth J Phillips, Paul Bigliardi, Andreas J Bircher, Ana Broyles, Yoon-Seok Chang, Wen-Hung Chung, et al., Controversies in drug allergy: Testing for delayed reactions. *J Allergy Clin Immunol*. 2019; 143(1): 66-73.
85. M Blanca, A Romano, MJ Torres, J Fernández, C Mayorga, J Rodriguez, et al., Update on the evaluation of hypersensitivity reactions to betalactams. *Allergy*. 2009; 64(2): 183-193.
86. Antonino Romano, Rocco Luigi Valluzzi, Cristiano Caruso, Michela Maggioletti, Francesco Gaeta. Non-immediate Cutaneous Reactions to Beta-Lactams: Approach to Diagnosis. *Curr Allergy Asthma Rep*. 2017; 17(4): 23.
87. Barbaud A. Skin testing in delayed reactions to drugs. *Immunol Allergy Clin North Am*. 2009; 29(3): 517-535.
88. A Barbaud, M Gonçalo, D Bruynzeel, A Bircher. Guidelines for performing skin tests with drugs in the investigation of cutaneous adverse drug reactions. *Contact Dermatitis*. 2001; 45(6): 321-328.
89. A Padial, C Antunez, N Blanca-Lopez, TD Fernandez, JA Cornejo-Garcia, C Mayorga, et al., Non-immediate reactions to beta-lactams: diagnostic value of skin testing and drug provocation test. *Clin Exp Allergy*. 2008; 38(5): 822-828.
90. Marie Baeck, Julie-Anne Chemelle, Raphaël Terreur, Jacques Drieghe, An Goossens. Delayed hypersensitivity to corticosteroids in a series of 315 patients: clinical data and patch test results. *Contact Dermatitis*. 2009; 61(3): 163-1675.
91. Sae-Hoon Kim, Eun-Jung Jo, Mi-Yeong Kim, Seung-Eun Lee, Min-Hye Kim, Min-Suk Yang, et al., Clinical value of radiocontrast media skin tests as a prescreening and diagnostic tool in hypersensitivity reactions. *Ann Allergy Asthma Immunol*. 2013; 110(4): 258-262.
92. Elina Tan, Grace Thompson, Charlotta Ekstrom, Michaela Lucas. Non-immediate heparin and heparinoid cutaneous allergic reactions: a role for fondaparinux. *Intern Med J*. 2018; 48(1): 73-77.
93. K Brockow, A Romano, M Blanca, J Ring, W Pichler, P Demoly. General considerations for skin test procedures in the diagnosis of drug hypersensitivity. *Allergy*. 2002; 57(1): 45-51.
94. Eleni Papakonstantinou, Sabine Müller, Jan H Röhrbein, Dorothea Wiczorek, Alexander Kapp, Thilo Jakob, et al., Generalized reactions during skin testing with clindamycin in drug hypersensitivity: a report of 3 cases and review of the literature. *Contact Dermatitis*. 2018; 78(4): 274-280.
95. Maria Isabel Montañez, Cristobalina Mayorga, Gador Bogas, Esther Barrionuevo, Ruben Fernandez-Santamaria, Angela Martin-Serrano, et al., Epidemiology, Mechanisms, and Diagnosis of Drug-Induced Anaphylaxis. *Front Immunol*. 2017; 8: 614.
96. I Luque, L Leyva, M José Torres, M Rosal, C Mayorga, J M Segura, et al., *In vitro* T-cell responses to beta-lactam drugs in immediate and nonimmediate allergic reactions. *Allergy*. 2001; 56(7): 611-618.
97. Francisca Vilchez-Sánchez, David Loli-Ausejo, Amelia Rodriguez-Mariblanca, Jaime Montserrat-Villatoro, Elena Ramírez, Javier Domínguez-Ortega, et al., Lymphocyte transformation test can be useful for the diagnosis of delayed adverse reactions to sulfonamides. *Allergy*. 2020; 75(12): 3267-3272.
98. R Cabañas, O Calderón, E Ramírez, A Fiandor, T Caballero, R Heredia, et al., Sensitivity and specificity of the lymphocyte transformation test in drug reaction with eosinophilia and systemic symptoms causality assessment. *Clin Exp Allergy*. 2018; 48(3): 325-333.
99. Francesca Mori, Lucia Fili, Lucrezia Sarti, Manuela Capone, Giulia Liccioli, Mattia Giovannini, et al., Sensitivity and specificity of lymphocyte transformation test in children with mild delayed hypersensitivity reactions to beta-lactams. *Allergy*. 2020; 75(10): 2696-2699.
100. M Movsisyan, A Fiandor, M González-Muñoz, S Quirce, T Bellón, A Hakobyan, et al., The Lymphocyte Transformation Test Is Useful in the Diagnosis of Fixed Drug Eruption Induced by Etoricoxib. *J Invest Allergol Clin Immunol*. 2019; 29(4): 307-309.
101. Nyfeler B, WJ Pichler. The lymphocyte transformation test for the diagnosis of drug allergy: sensitivity and specificity. *Clin Exp Allergy*. 1997; 27(2): 175-181.
102. Rebeca Rodriguez-Pena, Soledad Lopez, Cristobalina Mayorga, Cristina Antunez, Tahia D Fernandez, Maria J Torres, et al., Potential involvement of dendritic cells in delayed-type hypersensitivity reactions to beta-lactams. *J Allergy Clin Immunol*. 2006; 118(4): 949-956.
103. Soledad Lopez, Maria J Torres, Cristina Antunez, Rebeca Rodríguez-Pena, Gabriela Canto, Miguel Blanca, et al., Specific immunological response to budesonide in a patient with delayed-type hypersensitivity reaction. *J Invest Dermatol*. 2010; 130(3): 895-897.
104. N Blanca-López, L Zapatero, E Alonso, MJ Torres, V Fuentes, MI Martínez-Molero, et al., Skin testing and drug provocation in the diagnosis of nonimmediate reactions to aminopenicillins in children. *Allergy*. 2009; 64(2): 229-233.

105. W Aberer, A Bircher, A Romano, M Blanca, P Campi, J Fernandez, et al., Drug provocation testing in the diagnosis of drug hypersensitivity reactions: general considerations. *Allergy*. 2003; 58(9): 854-863.
106. Maria A Zambonino, Jose Luis Corzo, Candelaria Muñoz, Gloria Requena, Adriana Ariza, Cristobalina Mayorga, et al., Diagnostic evaluation of hypersensitivity reactions to beta-lactam antibiotics in a large population of children. *Pediatr Allergy Immunol*. 2014; 25(1): 80-87.
107. ER Gomes, K Brockow, S Kuyucu, F Saretta, F Mori, N Blanca-Lopez, et al., Drug hypersensitivity in children: report from the pediatric task force of the EAACI Drug Allergy Interest Group. *Allergy*. 2016; 71(2): 149-161.
108. Moral L, JC Caubet. Oral challenge without skin tests in children with non-severe beta-lactam hypersensitivity: Time to change the paradigm? *Pediatr Allergy Immunol*. 2017; 28(8): 724-727.
109. S Terrados, M Blanca, J Garcia, J Vega, MJ Torres, MJ Carmona, et al., Nonimmediate reactions to betalactams: prevalence and role of the different penicillins. *Allergy*. 1995; 50(7): 563-567.
110. Depta JP, WJ Pichler. Cross-reactivity with drugs at the T cell level. *Curr Opin Allergy Clin Immunol*. 2003; 3(4): 261-267.
111. Brian L Strom, Rita Schinnar, Andrea J Apter, David J Margolis, Ebbing Lautenbach, Sean Hennessy, et al., Absence of cross-reactivity between sulfonamide antibiotics and sulfonamide nonantibiotics. *N Engl J Med*. 2003; 349(17): 1628-1635.
112. Coopman S, H Degreef, A Doooms-Goossens. Identification of cross-reaction patterns in allergic contact dermatitis from topical corticosteroids. *Br J Dermatol*. 1989; 121(1): 27-34.
113. DS Wilkinson, S Fregert, B Magnusson, HJ Bandmann, CD Calnan, E Cronin, et al., Terminology of contact dermatitis. *Acta Derm Venereol*. 1970; 50(4): 287-292.
114. Xiao-Wei Cai, Rong Zhu, Lei Ran, Yi-Qian Li, Ke Huang, Jing Peng, et al., A novel noncontact communication between human keratinocytes and T cells: Exosomes derived from keratinocytes support superantigeninduced proliferation of resting T cells. *Mol Med Rep*. 2017; 16(5): 7032-7038.
115. Erika Parkinson, Maja Aleksic, Richard Cubberley, Gushinder Kaur-Atwal, Johannes PC Vissers, Paul Skipp. Determination of Protein Haptenation by Chemical Sensitizers Within the Complexity of the Human Skin Proteome. *Toxicol Sci*. 2018; 162(2): 429-438.
116. Jörg Bartel, Jan Krumsiek, Katharina Schramm, Jerzy Adamski, Christian Gieger, Christian Herder, et al., The Human Blood Metabolome-Transcriptome Interface. *PLoS Genet*. 2015; 11(6): e1005274.