



The Molecular Docking Study of Interaction of Newly Synthesised Benzamide Appended by Pyrazolone Derivatives as Ligand Molecule with the Target Protein 6LU7 of Novel Corona Virus

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ABSTRACT

Plants and bioactive compounds have played an important role in the development of several clinically useful therapeutic agents since time immemorial. The global health emergency of novel COVID-19 is due to severe acute respiratory syndrome corona virus-2 (SARS-CoV-2). As if now there are no approved drugs for the treatment of corona viral disease (COVID-19), although some of the drugs have been tried. The virtual interaction of the COVID-19 main protease in complex with the inhibitor N³ (Research Collaborators for Structural Bioinformatics Protein Data Bank (PDB) ID: 6LU7), Hence this is chosen as target protein molecule. The newly synthesised compounds of benzamide appended pyrazolones derivatives are made as ligand molecules. The synthesis of these organic ligand molecules is done through four steps using different reagents and different environmental conditions. In this article we have discussed the interaction of our synthesized ligands with the target protein molecule using autodock tools. Firstly, the ligands were prepared using commercial ACD/chemsketch tool in PDB format. Desired protein target 6LU7 is downloaded from Protein Data Bank. Further protein optimization done by removing co-ordinates and hetero atoms, energy minimization of protein is done by Swiss PDB viewer 4.1.0. To change the file format open babel 2.4.1 is used. Docking of protein and ligand is done using autodock 4.2 and the results are visualized by pymol and tabulated using the Lamarckian genetic algorithm. After docking comparative study of the binding energies, inhibitory constant and hydrogen bonding of all interactions were discussed and tabulated for the good results.

Keywords: Benzamide; Pyrazolones; Protein-ligand interaction; Molecular docking; Autodock 4.2; Pymol

INTRODUCTION

The molecular docking approaches are used to model the interaction between a small molecule and a protein at the atomic level, which allow us to characterize the behavior of small molecules in the binding site of target proteins as well as to elucidate fundamental biochemical processes and study of its features. The docking process involves two basic steps they are prediction of the ligand conformation as well as its position and orientation within these sites (usually referred to as pose) and also assessment of the binding affinity [1].

Pyrazolones are proven to be valuable over wide range of applications in the pharmaceutical industries. The most classic approach to prepare pyrazolones is *via* the reaction between α , β -ketoester, α , β -cyanoester or α , β -unsaturated esters and hydrazine derivatives of alkanes, arenes and heterocycles. Synthesis of benzamide appended

heterocycles was initiated, to expand the application of synthesized compounds as the useful drugs [2].

Plan for synthesis of ligands

Synthesis of substituted benzamides 3a-f was achieved by reacting aminophenol and benzoyl chloride in presence of trimethylamine at room temperature for 24 hrs, further synthesis of compounds 4a-f is accomplished by reaction of substituted benzamides 3a-f with ethylbromoacetate in presence of potassium carbonate (Scheme 1). The obtained acetates 4a-f were reacted with 80% hydrazine hydrate to yield substituted hydrazides 5a-f. These phenyl hydrazides 5a-f were used further for cyclisation with ethylacetoacetate in presence of 20% tetrabutylammonium hydroxide in methanol with ethylene glycol as solvent to achieve substituted N-(4-(2-(5-methyl-3-oxo-2,3-dihydropyrazolyl-1)-2-oxoethoxy)phenyl) benzamide derivatives

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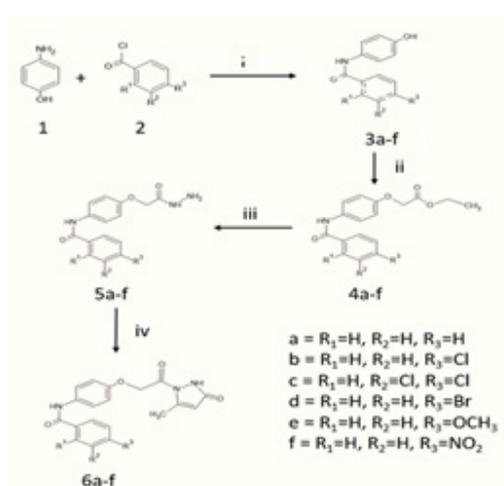
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6a-f which is our ligand molecule (Scheme 2). The structure of synthesized ligands were confirmed by characterization using spectroscopic techniques [3,4].

Chemicals and solvents used were purchased from Sigma Aldrich and used without further purification unless stated. Melting points were determined in open capillaries on a Buchi oil melting point apparatus and are uncorrected Equation(c). Reactions were monitored by using Thin Layer Chromatography (TLC) on aluminum sheet precoated with silica gel 60 F254 (0.2 mm Merck). Chromatographic spots were visualized by UV light and/or with iodine. For column chromatography, silica gel of 100-200 mesh size was used. ¹H NMR spectra were acquired on a Bruker NRC-IISC instrument, Bangalore at ambient temperature at 400 MHz and/or 300 MHz in DMSO-d₆ or CDCl₃ and TMS was used as an internal reference. ¹³C NMR spectra were recorded on a Bruker AMX-400(100 MHz) with complete proton decoupling. Chemical shift are reported in ppm from TetramethylSilane (TMS) with the solvent as the internal reference (CDCl₃: δ 77.0 ppm or DMSO δ 40.0 ppm). LC-MS was performed on Agilent LCMS system equipped with a BEH C8, 30 \times 4.6 mm, 1.7 μ m column at a flow rate of 1.0 ml/min [5].

Scheme 1: Synthesis of ligand molecules



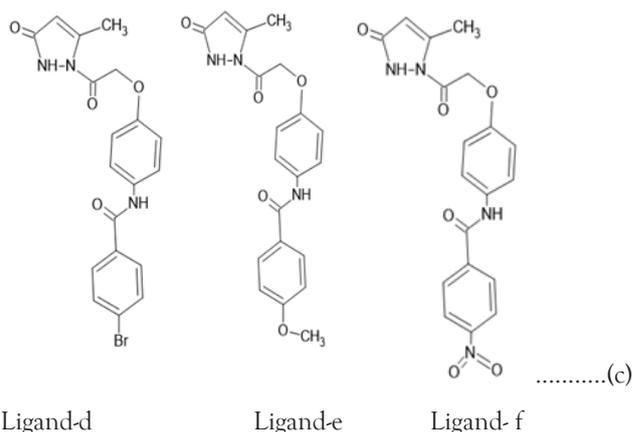
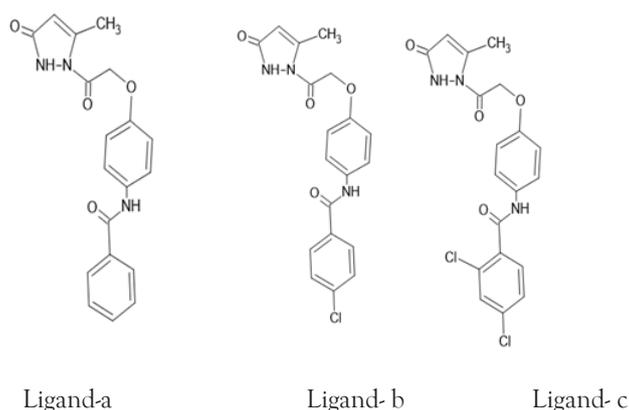
Reagents: i=Dry THF, RT, TEA

ii=Ethylbromoacetate, K₂CO₃, MeOH

iii=80% Hydrazine hydrate, Methanol

iv=Ethylacetoacetate, TBAH in Methanol, Ethylene glycol, RT

Scheme 2: Structure of benzamide appended by pyrazolones acting as ligands



Molecular docking

Molecular docking is a well-established computational technique which predicts the interaction energy between two molecules. This technique basically incorporates algorithms like molecular dynamics, fragment search methods and so on. Molecular docking also gives the clear idea that what biochemical mechanism is going on between protein and the ligand. In this article we used to study the interaction of two molecules to fit the best orientation of ligand which would form a complex with overall minimum binding energy [6]. We start with the *in-silico* generation of ligand and conversion of file format. Further selection of protein and preparation of protein by optimization of protein and energy minimization. Next using Autodock Vina 4.2 autodock was run to check the interaction and finally analysed by visualization tools and creation of algorithm table [7].

Using commercial ACD/chemsketch tool we have drawn the ligand structure and saved in MDL mol.files and further this file is converted into PDB file format by using open babel 2.4.1 which is a necessary format to run autodock. The selected protein is downloaded from RCSB Protein Data Bank in PDB format. In this article we have selected COVID-19 main protease in complex with the inhibitor N³ (Research Collaborators for Structural Bioinformatics Protein Data Bank (PDB) ID: 6LU7. Next the protein optimization and energy minimization is done using swiss PDB viewer 4.1.0. by deleting all hetero atoms and co-ordinates.

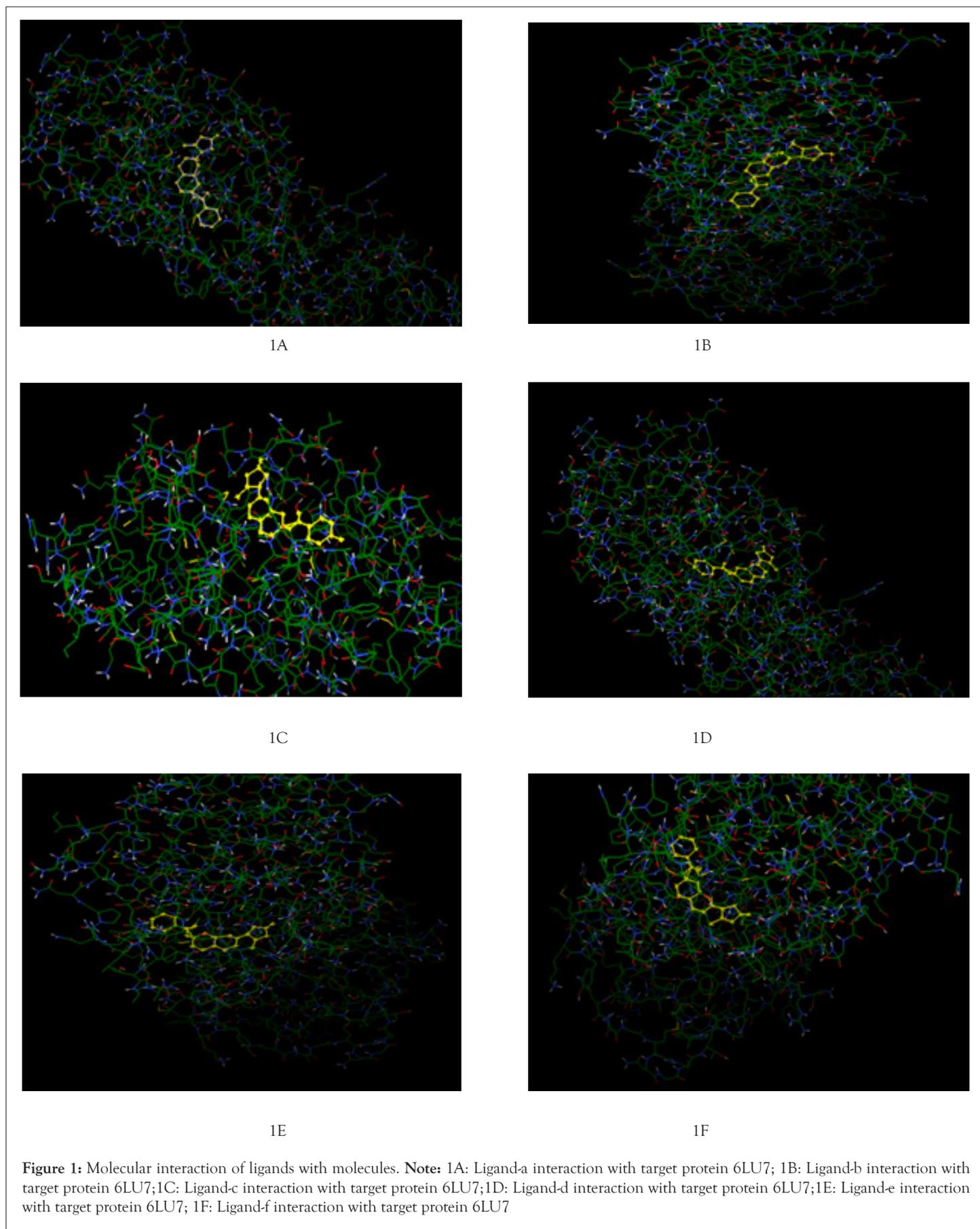
Both ligand and protein molecules are prepared in PDB format which is necessary to run autodock. During docking all water molecules are removed from the protein and polar hydrogen atoms and kollman charges were added to the protein and assigned with AD4 types of atoms. Grid selected around 0.512 value was fixed for every run around the active site of protein that is GLN189 which is obtained by literature survey. The docked files are saved in DLG format by Lamarckian Genetic Algorithm and further it is used for visualization and tabulation of results [8-12].

The results of molecular docking and the poses of ligand- protein interaction were studied using autodock tools for all interactions such as electrostatic forces, hydrogen bonding, and torsional effects and so on.

Further visualization of the poses of ligand-protein interaction by Pymol. Especially to study the hydrogen bonding interaction of ligand and protein in the selected grid region. The interacted amino acids were tabulated and studied for further. More the hydrogen bonds more will be the stability of the complex thus all six ligands were studied for their hydrogen bonding interaction

with the same protein 6LU7. The selection of grid box size is same for all type of ligands used in the molecular docking studies. Totally ten runs were considered for the best pose of ligand-protein interaction among them the lowest binding energy is considered as

the best pose. Further the selected interaction pose of ligand and protein were studied under pymol to get the hydrogen bonding interactions (Figures 1A-1F and Figures 2A-2F).



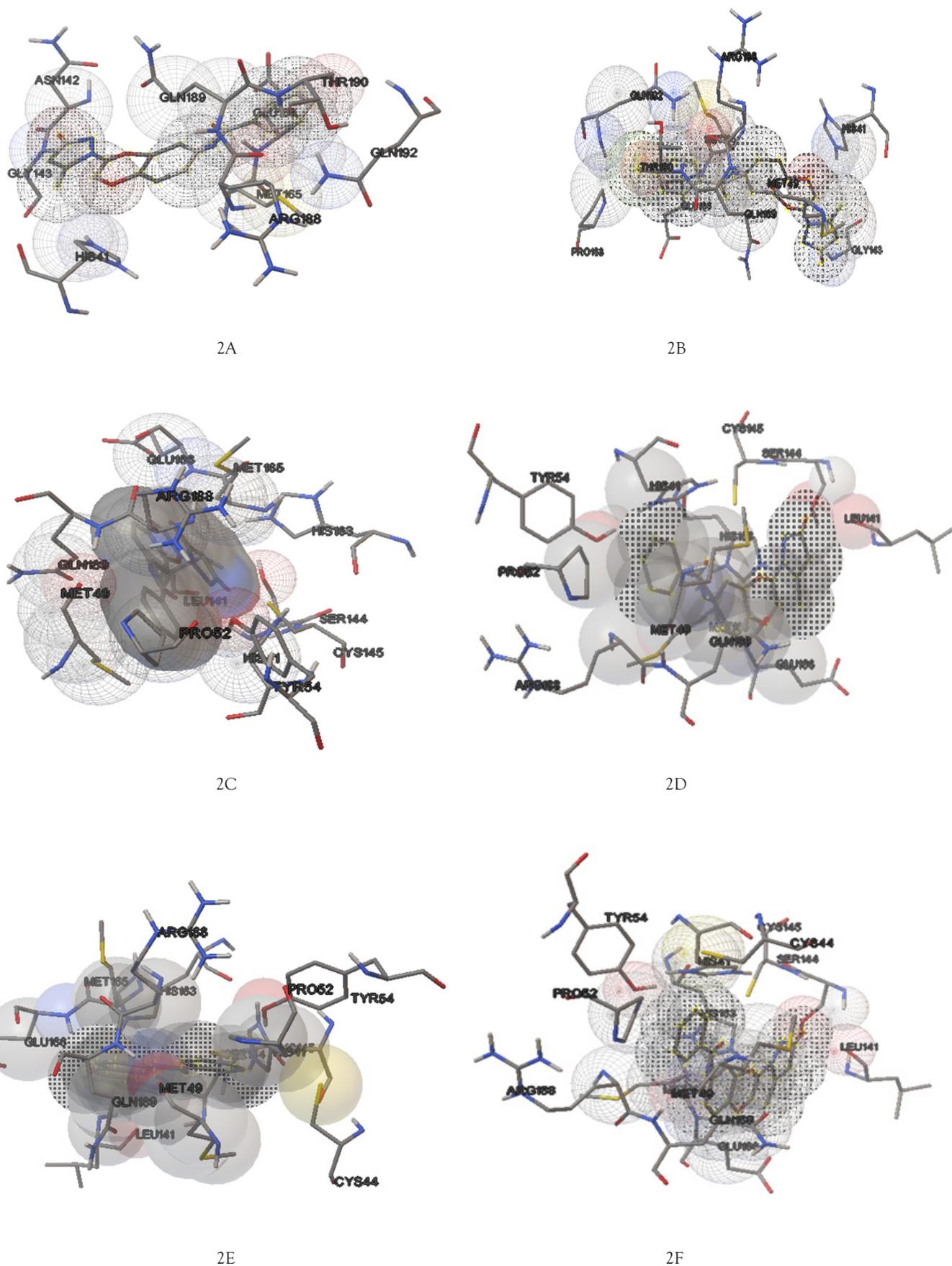


Figure 2: Visualization of protein-ligand interaction in terms of hydrogen bonding. Note: 2A: Visualization of ligand-a interaction with 6LU7; 2B: Visualization of ligand-b interaction with 6LU7; 2C: Visualization of ligand-c interaction with 6LU7; 2D: Visualization of ligand-d interaction with 6LU7; 2E: Visualization of ligand-e interaction with 6LU7; 2F: Visualization of ligand-f interaction with 6LU7

RESULTS AND DISCUSSION

Interaction of ligand-a (N-{4-[2-(5-methyl-3-oxo-2,3-dihydro-1H-pyrazol-1-yl)-2-oxoethoxy]phenyl}benzamide)

After docking the protein target with each of our ligands, we have observed several information regarding the interactions. Each ligand has slight difference in their interaction type with the selected target protein 6LU7 molecule. According to it for ligand-a and protein 6LU7 interaction we found the least binding energy as -8.90 in the 8th run with the inhibition constant of 301.49 nM and the entropy of the selected grid was found to be 0.41. When the complex in viewed in pymol we found that Ligand-a is having three hydrogen bonds with the target protein molecule 6LU7 with the selected grid box. The bond length of the three hydrogen bonds are 2.0, 2.4 and 2.5 with the amino acids GLU166, SER144 and LEU141 respectively [13].

Interaction of ligand-b (4-chloro-N-{4-[2-(5-methyl-3-oxo-2,3-dihydro-1H-pyrazol-1-yl)-2-oxoethoxy]phenyl}benzamide)

Ligand-b and protein 6LU7 interaction we found the least binding energy as -8.77 in the 7th run with the inhibition constant of 374.22 nM and the entropy of the selected grid was found to be 0.11. When the complex in viewed in pymol we found that Ligand-a is having one hydrogen bonds with the target protein molecule 6LU7 with the selected grid box. The bond length of that one hydrogen bonds is 2.0 with the amino acids GLY143 [14].

Interaction of ligand-c (2,4-dichloro-N-{4-[2-(5-methyl-3-oxo-2,3-dihydro-1H-pyrazol-1-yl)-2-oxoethoxy]phenyl}benzamide)

Ligand-c and protein 6LU7 interaction we found the least binding energy as -8.99 in the 3rd run with the inhibition constant of 255.23 nM and the entropy of the selected grid was found to be 0.14. When the complex in viewed in pymol we found that Ligand-a is having one hydrogen bonds with the target protein molecule 6LU7

with the selected grid box. The bond length of that one hydrogen bonds is 2.9 with the amino acids ARG188 [15].

Interaction of ligand-d (4-bromo-N-{4-[2-(5-methyl-3-oxo-2,3-dihydro-1H-pyrazol-1-yl)-2-oxoethoxy]phenyl}benzamide)

Ligand-d and protein 6LU7 interaction we found the least binding energy as -8.85 in the 7th run with the inhibition constant of 325.66 nM and the entropy of the selected grid was found to be 0.14. When the complex in viewed in pymol we found that Ligand-a is having one hydrogen bonds with the target protein molecule 6LU7 with the selected grid box. The bond length of that one hydrogen bonds is 3.2 with the amino acids GLU166 [16-18].

Interaction of ligand-e (4-methoxy-N-{4-[2-(5-methyl-3-oxo-2,3-dihydro-1H-pyrazol-1-yl)-2-oxoethoxy]phenyl}benzamide)

Ligand-e and protein 6LU7 interaction we found the least binding energy as -8.70 in the 3rd run with the inhibition constant of 420.33 nM and the entropy of the selected grid was found to be 0.10. When the complex in viewed in pymol we found that Ligand-a is having two hydrogen bonds with the target protein molecule 6LU7 with the selected grid box. The bond length of that two hydrogen bonds is 2.3 and 4.8 with the amino acids GLY143 and GLN189 respectively [19,20].

Interaction of ligand-f (N-{4-[2-(5-methyl-3-oxo-2,3-dihydro-1H-pyrazol-1-yl)-2-oxoethoxy]phenyl}-4-nitrobenzamide)

Ligand-f and protein 6LU7 interaction we found the least binding energy as -9.17 in the 8th run with the inhibition constant of 190.41 nM and the entropy of the selected grid was found to be 0.15. When the complex in viewed in pymol we found that Ligand-a is having three hydrogen bonds with the target protein molecule 6LU7 with the selected grid box. The bond length of that three hydrogen bonds is 2.1, 2.3 and 2.1 with the amino acids GLN192, GLN192 and GLY143 respectively (Figure 3A-3F) (Table 1).

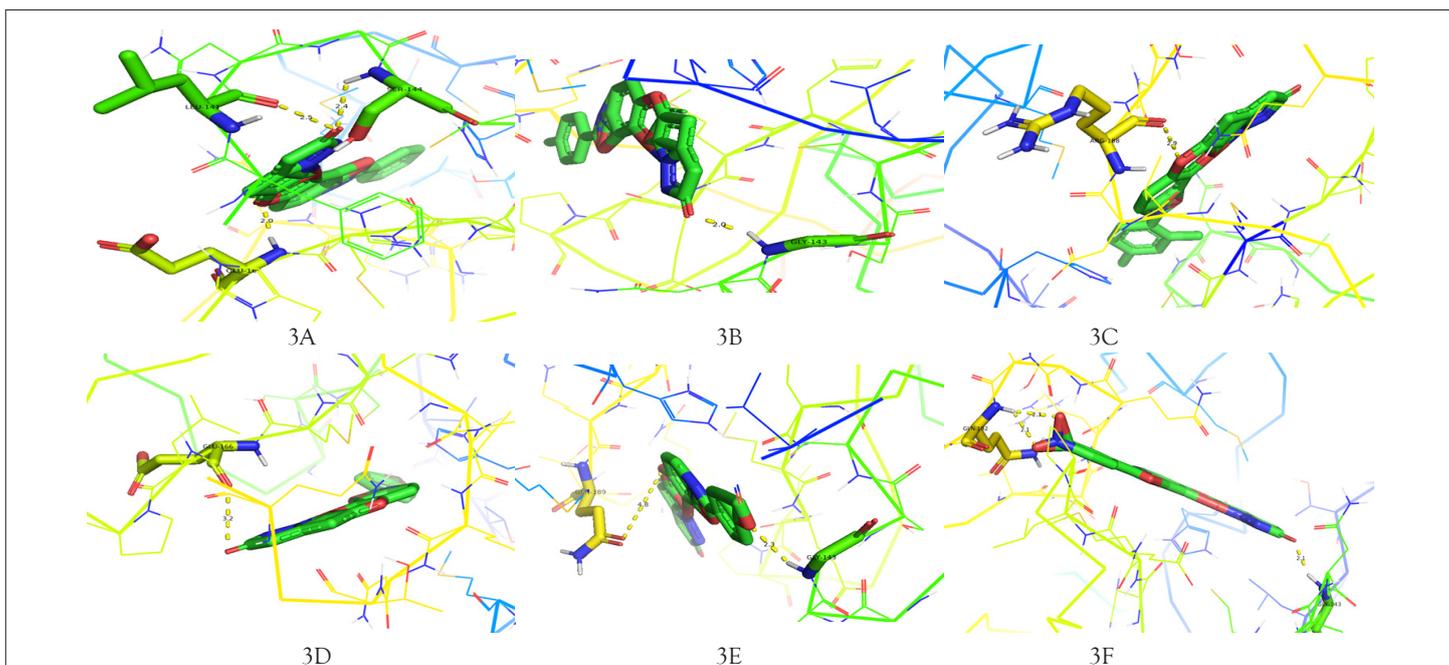


Figure 3: Pymol viewer of hydrogen bondings in the protein-ligand complex. **Note:** 3A: Ligand-a interaction with 6LU7 protein through hydrogen bonding; 3B: Ligand-b interaction with 6LU7 protein through hydrogen bonding; 3C: Ligand-c interaction with 6LU7 protein through hydrogen bonding; 3D: Ligand-d interaction with 6LU7 protein through hydrogen bonding; 3E: Ligand-e interaction with 6LU7 protein through hydrogen bonding; 3F: Ligand-f interaction with 6LU7 protein through hydrogen bonding

Table 1: Comparative study of interaction of ligand molecule with target protein 6LU7.

Name of ligand	Best Run	Binding energy	Inhibition constant ki	Entropy of cluster	Number of hydrogen bonds	Bond length	Interacted amino acid
Ligand-a	8	-8.9	301.49 nM	0.41	3	2	GLU166
						2.4	SER144
						2.5	LEU141
Ligand-b	7	-8.77	374.22 nM	0.11	1	2	GLY143
Ligand-c	3	-8.99	255.23 nM	0.14	1	2.9	ARG188
Ligand-d	7	-8.85	325.66 nM	0.14	1	3.2	GLU166
Ligand-e	3	-8.7	420.33 nM	0.1	2	2.3	GLY143
						4.8	GLN189
						2.1	GLN192
Ligand-f	8	-9.17	190.41 nM	0.15	3	2.3	GLN192
						2.1	GLY143

CONCLUSION

More the number of hydrogen bond in the complex more will be the stability and lesser the value of inhibition constant (Ki) more will be the interaction power of the ligand with protein which means a small concentration of ligand can show a good interaction with the protein molecule. When compare the interaction results of all the ligands with same selected protein target molecule, ligand-f is having a good interaction with protein when compare to all ligands because ligand-f is having binding energy as -9.17 in the 8th run with the inhibition constant of 190.41 nM.

DECLARATION

Ethical approval

The data and visualization images used in this research article are not plagiarized by anywhere.

Competing interests

Not applicable

Author's contribution

The whole article is written and data are obtained by author Smt. Pavithra V and the guidance for the research was provided by the corresponding author Dr. Sudha B S.

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Availability of data and materials

The data base and software used for this research article are reproducible and available for universal usage.

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