

**Open Access** 

# The Emergence of Bioenergetics: The Formation of a Gluconeogenesis System and Reductive Pentose Phosphate Pathway of CO<sub>2</sub> Fixation in Ancient Hydrothermal Systems

## Sergey A. Marakushev<sup>\*</sup> and Ol'ga V. Belonogova

Institute of Problems of Chemical Physics, Russian Academy of Sciences, Chernogolovka, Moscow Region, Russia \*Corresponding author: Marakushev SA, Institute of Problems of Chemical Physics, Russian Academy of Sciences, Chernogolovka, Russia, Tel: 496-522-7772; E-mail: marak@cat.icp.ac.ru

Rec date: Apr 18, 2016; Acc date: Nov 08, 2016; Pub date: Nov 11, 2016

**Copyright:** © 2016 Marakushev SA et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## Abstract

The origin of phosphorus metabolism is one of the central problems in the context of the emergence of life on Earth. It has been shown that the C–H–O system can be transformed into a four- component C–H–O–P system with the formation of a gluconeogenesis path in a possible Archean hydrothermal condition under the influence of a phosphorus chemical potential. This system became the energy supply basis for protometabolism, and facilitated the formation of a new  $CO_2$  fixation cycle (the reductive pentose phosphate pathway).

The modular design of central metabolism in the C–H–O–P system is derived from the parageneses (associations) of certain substances, and the emerging modules in turn associate with each other in certain physical and chemical hydrothermal conditions. The assembly of malate, oxaloacetate, pyruvate, and phosphoenolpyruvate is a reversible "turnstile - like" mechanism with a switching of reaction direction that determines the trend of specific metabolic systems development.

**Keywords:** Origin of metabolism energetics; Autocatalytic cycles; Gluconeogenesis; CO<sub>2</sub> fixation; Modularity; Parageneses; Chemical potentials

## Introduction

One of the most important concepts in the theories of the chemolithoautotrophic origin of life in the Archean volcanic hydrothermal environment is autocatalytic CO<sub>2</sub> fixation [1-9]. It has also been suggested that the last common ancestor (LCA) of all extant cell lineages was a chemolithoautotrophic thermophilic anaerobe [10-14] capable of synthesizing organic 'building blocks' from the inorganic carbon. Thus, these microorganisms can serve as a model for studying primordial metabolism. However, CO<sub>2</sub> fixation requires energy, which was provided by energy from high-energy intermediates, organic phosphates in particular, that are presently involved in the primary biochemical pathways.

In previous works we have considered the thermodynamic factors of natural selection in the coupled autocatalytic cycles of CO<sub>2</sub> fixation (redundant modular constructions) in the three - component C-H-O system [6,15]. It is rationalized that these systems are the primordial metabolic autocatalytic reductive citrate (RC) cycle (reductive tricarboxylic acid, Arnon-Buchanan cycles) and 3-hydroxipropionate (3-HP) cycle. An important factor in the stability and subsequent evolution of these coupled cycles is the reversibility of some key chemical reactions such as succinate  $\leftrightarrow$  fumarate, malate  $\leftrightarrow$  fumarate and others [16]. The further development of this system with negative feedback involves increasing the number of independent components by maintaining additional supply of phosphorus, nitrogen, and sulfur. Adding phosphorus (forming a four - component C-H-O-P system) leads to the development of the gluconeogenesis system, which

produces phosphorylated sugars, a basic source of energy for different biosynthetic pathways. The nodal intermediates and proposed autocatalysts in this process are malate, pyruvate, and oxaloacetate, the transformation of which into phosphoenolpyruvate permits the formation of phosphorylated trioses, pentoses, and hexoses.

The fundamental role of phosphorylated carbohydrates and the universality of the few modern metabolic interconversions suggest their origin at the earliest evolutionary stage. The possibility of gluconeogenesis first originating as a precursor to phosphorus metabolism in hyperthermophilic microorganisms was considered in [17]. Gluconeogenesis starting with phosphoenolpyruvate uses the unusual ancient enzyme fructose 1,6-bisphosphate aldolase/ phosphatase, which converts triose phosphates to fructose 1,6bisphosphate [18,13]. The ancient origin of gluconeogenesis with the consequent origin of the ancestral Aquificae, Chloroflexi, and Thermotogae taxa is postulated. Reactions converting triose phosphate into fructose 1,6-bisphosphate are generally reversible, but the reaction of fructose 1,6-bisphosphate into stable fructose 6-phosphate is irreversible and substantially determines the unidirectionality of gluconeogenesis, which precedes the emergence of glycolysis evolutionarily [17].

The early origin of gluconeogenesis is indirectly supported by data from the non-enzymatic metal - catalyzed formation of phosphorylated three-carbon sugars and pyruvate [19,20], and the widespread role of non-enzymatic catalysis in phosphate metabolism is considered in ref. [21]. It should be noted that both non-enzymatic and enzymatic reactions are based on the same fundamental thermodynamic laws. Experimental analysis of the conditions required for phosphorylated intermediate formation and stability in the pentose phosphate pathway is given in ref. [22].



**Figure 1:** Inferred coupling of the autocatalytic loops of the archaic reductive citrate (RC) and 3-hydroxipropionate (3-HP) cycles with the further development of phosphorus metabolism in the archaic systems of gluconeogenesis and the reductive pentose phosphate (RPP) pathway of  $CO_2$  fixation. The arrows show the direction of reactions. The circular arrows show the direction of reactions in the coupled RC and 3-HP cycles. In the RPP cycle, three molecules of fructose-6-phosphate regenerate three molecules of ribulose-1,5-bisphosphate and one molecule of 3-phosphoglycerate.

The reactions underlying the interconversion of phosphorylated carbohydrates were examined under the conditions found in a putative Archean ocean, which were determined based on the chemical composition of sedimentary rocks in this time period. It was shown that the simple inorganic ions (Fe (II), Co (II), Ni (II), Mo (IV)) found in rocks from the Archean period could catalyze the reactions observed in the extant metabolic pathways. It was also concluded that iron (at concentrations of 20  $\mu$ M to 5 mM) was the most effective catalyst of metabolic reactions requiring substrate phosphorylation in early anoxygenic Archean ocean. It is assumed that the ancient ocean was enriched not only by ferro - ions but also phosphates [22-24] that emanated from high-temperature alkaline hydrothermal fluids to the surface of the Earth [25]. Nevertheless, it requires some thermodynamic analysis of the possibility of phosphorylated carbohydrates formation.

A simplified biomimetic image of primordial anaerobic central carbon metabolism in the form of branching metabolic reactions with the formation of gluconeogenesis and the reductive pentose phosphate (RPP)  $CO_2$  fixation cycle (Calvin-Benson cycle) is presented in Figure

1, which shows the development of coupled autocatalytic systems (archaic RC and 3-HP cycles [26]) in the direction of phosphorus metabolites - triose phosphates and phosphorylate d sugars. It is assumed that these metabolic changes took place during of the orgin of life energetics aspect.

# **Methodical Approach**

The physicochemical analysis of parageneses (associations, assemblages) is widely used in geochemical researches of mineral systems [27]. This approach is based on a generalization of the thermodynamic and physical properties of minerals in order to detect the conditions responsible for the formation of the parageneses observed in rocks and ores. In this case, the thermodynamic potential method [27,28], which allows one to develop a system of geochemical mineral facies (thermodynamic stability areas), is used. Our preliminary calculations indicated that paragenetic analysis can be extended to organic substances, which form the systems listed below (the systems are listed in increasing complexity of chemical composition and structure): C–H–O, C–H–O–N, C–H–O–N–P and C–H–O–N–P-S [6,15,29].

The studies of organic substances assemblages (parageneses) and areas of their thermodynamic stability (facies) is based on the Gibbs phase rule, according to which the number of degrees of freedom for a thermodynamic system in equilibrium is equal to the number of system independent components plus two minus the number of phases. In this case, the number of independent components is the smallest number of chemical components; the compositions of all possible phases of the system can be obtained by their combination. The phase rule is not limited to consideration of the extensive parameters of the system and, hence, is completely applicable to open systems with chemical potential as an independent parameter [16,30]. Determining the total number of system independent parameters (intensive and extensive), and using this number together with the Gibbs phase rule provides a way to study the thermodynamic properties of organic systems.

The system state was calculated using free partial energy values for organic substance formation ( $\Delta$ Go f,T), depending on external conditions. The state diagrams, which reflect the facies of organic compounds and their parageneses, are graphically presented. When considering these organic compounds in fluid and aqueous hydrothermal systems, the points of major importance are shown with diagrams: a) chemical composition - paragenesis (at constant pressure (P), temperature (T), and chemical potential (µi)), b) chemical potential - temperature (at constant P), c) the relationship between chemical potentials of the components:  $\mu$ H<sub>2</sub>O -  $\mu$ CO<sub>2</sub>,  $\mu$ H<sub>2</sub> -  $\mu$ O<sub>2</sub>, etc. (at constant P and T).

The major equilibrium factors in the physicochemical analysis of parageneses are the chemical potential ( $\mu i$ ) of each component (representing its partial energy). The value  $\mu i$  is expressed through activity, *ai*, and fugacity, *fi*, as follows: ( $\mu i = (\mu^{\circ}i)T, p + RT \ln ai = (\mu^{\circ}i)T, p + RT \ln fi$ .)

The aqueous constants used were derived using the electrostatic model method, according to which the solvate constituent of a substance plays an essential role in the chemical potential at different temperatures, reflecting the contribution from electrostatic interactions between the substance and solvent ( $H_2O$ ) [31-34]. Thermodynamic calculations of the Gibbs standard free energy and analysis of the geochemical constraints showed that the abiotic

#### Page 3 of 6

synthesis of organic compounds in hydrothermal systems is limited by the metastable equilibrium that results from kinetic barriers, which prevent the achievement of stable equilibrium [32,35-37]. Most of the organic substances in condensed and dissolved phases are in a metastable state, i.e., these substances do not reach the minimum Gibbs free energy for the given composition of elements, and thus are "kinetic" or "metastable" phases [38,39].

Previously, we calculated a diagram of composition - paragenesis for the ternary C-H-O system [e.g., 6]. The addition of phosphorus to the C-H-O system forms the quaternary system C-H-O-P, in which the independent components – carbon, hydrogen, oxygen, and phosphorus – are extensive parameters (*fex*). Figure 2 presents the phase diagram of compositions for the compounds shown in Figure 1. In this ternary C-H-O diagram, the phases of the phosphorylated compounds are represented by the subtracting of orthophosphoric acid (H<sub>3</sub>PO<sub>4</sub>). If, in thermodynamic calculations, the chemical potential is determined for  $H_3PO_4$  ( $\mu H_3PO_4$ ), and phosphorus becomes an intensive parameter (*fin*), then the quaternary system becomes ternary (C–H–O). If the chemical potential is calculated for hydrogen or methane ( $\mu$ CH<sub>4</sub>,  $\mu$ H<sub>2</sub>), and hydrogen becomes intense parameter (*fin*), then the ternary system becomes a binary system, C–O (Figure 2a and b). The dashed conodes connect CH<sub>4</sub> and H<sub>2</sub> with substances phases, which are represented by stars on the triangle side. The composition diagram (Figure 2c) shows that, if the chemical potential is calculated for CO<sub>2</sub> ( $\mu$ CO<sub>2</sub>), and CO<sub>2</sub> becomes an intensive parameter (*fin*), then the ternary system becomes a binary system, C–H (dashed conodes connect the CO<sub>2</sub> point, and substance phases represented by rhombs on the triangle base). The free energies of aqueous ionized phosphorus substance formation were taken from ref. [40] and were designed for non-ionized forms using the method described in refs., [40,41]. The constants for substances in the C–H–O system were taken from [16].



**Figure 2:** C–H–O phase diagram representing the compositions of the compounds shown in the modular scheme (Figure 1). The twocomponent C–O system is generated when the chemical potentials of  $CH_4$  and  $H_2$  (**a** and **b**) are used. The two-component C–H system (**c**) is generated when the chemical potential of  $CO_2$  is used. Phosphorylated compounds are represented by the subtracting of orthophosphoric acid ( $H_3PO_4$ ) composition. Substances in the C–H–O system are indicated with filled triangles and substances in the C–H–O–P system are indicated with empty boxes. Designations of the substances: 1 - fumarate (Fum), 2- succinate (Suc), 3 - acetate (Acet), 4 - pyruvate (Pyr), 5 malate (Mal), 6 - glyoxylate (Glx), 7 - oxaloacetate (Oxal), 8 - phosphoenolpyruvate (PEP), 9 - 3-phosphoglycerate (PG), 10 – glyceraldehyde -3- phosphate (GAP), 11 - fructose 6-phosphate, 12 - ribulose 1,5 bisphosphate, 13 - fructose 1,6-bisphosphate (FBP).

Certainly, the hydrothermal systems are generally characterized by a more high- pressure and high-temperature conditions. However, these conditions cannot fundamentally change the character the diagrams of chemical potentials - as a rule, the equilibria shifts in a direction of its higher values.

It is assumed that the anoxygenic Archean ocean saturated with siliceous compounds and Fe (II) contained a higher level of dissolved orthophosphate than the modern ocean [23,42,43]. These conditions have primarily been determined with the use of the orthophosphate chemical potential in thermodynamic calculations of the origin and the evolution of protometabolic pathways related to the four component C-H-O-P system.

# The Chemical Potentials of Methane, Molecular Hydrogen, and Carbon Dioxide

The most important energy factors in the generation of organic compounds, which are intermediates of protometabolic pathways in

hydrothermal systems, are the chemical potentials of molecular hydrogen and hydrocarbons. The predominant widespread hydrocarbon is methane, the concentration of which (e.g., in volcanic oceanic emissions) is usually more than two orders of magnitude higher than the concentration of other hydrocarbons [44].

At the present time, the abiotic synthesis of organic compounds in the hydrothermal systems of mid-ocean ridges has been confirmed in a number of publications (e.g., [44-46]). Alkanes and carboxylic acids represent some of the most abundant organic structure types found in natural hydrothermal systems and sedimentary basin fluids.

The diagram in Figure 3 is based on aqueous constants at standard conditions and shows the formation and development of coupled C–H–O–P metabolic systems, Figure 1, under the influence of the chemical potentials of methane ( $\mu$ CH<sub>4</sub>) and phosphoric acid ( $\mu$ H<sub>3</sub>PO<sub>4</sub>).



**Figure 3:** Diagram of the chemical potentials of CH<sub>4</sub> and H<sub>3</sub>PO<sub>4</sub> ( $\mu$ CH<sub>4</sub>=RTlnαCH<sub>4</sub>,  $\mu$ H<sub>3</sub>PO<sub>4</sub>=RTlnαH<sub>3</sub>PO<sub>4</sub>, where α represents the activities of the corresponding respective substances in aqueous solution at standard conditions). The free energy of aqueous substance formation values ( $\Delta$ G°<sub>298</sub>) are given in the table (values are at standard conditions). The shaded field indicates the partially overlapping facies of fumarate and succinate. The parageneses of substances for each facies are shown in the linear diagram of the C–O system. 3 HP and RC are the archaic 3-hydroxipropionate and reductive citrate cycles. The substance designations correspond to the designations in Figure 2.

According to the Gibbs phase rule, the diagram represents the fourphase nonvariant equilibria in this two-component C-O system (see Figure 2a), whereas the monovariant equilibria are three-phase, separating divariant fields of phase stability and their parageneses and are identified by linear diagrams in the system facies. Increasing the H<sub>3</sub>PO<sub>4</sub> chemical potential results in the stable paragenesis phosphoenolpyruvate - oxaloacetate at a relatively low chemical potential of methane (<-100 kJ/mol). Further development of the system leads to the facies formation of 3-phosphoglycerate and glyceraldehyde phosphate, permitting the development of the ancient pathway of substrate phosphorylation - gluconeogenesis and the reductive pentose phosphate (RPP) cycle of CO<sub>2</sub> fixation. Attention should be drawn to the overlapping facies (shaded field) of succinate (RC-cycle) and fumarate (3-HP cycle) with oxaloacetate - succinate fumarate - phosphoenolpyruvate paragenesis. This field is an area of thermodynamic stability of coupled RC + 3-HP bicycle [6] and the new metabolic pathways apparently originated at the beginning of these redundant autocatalytic metabolic systems.

The chemical potential of H<sub>2</sub> divides the lower region of the diagram (Figure 4) on the facies of the dicarboxylic acids (succinate  $\leftrightarrow$  fumarate) and thus fixes the phase areas of stability of the archaic autotrophic 3-HP and RC cycles of CO2 assimilation. With an increase in the chemical potential of H<sub>3</sub>PO<sub>4</sub>, facies of fumarate (3-HP cycle initiation) transform into facies of phosphoenolpyruvate - oxaloacetate, and a further increase in the chemical potentials of H<sub>2</sub> and H<sub>3</sub>PO<sub>4</sub> leads to the decomposition of the metastable paragenesis with the formation of facies of 3-phosphoglycerate, which is a central metabolite (and autocatalyst [47]) in both gluconeogenesis and the RPP autocatalytic CO<sub>2</sub> fixation cycle (see Figure 1). Similarly, increasing the H<sub>3</sub>PO<sub>4</sub> chemical potential in facies of succinate (RC

cycle initiation) also leads to the formation of facies with phosphoenolpyruvate - oxaloacetate paragenesis and further to facies of 3-phosphoglycerate.

According to [11], phosphoenolpyruvate and oxaloacetate are nodal molecules of all anabolic networks (in addition to acetate, pyruvate, and 2-oxoglutarate). From the diagrams in Figures 3 and 4, it is clear that facies with phosphoenolpyruvate - oxaloacetate paragenesis are an area of bifurcation (network node) that determines the development of the primordial 3-HP (fumarate) and RC cycles (succinate), system of gluconeogenesis (3-phosphoglycerate  $\leftrightarrow$  fructose 1,6-bisphosphate  $\rightarrow$ fructose 6-phosphate) and the RPP cycle (3-phosphoglycerate  $\rightarrow$ ribulose-1,5-bisphosphate  $\rightarrow$  3-phosphoglycerate). It is obvious that 3phosphoglycerate must be added to the five nodal molecules of autotrophic anabolic networks mentioned above. Thus, the diagram in Figure 3 and 4 represents a thermodynamic basis of "encrustation" of the C-H-O system by a chemical "shell" of phosphorus under the influence of a chemical potential of methane and/or hydrogen and phosphorus. However, the autotrophic nature of the parageneses formation of carboxylic acids and triose phosphates is manifested the best with consideration of the chemical potential of carbon dioxide.



**Figure 4:** Diagram of the chemical potentials of H<sub>2</sub> and H<sub>3</sub>PO<sub>4</sub> ( $\mu$ H<sub>2</sub>=RTln $\alpha$ H<sub>2</sub>, where  $\alpha$  represents the activity of molecular hydrogen in solution) at P=1 bar and T=298 K. The substance designations correspond to the designations in Figure 2. The arrows show the reversible reactions.

Figure 4 shows a diagram of the chemical potentials of hydrogen and  $H_3PO_4$ . In general, this diagram is similar to Figure 3 (also see Figures 2a and 2b), and the monovariant equilibria are three-phase or degenerate two-phase. As we have shown previously [6,26], the association of succinate with fumarate acts as a redox switch, turning electron flow in the direction of archaic chemoautotrophic 3-HP and RC cycles in the wide temperature range.

On the diagram of  $CO_2$  and  $H_3PO_4$  chemical potentials (Figure 5), a phosphoenolpyruvate - malate paragenesis is located in a sufficiently limited range of the considered chemical potentials (center diagrams) and develops in the direction of the discussed metabolic systems (Figure 1) at varying chemical potentials. Thus, increasing the chemical potential of phosphorus leads to the emergence of phases of 3-phosphoglycerate and glyceraldehyde phosphate (emerging at a low chemical potential of  $CO_2$ ), and these compounds (phosphorylated acid and aldehyde) are the initiators of gluconeogenesis and the RPP

#### Page 5 of 6

cycle. It should be noted that glyceraldehyde phosphate is also one of the main centers of the ancient evolutionary development of the nonoxidative branch of the pentose phosphate pathway, which leads to the formation of a series of phosphorylated sugars [48].



**Figure 5:** Diagram of the chemical potentials of CO<sub>2</sub> and H<sub>3</sub>PO<sub>4</sub> ( $\mu$ CO<sub>2</sub> = RTln $\alpha$ CO<sub>2</sub>, where  $\alpha$  represents the activity of CO<sub>2</sub> in aqueous solution at standard conditions). The parageneses of substances for each facies are shown in the linear diagram of the C–H system. Abbreviations as above.

Decreasing the chemical potential of phosphorus leads to the disappearance of the phosphoenolpyruvate - malate paragenesis and the appearance of phosphoenolpyruvate - pyruvate (with decreasing µCO<sub>2</sub>) and phosphoenolpyruvate - oxaloacetate (with increasing  $\mu$ CO<sub>2</sub>) parageneses. In general, increasing the CO<sub>2</sub> chemical potential leads to the carboxylation of compounds and their association with the formation of nodal facies - the center of phosphorus metabolism (malate - oxaloacetate - phosphoenolpyruvate). In this facies, as depicted in the diagram (Figures 3 and 4), the autocatalytic oxaloacetate - phosphoenolpyruvate paragenesis is also a "paragenesis bifurcation", meaning that this paragenesis determines the direction of natural selection of the primitive gluconeogenesis and CO<sub>2</sub> fixation metabolic pathways. According to Figure 1, phosphoenolpyruvate, oxaloacetate, and pyruvate assemblage are "turnstile - like" chemical mechanisms, allowing the direction of development of metabolic systems to change.

## Conclusion

The energy processes within the living cells are chiefly determined by an element such as phosphorus, and phosphates play a crucial role in the activation of organic molecules through phosphorylation. Therefore, it is reasonable to assume that some types of phosphorylated compounds also played a major role in the formation of the first protometabolic systems on the early Earth. Early evolution likely ensured the storage and consumption of energy in the substrate phosphorylation process [e.g., 49], which was the main driving force behind the development of both other metabolic systems and replication, and allowed the further development of energy pathways in aerobic and anaerobic organisms in all three domains of life.

Unlike other designs, the modular systems of metabolism are much more capable of evolutionary development [50,51]. The modules of the

core metabolic pathways are derived from the parageneses of certain substances (micro modules - associations of chemical compounds which ultimately function together), and the resulting modules are in turn in paragenesis with each other in certain physical and chemical conditions. Malate, oxaloacetate, pyruvate, and phosphoenolpyruvate (Figure 1) form the reversible "turnstile - like" mechanism capable of switching reaction directions. A change in the external conditions shifts the coupled modular autocatalytic system toward developing in the direction that is most favorable to the formation of specific metabolic systems. The chemical C-H-O system acquired new "layers" from phosphorus, nitrogen, and sulfur. The present work investigated only the phosphorus "layer", which led to the development of new protometabolic energy systems. Thus, in early chemical protometabolic reactions of the CO<sub>2</sub> fixation energetics, which primordially was provided by the partial energy of environmental chemical potentials (mainly due to the endogenous flow of hydrogen and hydrocarbons), was then replaced with energy from high-energy organic phosphates.

## Acknowledgments

This study was supported by the program of the Presidium of the Russian Academy of Sciences on fundamental researches of the Evolution of Organic World and Planetary Processes.

## References

- 1. Wächtershäuser G (1988) Before enzymes and templates: theory of surface metabolism. Microbiol Rev 52: 452-484.
- Wächtershäuser G (1990) Evolution of the first metabolic cycles. Proc Natl Acad Sci USA 87: 200–204.
- Morowitz HJ, Kostelnik JD, Yang J, Cody GD (2000) The origin of intermediary metabolism. Proc Natl Acad Sci U S A 97: 7704-7708.
- Smith E, Morowitz HJ (2004) Universality in intermediary metabolism. Proc Natl Acad Sci U S A 101: 13168-13173.
- Lindahl PA (2004) Stepwise evolution of nonliving to living chemical systems. Orig Life Evol Biosph 34: 371-389.
- Marakushev SA, Belonogova OV (2009) The parageneses thermodynamic analysis of chemoautotrophic CO2 fixation archaic cycle components, their stability and self-organization in hydrothermal systems. J Theor Biol 257: 588-597.
- Zachar I, Szathmáry E (2010) A new replicator: a theoretical framework for analysing replication. BMC Biol 8: 21.
- Peretó J1 (2012) Out of fuzzy chemistry: from prebiotic chemistry to metabolic networks. Chem Soc Rev 41: 5394-5403.
- 9. Braakman R, Smith E (2012) The emergence and early evolution of biological carbon-fixation. PLoS Comput Biol 8: e1002455.
- 10. Wächtershäuser G1 (2007) On the chemistry and evolution of the pioneer organism. Chem Biodivers 4: 584-602.
- 11. Srinivasan V, Morowitz HJ (2009) Analysis of the intermediary metabolism of a reductive chemoautotroph. Biol Bull 217: 222-232.
- 12. Hügler M, Sievert SM (2011) Beyond the Calvin cycle: autotrophic carbon fixation in the ocean. Ann Rev Mar Sci 3: 261-289.
- 13. Fuchs G1 (2011) Alternative pathways of carbon dioxide fixation: insights into the early evolution of life?Annu Rev Microbiol 65: 631-658.
- Stueken EE, Anderson RE, Bowman JS, Brazelton WJ, Colangelo-Lillis J, et. al. (2013) Did life originate from a global chemical reactor? Geobiology 11: 101-126.
- Marakushev SA, Belonogova OV (2010) Evolution of carbon dioxide archaic chemoautotrophic fixation system in hydrothermal systems. Dokl Biochem Biophys 433: 168– 174.
- Marakushev SA, Belonogova OV (2013) The divergence and natural selection of autocatalytic primordial metabolic systems. Orig Life Evol Biosph 43: 263-281.

Page 6 of 6

- Ronimus RS, Morgan HW (2003) Distribution and phylogenies of enzymes of the Embden-Meyerhof-Parnas pathway from archaea and hyperthermophilic bacteria support a gluconeogenic origin of metabolism. Archaea 1: 199–221.
- 18. Say RF, Fuchs G (2010) Fructose ,6-bisphosphate aldolase/phosphatase may be an ancestral gluconeogenic enzyme. Nature 464: 1077 1081.
- Pitsch S, Eschenmoser A, Gedulin B, Hui S, Arrhenius G (1995) Mineral induced formation of sugar phosphates. Orig Life Evol Biosph 25: 297-334.
- Cody GD, Boctor NZ, Filley TR, Hazen RM, Scott JH, et al. (2000) Primordial carbonylated iron-sulfur compounds and the synthesis of pyruvate. Science 289: 1337-1340.
- Keller MA, Piedrafita G, Ralser M2 (2015) The widespread role of nonenzymatic reactions in cellular metabolism.Curr Opin Biotechnol 34: 153-161.
- 22. Keller MA, Turchyn AV, Ralser M (2014) Non-enzymatic glycolysis and pentose phosphate pathway-like reactions in a plausible Archean ocean. Mol Syst Biol 10: 725.
- 23. Van Cappellen P, Ingall ED (1996) Redox stabilization of the atmosphere and oceans by phosphorus-limited marine productivity. Science 271: 493-496.
- 24. Pasek MA, Harnmeijer JP, Buick R, Gull M, Atlas Z (2013) Evidence for reactive reduced phosphorus species in the early Archean ocean.Proc Natl Acad Sci U S A 110: 10089-10094.
- 25. Shibuya T, Komiya T, Nakamura K, Takai K, Maruyama S (2010) Highly alkaline, high-temperature hydrothermal fluids in the early Archean ocean. Precambrian Res 182: 230–238.
- Marakushev SA, Belonogova OV (2011) Emergence of the chemoautotrophic metabolism in hydrothermal environments and the origin of ancestral bacterial taxa. Dokl Biochem Biophys 439: 161-166.
- 27. Korzhinskii DS (1959) Physicochemical basis of the analysis of the paragenesis of minerals. Consultants Bureau, New York.
- 28. Gibbs JW (1878) On the equilibrium of heterogeneous substances. Transact Connect Acad Arts & Sci 3: 108–248, 343–524.
- Marakushev AA, Marakushev SA (2006) PT facies of elementary, hydrocarbon, and organic substances in the C-H-O system. Dokl Earth Sci 406: 141–147.
- 30. Korzhinskii DS (1966) On thermodynamics of open systems and phase rule. Geochim Cosmochim Acta 30: 829-836.
- 31. Shock EL1 (1992) Chemical environments of submarine hydrothermal systems. Orig Life Evol Biosph 22: 67-107, 191-242.
- 32. Helgeson HC, Knox AM, Owens CE, Shock EL (1993) Petroleum, oil field waters, and authigenic mineral assemblages. Are they in metastable equilibrium in hydrocarbon reservoirs. Geochim Cosmochim Acta 57:P. 3295-3339.
- 33. Oelkers EH, Helgeson HC, Shock EL, Sverjensky DA, Johnson JW, et. al. (1995) Summury of the apparent standard partial molal Gibbs free energies of the formation of aqueous species, minerals, and gases at

pressures 1 to 5000 bars and temperatures 25 to 1000°C. J Phys Chem Ref Data 24: 1401-1560.

- Amend JP, Shock EL (2001) Energetics of overall metabolic reactions of thermophilic and hyperthermophilic Archaea and bacteria.FEMS Microbiol Rev 25: 175-243.
- Shock EL (1990) Geochemical constraints on the origin of organics compounds in hydrothermal systems. Orig Life Evol Biosph 20: 331-367.
- Shock EL, McCollom T, Schulte MD (1995) Geochemical constraints on chemolithoautotrophic reactions in hydrothermal systems. Orig Life Evol Biosph 25: 141-159.
- Shock EL, Schulte MD (1998) Organic synthesis during fluid mixing in hydrothermal systems. J Geophys Res 103: 28513–28527.
- 38. Desiraju GR1 (2002) Cryptic crystallography. Nat Mater 1: 77-79.
- Brazhkin VV (2006) Metastable phase and 'metastable' phase diagrams. J Phys Condens Matter 18: 9643-9650.
- 40. Mavrovouniotis ML (1991) Estimation of standard Gibbs energy changes of biotransformations. J Biol Chem 266: 14440-14445.
- 41. Alberty RA (2001) Effect of temperature on standard transformed Gibbs energies of formation of reactants at specified pH and ionic strength and apparent equilibrium constants of biochemical reactions. J Phys Chem B 105: 7865–7870.
- 42. Konhauser KO, Lalonde SV, Amskold L, Holland HD (2007) Was there really an Archean phosphate crisis?Science 315: 1234.
- 43. Li W, Czaja AD, Van Kranendonk MJ, Beard BL, Roden EE, et. al. (2013) An anoxic, Fe(II)-rich, U-poor ocean 3.46 billion years ago. Geochim Cosmochim Acta 120: 65-79.
- 44. Shock EL, Canovas P, Yang Z, Boyer G, Johnson K, et. al. (2013) Thermodynamics of organic transformations in hydrothermal fluids. Rev Mineral Geochem 76: 311-350.
- 45. Fu Q, Socki RA, Niles PB (2015) Evaluating reaction pathways of hydrothermal abiotic organic synthesis at elevated temperatures and pressures using carbon isotopes. Geochim Cosmochim Acta 154: 1–17.
- 46. McDermott JM, Seewald JS, German CR, Sylva SP (2015) Pathways for abiotic organic synthesis at submarine hydrothermal fields. Proc Natl Acad Sci U S A 112: 7668-7672.
- 47. Kun A, Papp B, Szathmary E (2008) Computational identification of obligatorily autocatalytic replicators embedded in metabolic networks. Genome Biol 9: R51.1-R51.11.
- 48. Stincone A., Prigione A, Cramer T, Wamelink MM, et al. (2015) The return of metabolism: biochemistry and physiology of the pentose phosphate pathway. Biol Rev Camb Philos Soc 90: 927-963.
- Ferry JG, House CH (2006) The stepwise evolution of early life driven by energy conservation. Mol Biol Evol 23: 1286-1292.
- Braakman R, Smith E (2013) The compositional and evolutionary logic of metabolism. Phys Biol 10: 011001.
- Clune J, Mouret JB, Lipson H (2013) The evolutionary origins of modularity. Proc Biol Sci 280: 20122863.