

# T cell Migration and Graft Versus Host Disease

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## Abstract

Graft-Versus-Host Disease (GVHD) remains the major obstacle to a more favorable therapeutic outcome of allogeneic hematopoietic stem cell transplantation (HSCT). GVHD is mediated by immunocompetent donor T cells. The mature allo-reactive T cells, either CD4<sup>+</sup> T cells or CD8<sup>+</sup> T cells, within the "graft", can mediate GVHD. In this review article, we also describe the GVHD pathophysiologic events after bone marrow transplantation, including GVHD target recipient organs with the distribution of donor T cells, and distribution kinetics of donor T cells with accompanying cytokine expression.

**Keywords:** Graft-Versus-Host-Disease (GVHD); T cell migration

## Introduction

Allogeneic Bone Marrow Transplantation (Allo-BMT) is a curative therapy for leukemia, aplastic anemia and immune deficiencies. The alloreactive donor T-cells that induce graft-versus-leukemia (GVL) effect may also initiate graft-versus-host disease (GVHD). GVHD is a serious problem that limits the use of allogeneic BMT. GVHD is mediated by immunocompetent donor T cells, which migrate to lymphoid tissues soon after infusion, recognize host alloantigens, and become activated upon interaction with host antigen presenting cells (APCs). Although the pathophysiologic mechanisms of GVHD still remains unclear now, it has been reported that GVHD develops in three consecutive stages: (1) Pre-transplant conditioning results in inflammation coupled with a cytokine storm; (2) Activation of donor T-cells; (3) Finally, the activated donor T-cells assault certain tissues, such as Intestine, skin, liver, and lungs [1]. The occurrence and severity of GVHD depend on several factors, including the intensity of conditioning, the presence and number of donor T-cells in the graft, and the antigenic disparity between donor and recipient [2,3]. This review addresses the distribution, dynamic of activation and migration pattern of donor T cells in GVHD.

## Either CD4<sup>+</sup> T cells or CD8<sup>+</sup> T cells can Initiate Lethal GVHD

In Allogeneic transplantations, in most cases, are performed between HLA-matched sibling's donors. The alloresponse is directed to minor histocompatibility antigens (miHA) expressed on host tissues [4-6]. The miHA are processed self-protein degradation products that can be presented in association with either major histocompatibility complex (MHC) class I or class II molecules on host parenchymal and antigen-presenting cells (APC), resulting in stimulation of donor CD4<sup>+</sup> and CD8<sup>+</sup> T cells, respectively [7-9]. Studies have shown that either the recipient's or donor's DCs can present allo-antigens to donor T-cells [10,11]. Allogeneic donor T cells with the same MHC molecules can thus recognize miHA that is not expressed by themselves. During the process of GVHD, activated donor T-cells migrate to target tissue and induce GVHD via either direct cell contactor (cytotoxic T-cells) or cytokine mediated toxicity (T-helper cells) [12]. Our group found that either CD4<sup>+</sup> T cells or CD8<sup>+</sup> T cells could initiate lethal GVHD independently in Allo-BMT mouse model. Friedman et al. [13] analyzed the T-cell responses after transplantation by CDR3-size spectratyping in B6 --> BALB/C Allo-BMT model. They revealed clonal or oligoclonal expansions of the V $\beta$  2, 4, and 6 to 14 families

for the CD4<sup>+</sup> response and of the V $\beta$  4, 6, 8 to 11, and 14 families for the B6 CD8<sup>+</sup> response. Appropriate positive selection of these T-cell receptor V $\beta$ -skewed CD4<sup>+</sup> and CD8<sup>+</sup> T-cell subsets and their subsequent transfer into lethally irradiated BALB.B recipients resulted in fatal GVHD induction. In contrast, BALB B mice transplanted with non-skewed V $\beta$  T cells survived, with minimal symptoms of GVHD. This indicates that there is special T cell subsets- T cell receptor V $\beta$ -skewed T cells, in CD4<sup>+</sup>T cells or CD8<sup>+</sup>T cells, which involve in GVHD. It should be the special T cell subsets, T cell receptor V $\beta$ -skewed T cells in both CD4<sup>+</sup> T cells and CD8<sup>+</sup> T cells, initiate lethal GVHD in the Allo-BMT mouse model.

## T cell Migration in GVHD

Panoskaltis-Mortari et al. [14] tracked the migration of eGFP transgenic donor cells post-transplant in a fully MHC-mismatched murine allo-BMT model. Within 2~3 days after transplantation, allogeneic T cells expanded in lymphoid tissues. Between 3 and 7 days post-transplant, allogeneic T cell numbers increased in GVHD target organs including the gastro-intestinal (GI) tract, liver, lungs, skin, central nervous system, gingiva, and nasal mucosal. In our study, donor T cells infiltrated the liver, spleen, skin, lungs, intestine, tongue, and small amounts eGFP<sup>+</sup> cells were noted in the kidney and brain; but no eGFP<sup>+</sup> cells were seen in the cardiac muscle or skeletal muscle [15]. These indicated that GVHD attacked not only liver, intestines and skin, but also lungs, tongue, and even kidneys or brain. For example, lungs were possibly the important GVHD target organ, which has been reported [16]. It has been found in clinic that GVHD was closely related to interstitial pneumonia. Perhaps, interstitial pneumonia post Allo-BMT was resulted from the secondary infection on the basis of GVHD immunologic injury of lungs [17]. GVHD involvement of the central nervous system and the brain has also been previously described in mice [18] and humans [19].

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Data demonstrated, in early post-transplant, donor T cells presented in lymphoid organ, such as spleen, but not in non-lymphoid organs such as liver and skin [20,21]. The sphingosine-1-phosphate receptor inhibitor FTY720, which prevents lymphocyte egress from lymphoid organs, inhibited target organ infiltration and GVHD lethality in a murine model, suggesting that these donor T cells migrated after previous activation in lymphoid tissues [22,23].

## IL-2 and CD25 Expression during T cell Migration

IL-2 is an important cytokine in immunologic responses. Secretion begins at 45 minutes after T-cell activation. CD25, the  $\alpha$ -chain of the IL-2 receptor, is expressed at 2 hours after T-cell activation, combining to build a trimer with the  $\beta$  and  $\gamma$ -chains on the cellular membrane. Then, IL-2 combines to the receptor transmitting and signal for activation, proliferation, and differentiation [24]. As second signal of pCTL, the precursor cell of cytotoxic T lymphocytes, IL-2 combines with membrane CD25 on pCTL for activation, leading to antigenic-specific amplification, as antigenic specific CTL. Where T cells became effectors, the population showing with high CD25 expression and high IL-2 secretion disappears. Possible mechanisms of the process include down-regulation of IL-2 or CD25 expression and or activation-induced cell death after completing the immunologic response. Using mouse GVHD model, Via et al. [25] established a mouse GVHD model, and found the peak expression IL-2 on day 2~3 post transplantation, and decreased afterward. This short period of IL-2 expression on early stage of BMT also was confirmed by detecting of IL-2 mRNA by RT-PCR [26,27]. It was possible that the high expression of IL-2 and CD25 in the early stage of transplantation play a role in T cell activation.

Our group found recently the highest level of T cell population in spleen at day+4 was synchronized with the peak expression of CD25 by the donor T cells, and the highest serum levels of IL-2 [28]. Thus, it was highly possible that these T cells were activated in spleen and migrated to GVHD target organs, such as liver, GI tract, lungs, skin and so on, inducing tissue damage and the appearance of clinical manifestations. In fact, we noted that from days 2 to 16 donor T cells decreased in the spleen and increased in the liver [28].

These data support the donor T-cell migration hypothesis: During the development of GVHD, donor T cells migrate to lymphoid organs, such as the spleen, where they are activated. Thereafter they migrate to GVHD target organs to induce damage.

## References

- Teshima T, Ferrara JLM (2002) Understanding the alloresponse: new approaches to graft-versus-host disease prevention. *Semin Hematol* 39(1):15-22.[PubMed]
- Pérez-Simón JA, Díez-Campelo M, Martino R, Brunet S, Urbano A, et al. (2005) Influence of the intensity of the conditioning regimen on the characteristics of acute and chronic graft-versus-host disease after allogeneic transplantation. *Br J Haematol* 130(3):394-03.[PubMed]
- Ferrara JLM, Cooke KR, Teshima T (2003) The pathophysiology of acute graft-versus-host disease. *Int J Hematol* 78(3):181-187.[PubMed]
- Sadeghi B, Aghdami N, Hassan Z, Forouzanfar M, Rozell B, et al. (2005) GVHD after chemotherapy conditioning in allogeneic transplanted mice. *ukocyte migration and Graft-versus-host disease. Blood* 11: 4191-4199.
- Perreault C, Décarý F, Brochu S, Gyger M, Bélanger R, et al. (1990) Minor histocompatibility antigens. *Blood* 76(7): 1269-1281.[PubMed]
- Goulmy E (1996) Human minor histocompatibility antigens. *Curropin Immunol* 8(1): 75-82.[PubMed]
- Simpson E, Scott D, James E, Lombardi G, Cwynarski K, et al. (2002) Minor H antigens: genes and peptides. *Transpl Immunol* 10(2-3): 115-123.[PubMed]
- Teshima T, Ferrara JL (2002) Understanding the alloresponse: new approaches to graft-versus-host disease prevention. *Semin Hematol* 39(1):15-22.[PubMed]
- Shlomchik WD (2007) Graft-versus-host disease. *Nat Rev Immunol* 7(5): 340-352.[PubMed]
- Matte CC, Liu J, Cormier J, Anderson BE, Athanasiadis I, et al. (2004) Donor APCs are required for maximal GVHD but not for GVL. *Nat Med* 10(9): 987-992.[PubMed]
- Anderson BE, McNiff JM, Jain D, Blazar BR, Shlomchik WD, et al. (2005) Distinct roles for donor- and host-derived antigen-presenting cells and co-stimulatory molecules in murine chronic graft-versus-host disease: requirements depend on target organ. *Blood* 105(5): 2227-2234.[PubMed]
- Kataoka Y, Iwasaki T, Kuroiwa T, Seto Y, Iwata N, et al. (2001) The role of donor T cells for target organ injuries in acute and chronic graft-versus-host disease. *Immunology* 103(3): 310-318.[PubMed]
- Friedman TM, Jones SC, Statton D, Murphy GF, Korngold R (2004) Evolution of responding CD4+ and CD8+ T-cell repertoires during the development of graft-versus-host disease directed to minor histocompatibility antigens. *Biol Blood Marrow Transplant* 10(4): 224-235.[PubMed]
- Panoskaltis-Mortari A, Hermanson JR, Taras E, Wangenstein OD, Serody JS, et al. (2003) Acceleration of idiopathic pneumonia syndrome (IPS) in the absence of donor MIP-1 alpha (CCL3) after allogeneic BMT in mice. *Blood* 101(9): 3714-3749.[PubMed]
- Wen HS, Wang JM, Zhou H, Xia R, Qiu HY, et al. (2006) Migration and Distribution of Allogeneic T Lymphocytes in Organs of Graft-Versus-Host Disease Mouse Model. *J Exp Hematol* 14(5): 919-923.[PubMed]
- Panoskaltis-Mortari A, Price A, Hermanson JR, Taras E, Lees C, et al. (2004) In vivo imaging of graft-versus-host-disease in mice. *Blood* 103(9): 3590-3598. [PubMed]
- Trisolini R, Stanzani M, Lazzari Agli L, Colangelo A, Bonifazi F, et al. (2001) Delayed non-infectious lung disease in allogeneic bone marrow transplant recipients. *Sarcoidosis Vasc Diffuse Lung Dis* 18(1): 75-84.[PubMed]
- Padovan CS, Gerbitz A, Sostak P, Holler E, Ferrara JL, et al. (2001) Cerebral involvement in graft-versus-host disease after murine bone marrow transplantation. *Neurology* 56(8): 1106-1108.[PubMed]
- Ma M, Barnes G, Pulliam J, Jezek D, Baumann RJ, et al. (2002) CNS angitis in graft vs host disease. *Neurology* 59(12): 1994-1997.[PubMed]
- Ichiba T, Teshima T, Kuick R, Misesk DE, Liu C, et al. (2003) Early changes in gene expression profiles of hepatic GVHD uncovered by oligonucleotide microarrays. *Blood* 102(2): 763-771.[PubMed]
- Sugerman PB, Faber SB, Willis LM, Petrovic A, Murphy GF, et al. (2004) Kinetics of gene expression in murine cutaneous graft-versus-host disease. *Am J Pathol* 164(6): 2189-2202.[PubMed]
- Matloubian M, Lo CG, Cinamon G, Lesneski MJ, Xu Y, Brinkmann V, et al. (2004) Lymphocyte egress from thymus and peripheral lymphoid organs is dependent on S1P receptor1. *Nature* 427(6972): 355-360.[PubMed]
- Kim YM, Sachs T, Asavaroengchai W, Bronson R, Sykes M (2003) GVHD can be separated from graft-versus-lymphoma effects by control of lymphocyte trafficking with FTY720. *J Clin Invest* 111: 659-669.[PubMed]
- Laurence A Turka, Wayne W. Hancock (2003) Transplantation immunobiology. Blackwell Pub.
- Via CS, Finkelman FD (1993) Critical role of interleukin-2 in the development of acute graft-versus-host disease. *Int Immunol* 5(6): 565-572.[PubMed]
- Ferrara JL, Abhyankar S, Gilliland DG (1993) Cytokine storm of graft-versus-host disease: a critical effector role for interleukin-1. *Transplant Proc* 25(1 pt 2): 1216-1217.[PubMed]
- Garlisi CG, Pennline KJ, Smith SR, Siegel MI, Umland SP (1993) Cytokine gene expression in mice undergoing chronic graft-versus-host disease. *Mol Immunol* 30(7): 669-677.[PubMed]
- Wen HS, Wang JM, Zhou H, Gong SI, Gao L, et al. (2013) Migration and activation of T cells during development of graft-versus-host disease in a mouse model. *Transplant Proc* 45(2):713-718.[PubMed]