

## The Cancer Stem Cell Conundrum in Multiple Myeloma

Robert G. Hawley<sup>1,2\*</sup>

<sup>1</sup>Department of Anatomy and Regenerative Biology, George Washington University, Washington, DC, USA

<sup>2</sup>Sino-US Joint Laboratory of Translational Medicine, Jining Medical University Affiliated Hospital, Jining Medical University, Jining, Shandong, China

The cancer stem cell (CSC) hypothesis in its original form postulates that a small subpopulation of cancer cells is responsible for propagation of the tumor [1]. By comparison to normal stem cells, CSCs are predicted to be drug-resistant due to increased expression of proteins such as anti-alkylating enzymes like aldehyde dehydrogenase (ALDH) that neutralize the therapeutic agents [2] or members of the ATP-binding cassette (ABC) family of transporters that efflux them out of the cells [3].

Multiple myeloma (MM) is an incurable malignancy of B-lymphoid cells characterized by the accumulation of differentiated plasma cells in the bone marrow. MM is responsible for over 30,000 deaths each year in the United States and the European Union. While patients initially respond to therapy, they eventually relapse because the MM cells acquire drug resistance [4].

Demonstration of a low percentage of clonogenic cells in the bulk tumor mass prompted a search for the CSC in MM [5]. But contradictory results have been obtained regarding the phenotype of the proposed tumor-propagating cells; moreover, the relationship between drug-resistant MM cells at relapse and putative MM CSCs remains a matter of much debate [6-18]. A subpopulation of clonogenic MM cells has been described having a memory B cell-like phenotype (CD19<sup>+</sup>CD20<sup>+</sup>CD27<sup>+</sup>) [6]. Although CD19<sup>+</sup>CD20<sup>+</sup>CD27<sup>+</sup> MM cells lacked the characteristic plasma cell antigen CD138, they were capable of differentiating into CD138<sup>+</sup> plasma cells [7]. These studies suggested that MM is organized in a hierarchical manner and that CD19<sup>+</sup>CD20<sup>+</sup>CD27<sup>+</sup> MM cells might represent a putative MM CSC [19]. However, other work indicates that such cells might represent a premalignant intermediate [20]. Their biological significance has also been questioned based on their rarity. For example, one study investigating the clonal hierarchy in light chain MM was unable to confirm the presence of tumor-specific immunoglobulin sequences in the memory B cell compartment [10]. A number of other recent reports have also failed to obtain evidence in support of this supposition [13,14,16]. These latter results are consistent with the prevailing assumption that neoplastic transformation in MM occurs at a post-memory B cell stage when somatic hypermutation of immunoglobulin genes has ceased [21,22].

Various strategies have been employed to prospectively isolate and study CSC-like tumor-propagating cells. One common approach is based on the expression of cell surface markers that are characteristic of the stem cell phenotype of the corresponding normal tissue. An example of this approach involves expression of the CD34 cell surface marker of immature hematopoietic cells (see [23] for review). It is of interest in this regard that a subpopulation of CD138<sup>+</sup> MM cells has been reported to express CD34 [8].

Another approach capitalizes on the functional properties of stem cells. As noted above, stem cells are highly resistant to damage by toxic agents through a combination of mechanisms [2,3,24,25]. Some of these attributes can be exploited by flow cytometry-based procedures to enrich for stem-like cells [23,26]. Efflux of the vital dye Hoechst 33342 by the ABCG2 and/or ABCB1 transporters identifies a subset of cells in a variety of normal and malignant tissues—termed “side

population” (SP) cells—which displays stem cell-like properties [27,28]. Interestingly, variable results have also been obtained concerning the SP phenotype in human MM cell lines and patient samples. Using this assay, one group identified a clonogenic CD138<sup>neg</sup> MM subpopulation that was resistant to the anti-MM agent lenalidomide [7] whereas another group subsequently described the characterization of clonogenic SP cells in MM that primarily expressed CD138 and were sensitive to lenalidomide [9]. Likewise, ALDH has been shown to be a marker of CSC-like cells in a wide range of tumors, including the B-lymphoid malignancies Hodgkin lymphoma and mantle cell lymphoma [29,30]. Although it has been reported that certain MM cell cultures as well as patient samples contained subpopulations of ALDH<sup>+</sup> cells with a CSC-like phenotype [7,18], the generality of this finding has been questioned [17].

How can these discrepant observations be reconciled? On the one hand, it is important to appreciate that MM is characterized by significant molecular heterogeneity, comprising at least seven disease subtypes [31]. A potential scenario that could also help to integrate the incongruent observations would be if malignant transformation of CD138<sup>+</sup> post-memory B cells results in the acquisition of a CSC-like phenotype [32], e.g., by a “dedifferentiation” mechanism that is akin to the cellular reprogramming that occurs during the generation of induced pluripotent stem cells [22,33,34]. Indeed, activation of the *MYC* proto-oncogene, one of four transcription factors used in the initial reprogramming experiments [35], is a recurring event in MM pathogenesis [36].

Thus, the putative MM CSC would not be expected to be a single genetic entity; rather, genetically-distinct subtype-associated CSCs are predicted. Furthermore, it would not be surprising if MM CSCs exhibit phenotypic variability during tumor progression as a result of epigenetic changes and genomic instability [37]. Considered in this light, it will be a challenging task but well worth the effort to delineate all of the MM CSC subpopulations. The clinical implications are profound in that subtype-specific concerted therapies targeting the bulk as well as the various CSC fractions of the tumor will undoubtedly be necessary if an effective cure is to be found for this devastating collection of diseases.

### Acknowledgement

This work was supported by a Grant from the Dr. Cyrus and Myrtle Katzen Cancer Research Center at The George Washington University.

\*Corresponding author: Robert G. Hawley, Department of Anatomy and Regenerative Biology, George Washington University, Washington, DC, USA, E-Mail: [rghawley@gwu.edu](mailto:rghawley@gwu.edu)

Received October 19, 2012; Accepted October 20, 2012; Published October 22, 2012

Citation: Hawley RG (2012) The Cancer Stem Cell Conundrum in Multiple Myeloma. J Stem Cell Res Ther 2:e110. doi:10.4172/2157-7633.1000e110

Copyright: © 2012 Hawley RG. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## References

1. Dick JE (2008) Stem cell concepts renew cancer research. *Blood* 112: 4793-4807.
2. Alison MR, Guppy NJ, Lim SM, Nicholson LJ (2010) Finding cancer stem cells: are aldehyde dehydrogenases fit for purpose? *J Pathol* 222: 335-344.
3. Moitra K, Lou H, Dean M (2011) Multidrug efflux pumps and cancer stem cells: insights into multidrug resistance and therapeutic development. *Clin Pharmacol Ther* 89: 491-502.
4. Mahindra A, Laubach J, Raje N, Munshi N, Richardson PG, et al. (2012) Latest advances and current challenges in the treatment of multiple myeloma. *Nat Rev Clin Oncol* 9: 135-143.
5. Hamburger A, Salmon SE (1977) Primary bioassay of human myeloma stem cells. *J Clin Invest* 60: 846-854.
6. Matsui W, Huff CA, Wang Q, Malehorn MT, Barber J, et al. (2004) Characterization of clonogenic multiple myeloma cells. *Blood* 103: 2332-2336.
7. Matsui W, Wang Q, Barber JP, Brennan S, Smith BD, et al. (2008) Clonogenic multiple myeloma progenitors, stem cell properties, and drug resistance. *Cancer Res* 68: 190-197.
8. Kuranda K, Berthon C, Dupont C, Wolowiec D, Leleu X, et al. (2010) A subpopulation of malignant CD34+CD138+B7-H1+ plasma cells is present in multiple myeloma patients. *Exp Hematol* 38: 124-131.
9. Jakubikova J, Adamia S, Kost-Alimova M, Klippel S, Cervi D, et al. (2011) Lenalidomide targets clonogenic side population in multiple myeloma: pathophysiological and clinical implications. *Blood* 117: 4409-4419.
10. Pfeifer S, Perez-Andres M, Ludwig H, Sahota SS, Zojer N, et al. (2011) Evaluating the clonal hierarchy in light-chain multiple myeloma. *Leukemia* 25: 1213-1216.
11. Van Valckenborgh E, Matsui W, Agarwal P, Lub S, Dehui X, et al. (2012) Tumor-initiating capacity of CD138- and CD138+ tumor cells in the 5T3 multiple myeloma model. *Leukemia* 26: 1436-1439.
12. Chiron D, Surget S, Maïga S, Bataille R, Moreau P, et al. (2012) The peripheral CD138+ population but not the CD138- population contains myeloma clonogenic cells in plasma cell leukaemia patients. *Br J Haematol* 156: 679-683.
13. Trepel M, Martens V, Doll C, Rahlf J, Gösch B, et al. (2012) Phenotypic detection of clonotypic B cells in multiple myeloma by specific immunoglobulin ligands reveals their rarity in multiple myeloma. *PLoS One* 7: e31998.
14. Hosen N, Matsuoka Y, Kishida S, Nakata J, Mizutani Y, et al. (2012) CD138-negative clonogenic cells are plasma cells but not B cells in some multiple myeloma patients. *Leukemia* 26: 2135-2141.
15. Christensen JH, Jensen PV, Kristensen IB, Abildgaard N, Lodahl M, et al. (2012) Characterization of potential CD138 negative myeloma "stem cells". *Haematologica* 97: e18-e20.
16. Kim D, Park CY, Medeiros BC, Weissman IL (2012) CD19-CD45low/-CD38high/CD138+ plasma cells enrich for human tumorigenic myeloma cells. *Leukemia* doi: 10.1038/leu.2012.140 [Epub ahead of print].
17. Páino T, Ocio EM, Paiva B, San-Segundo L, Garayoa M, et al. (2012) CD20 positive cells are undetectable in the majority of multiple myeloma cell lines and are not associated with a cancer stem cell phenotype. *Haematologica* 97: 1110-1114.
18. Boucher K (2012) Stemness of B cell progenitors in multiple myeloma bone marrow. *Clin Cancer Res*: doi: 10.1158/1078-0432.CCR-12-0531 [Epub ahead of print].
19. Szczepek AJ, Bergsagel PL, Axelsson L, Brown CB, Belch AR, et al. (1997) CD34+ cells in the blood of patients with multiple myeloma express CD19 and IgH mRNA and have patient-specific IgH VDJ gene rearrangements. *Blood* 89: 1824-1833.
20. Rasmussen T, Haaber J, Dahl IM, Knudsen LM, Kerndrup GB, et al. (2010) Identification of translocation products but not K-RAS mutations in memory B cells from patients with multiple myeloma. *Haematologica* 95: 1730-1737.
21. Bakkus MH, Heirman C, Van Riet I, Van Camp B, Thielemans K, et al. (1992) Evidence that multiple myeloma Ig heavy chain VDJ genes contain somatic mutations but show no intracлонаl variation. *Blood* 80: 2326-2335.
22. Takishita M, Kosaka M, Goto T (1994) Cellular origin and extent of clonal involvement in multiple myeloma: genetic and phenotypic studies. *Br J Haematol* 87: 735-742.
23. Hawley RG, Ramezani A, Hawley TS (2006) Hematopoietic stem cells. *Methods Enzymol* 419: 149-179.
24. Diehn M, Cho RW, Lobo NA, Kalisky T, Dorie MJ, et al. (2009) Association of reactive oxygen species levels and radioresistance in cancer stem cells. *Nature* 458: 780-783.
25. Riz I and Hawley RG (2009) Genomic stability in stem cells. In: Rajasekhar VK, Vemuri MC, editors. *Regulatory Networks in Stem Cells*. New York, NY: Humana Press/Springer: 67-74.
26. Eaker SS, Hawley TS, Ramezani A, Hawley RG (2004) Detection and enrichment of hematopoietic stem cells by side population phenotype. *Methods Mol Biol* 263: 161-180.
27. Goodell MA, Brose K, Paradis G, Conner AS, Mulligan RC (1996) Isolation and functional properties of murine hematopoietic stem cells that are replicating in vivo. *J Exp Med* 183: 1797-1806.
28. Ramos CA, Venezia TA, Camargo FA, Goodell MA (2003) Techniques for the study of adult stem cells: be fruitful and multiply. *BioTechniques* 34: 572-578, 580-584, 586-591.
29. Jones RJ, Gocke CD, Kasamon YL, Miller CB, Perkins B, et al. (2009) Circulating clonotypic B cells in classic Hodgkin lymphoma. *Blood* 113: 5920-5926.
30. Brennan SK, Meade B, Wang Q, Merchant AA, Kowalski J, et al. (2010) Mantle cell lymphoma activation enhances bortezomib sensitivity. *Blood* 116: 4185-4191.
31. Zhan F, Huang Y, Colla S, Stewart JP, Hanamura I, et al. (2006) The molecular classification of multiple myeloma. *Blood* 108: 2020-2028.
32. Luckey CJ, Bhattacharya D, Goldrath AW, Weissman IL, Benoist C, et al. (2006) Memory T and memory B cells share a transcriptional program of self-renewal with long-term hematopoietic stem cells. *Proc Natl Acad Sci USA* 103: 3304-3309.
33. Yaccoby S (2005) The phenotypic plasticity of myeloma plasma cells as expressed by dedifferentiation into an immature, resilient, and apoptosis-resistant phenotype. *Clin Cancer Res* 11: 7599-7606.
34. Vierbuchen T, Wernig M (2012) Molecular roadblocks for cellular reprogramming. *Mol Cell* 47: 827-838.
35. Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T, et al. (2007) Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell* 131: 861-872.
36. Chng WJ, Huang GF, Chung TH, Ng SB, Gonzalez-Paz N, et al. (2011) Clinical and biological implications of MYC activation: a common difference between MGUS and newly diagnosed multiple myeloma. *Leukemia* 25: 1026-1035.
37. Visvader JE, Lindeman GJ (2012) Cancer stem cells: current status and evolving complexities. *Cell Stem Cell* 10: 717-728.