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Synthesis and Characterization of α , α -Dimethyl-4-[1-Hydroxy-4-[4-(Hydroxyldiphenyl- Methyl)-1-Piperidinyl]Butyl] Benzeneacetic Acid Metal Complexes of Biological Interest

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Abstract

 α,α -Dimethyl-4-[1-hydroxy-4-[4-(hydroxydiphenylmethyl)-1-piperidinyl]butyl]benzene acetic acid generically known as fexofenadine or carboxyterfenadine is a second generation H₁-receptor antagonist, much widely used due to its non-sedative effects. Metal complexes of fexofenadine with various essential and trace elements of biological interest, have been synthesized and characterized by FT-IR, ¹H-NMR, UV, CHN elemental analysis. Conductometric titration was carried out to determine the mole ratio of interaction of the drug and metal. Spectral data clearly showed the complexation of fexofenadine with nitrogen of the piperidine ring in case of magnesium, calcium, chromium and manganese complexes while oxygen of carboxylato group is involved in complexation with iron, cobalt nickel, copper, zinc and cadmium. Elemental analysis reveals monodentate character of these complexes.

Keywords: Fexofenadine; H₁-receptor antagonist; Essential and trace elements; FT-IR; ¹H-NMR

Introduction

Fexofenadine (carboxyterfenadine) (Figure 1), is α,α -Dimethyl-4-[1-hydroxy-4-[4-(hydroxydiphenylmethyl)-1-piperidinyl]butyl] benzeneacetic acid is a non-anticholinergic, non sedating-type histamine H -receptor antagonist [1, 2] and an active metabolite of terfenadine [3-5]. Bioavailability of fexofenadine decreases when the drug is co-administered with grapefruit juice, orange juice and apple juice which potentially reduced pharmacokinetic effects of fexofenadine [6] and also lowers the plasma drug concentrations by reducing drug absorption catalyzed by the Organic Anion Transporting Polypeptide (OATP) [7].

Literature survey reveals few studies on the interaction of metal ions with fexofenadine [8]. Metals are considered essential to a human body being an integral part of an organic structure in performing physiologically important and vital functions in the body [9]. It seems that the role of metal ions is imperative for the way of function of fexofenadine. Synthesis and characterization of new metal complexes with fexofenadine are of great importance for better understanding of drug-metal ion interactions [10]. A thorough survey of the literature has revealed that no work has been reported on the synthesis of fexofenadine metal complexes, though our research group has reported a number of metal complexes with other co-administered drugs as



levofloxacin [11], enoxacin [12,13], sparfloxacin [14], gatifloxacin [15], gliquidone [16] and atenolol [17].

In this paper, we present the synthesis of a series of fexofenadinemetal complexes with metals of biological interest in an attempt to find the mode of coordination as well as to study drug-metal interactions. More specifically, the complexes have been synthesized and characterized by elemental analysis and diverse spectroscopic techniques (IR, UV-Vis and NMR techniques).

Experimental

Materials

Fexofenadine was a kind gift from Getz Pharmaceuticals (Pvt.) Ltd. Karachi, Pakistan. The metal salts and other chemicals were of analytical grade (Merck Germany). All the glassware's were washed with chromic acid followed by a thorough washing with deionized water which was freshly prepared in the laboratory.

Instrumentation

Conductometric studies were carried on Vernier LabPro. Data acquisition and analysis were carried out by using the Logger pro 3.2 software. Thin Layer Chromatography (TLC) was performed on a HSF-254 TLC plate and the samples were visualized under a UV lamp, melting point of complexes was recorded on a Gallenkamp electrothermal melting point apparatus and is uncorrected. The characterization of fexofenadine metal complexes was carried out by FT-IR spectrophotometer (Shimadzu Prestige-21 200 VCE), coupled to a P IV-PC and loaded with IR resolution software. The disks were

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placed in the holder directly in the IR beam. Spectra were recorded at a resolution of 2cm^{-1} , and 50 scans were accumulated. Stability of these complexes was studied 4.5 CF stability chamber from PARAMETER Generation and Control USA with constancy control \pm 0.5%.

Proton NMR studies of metal complexes were carried out on a Brucker AMX 500MHz instrument in deuterated methanol (CD_3OD) using TMS as an internal standard. CHN elemental studies were carried out by standard micro methods using Carlo Erba 1106.

Conductometric titrations

Prior to the synthesis of the metal complexes, the stoichiometry of complexes was determined using conductometric titration technique [18]. 0.1 mM metal solution was titrated against the drug solution at 298 K with constant stirring and change in conductivity was measured until the addition resulted in no change of conductivity values of the resulting solution. Similar process was repeated for all metals.

Synthesis of metal complexes

Metal complexes of fexofenadine were synthesized with magnesium, calcium, chromium, manganese, ferric, ferrous, cobalt, nickel, copper, zinc and cadmium hydrated salts. The synthesis of magnesium, calcium, chromium and manganese complexes with fexofenadine were carried out in M:L ratio of 1:2 while for rest of the metals M:L ratio was 1:1. These ratios were determined earlier by conductometric measurements. Drug and metal salts were dissolved separately in hot methanol (20 mL) and deionized water (10 mL). Both solutions were mixed with constant stirring and then refluxed for about three hours. The reactions were monitored by thin layer chromatography. The volume of the reaction mixture was then reduced by evaporation, filtered while hot and then kept undisturbed for crystal growth at room temperature.

The growth of crystals depended upon many factors like type of crystal, environment, concentration of solution, temperature etc. Due to this all these metal complexes had variable time of crystallization. Crystals of magnesium, calcium, chromium and manganese complexes were obtained within 10 to 15 days, while the iron complex took one month for crystallization. Cobalt, nickel, copper, zinc and cadmium complexes were crystallized in 20~25 days. Crystals of different colors for different complexes were obtained. Finally, these were filtered, washed with water and methanol, and vacuum dried. Their melting points were recorded and were recrystallized from absolute methanol till constant melting point. Their physicochemical parameters were noted.

Physical data

Fexofenadine: Color white, m.p. 190°C, UV nm (ε): 1.775(218 nm), IR (KBr) vmax: 3400-3350 db, s, 3058.04 s, 1700 s, 1600 and 1475sm 1447.60 m 1403.55 m, 1279 m, 1166.41 m,1099 s, 1068m, 983, 965 db, sm, 834 ms, 747, 702,638, 577, 525.53 dp, s. 1H NMR (MeOD, 400 MHz) : 7.19 (s, J=1.16 Hz), 7.11 (s, J=0.5 Hz) and 7.06 aromatic, 2.0 N-H, 1.52 CH₃, 2.29, 2.19 CH₂-N, 2.34 (t, J=12.2 Hz) -CH₂-CH₂-N, 2.36 (t, J=14.0 Hz), 1.39, 1.77 and 4.5 (s, J=1.0 Hz) -CH₂ open chain, 4.95 (s) alcoholic. Calculated for $C_{32}H_{39}NO_4$: C, 49.14; H, 5.96; N, 3.18. Found: C, 49.01; H, 5.74; N, 4.33.

Fexofenadine magnesium complex: Colorless, Yield 50%, m.p. 112°C, UV nm (ε): 0.4615(207 nm), IR (KBr) vmax: 3421 s, 2967 db, s, 1680 dp, s, 1446.45 sm, 1320 db,sm 0.4615(207 nm) 1300-1320 db, sm, 1100 sm, 1090 sm, 1410, 1637 sm, 1151 m, 750 sm, 707 sm, 1H NMR (MeOD, 400 MHz) δ: 7.10, 7.06 (s, J=0.69Hz) aromatic, 1.65 MgOOC-C-CH₂, 2.36, 1.19 (m, J=0.77Hz)CH₂-CH₂-N-Mg, 1.94 (t, J=0.99 Hz) CH₂-N-Mg, $-(CH_{2})_3$ -N-Mg, 1.79 1.69 (t, J=4.5Hz) Mg-N-CH₂, 2.42,

1.64, 1.44 -CH₂ open chain, 1.76 (s) alcoholic, 3.49, 1.58 (t) -CH₂-OH. Calculated for $C_{32}H_{39}ClMgNO_4$: C, 68.46; H, 7.00; N, 2.49. Found: C, 68.42; H, 7.07; N, 2.43.

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Fexofenadine calcium complex: Colorless Yield 42%, m.p. 72°C, UV nm (ε): 0.5173(209nm), IR (KBr) vmax: 3650 sm, 3411 m, 2967 db, sm, 1716.58 db, m 1619.35 0.5173(209nm) sm, -CH₂1446.84 m, 1383.80 sm, 1100-1140 db, 1490 sm, 750, sm, 706 sm, 668.14 sm, 602.08 sm. 1H NMR (MeOD, 400 MHz) δ: 7.34, 7.07, 7.23, 7.34 (s, J=0.63Hz) aromatic, 1.65 CaOOC-C-CH₂. 2.92 (t,J=3.5 Hz) CH₂-NH, 1.85 (m, J=3.5 Hz) -CH₂-NH, 1.94 (t, J=14.0Hz), 1.64, 1.44 -(CH₂)₃-NH open chain, 4.28 (J=9.0Hz) (piperidine)(Ph)-CH-OH, 1.90 (J=3.5Hz) NH-(CH₂)₂-CH, 1.43 (s) alcoholic. Calculated for $C_{32}H_{39}$ ClCaNO₄: C, 66.59; H, 6.81; N, 2.43. Found: C, 66.63; H, 6.85; N, 2.40.

Fexofenadine chromium complex: Color light green Yield 39%, m.p. 72°C, UV nm (ε): 0.28(257nm), IR (KBr) vmax: 3650 sm, 2967 db, sm 1725 db, 1691 sm, 1490 m, 1447 m, 1310 db, sm, 1146 m, 1471.97 db, m, 750 sm, 706 sm. 1H NMR (MeOD, 400 MHz) δ: 7.13 (s, J=7.67Hz) 7.18 (s, J=2.12Hz) aromatic, 4.16 (J=0.31Hz) -CH₂-COOCr, 1.91, 1.88 (t, J=12.2 Hz) CH₂-N-Cr, 1.35 (m, J=2.0 Hz) (m, J=7.69 Hz), -CH₂-CH₂-N, 2.40 (t, J=14.0Hz), 1.64, 1.50 (t, J=0.5 Hz) (CH₂)₃-N-Cr, 3.32 (J=4.8Hz) (piperidine)(Ph)-CH-OH, 1.76 (s) alcoholic. Calculated for $C_{32}H_{39}$ ClCrNO₄: C, 65.24; H, 6.67; N, 2.38. Found: C, 65.22; H, 6.69; N, 2.41.

Fexofenadine manganese complex: Colorless Yield 34%, m.p. 128°C, UV nm (ε): 0.7 (215 nm), IR (KBr) vmax: 3647 sm , 3420 sm, 2969.91 db, m, 1717 db, 1653 trp, 1490 sm, 1458 m, 1397 db, sm, 1365 db, 1146-1099.36 trp, 840 m, 750 sm, 706 sm. 1H NMR (MeOD, 400 MHz) : 7.13 (s, J=0.31Hz) aromatic, 7.18 (s, J=0.77Hz) 1.46, 3.12 -CH₂-COOMn, 2.27, 1.79, 1.69 CH₂-N-Mn, 1.69 (t, J=0.77 Hz) 1.46 (q, J=2.0 Hz) (t, J=12.2 Hz) -CH₂-CH₂-N, (CH₂)₃-N-Mn open chain at 1.39, 3.39, 1.44 (t, J=6.8 Hz) 4.5 (J=6.8Hz) -CH-O-Mn, 3.49 (J=4.8Hz) (piperidine) (Ph)-CH-OH, 2.0 (s) alcoholic. Calculated for C₃₂H₃₉ClMnNO₄ : C, 64.92; H, 6.64; N, 2.37. Found: C, 64.97; H, 6.62; N, 2.31.

Fexofenadine ferric chloride complex: Color Yellow Yield 19%, m.p. 112°C, UV nm (ε): 0.7 (215 nm), IR (KBr) vmax: 3425 m, 3055 db, m, 2968 db, 1716 m, 1591 sm, 481-1415 trp, sm, 1373 db, 1151 s, 790 s, 750sm, 636sm. 1H NMR (MeOD, 400 MHz) δ: 7.34 (s, J=0.69Hz), 7.23 (s, J=0.63Hz), 7.38 (s, J=7.21Hz) 7.07 (s, J=7.21Hz) (s, J=1.11Hz) 7.18 (s, J=0.77Hz) aromatic, 1.64 – (CH₃)₂-C-COOFe, 2.93, 2.92 CH₂-N, 1.73 1.85 -CH₂-CH₂-N, 2.41 1.52 1.64 (CH₂)₃-N, open chain at 4.61 (J=4.8Hz) (piperidine)(Ph)-CH-OH, 1.36 (s) alcoholic. Calculated for $C_{36}H_{51}FeNO_4$: C, 70.01; H, 8.32; N, 2.27. Found: C, 70.06; H, 8.37; N, 2.31.

Fexofenadine ferric ammonium citrate complex: Color light yellow Yield 19%, m.p. 210°C, UV nm (ε): 0.348 (254 nm), IR (KBr) vmax: 3405 sm, 2980 db, m, 1717 db, sm, 1600 s, 1491 sm, 1447 m, 1689 db, m, 1254 db, sm, 1154 sm, db, 1066 db, sm, 845 sm, 750 s, 704 sm, 637 sm. 1H NMR (MeOD, 400 MHz) δ: 7.34 (s, J=0.69Hz), 7.23 (s, J=0.63Hz), 7.38 (s, J=7.21Hz) 7.07 (s, J=7.21Hz) (s, J=1.11Hz) 7.18 (s, J=0.77Hz) aromatic, 1.64 – (CH₃)₂-C-COOFe, 2.93, 2.92 CH₂-N, 1.73 1.85 -CH₂-CH₂-N, 2.41 1.52 1.64 (CH₂)₃-N, open chain at 4.61 (J=4.8Hz) (piperidine)(Ph)-CH-OH, 1.36 (s) alcoholic. Calculated for $C_{36}H_{51}FeNO_4$: C, 70.01; H, 8.32; N, 2.27. Found: C, 70.06; H, 8.36; N, 2.22.

Fexofenadine ferrous sulfate complex: Color yellow Yield 15%, m.p. 137°C, UV nm (ϵ): 0.517(216nm), IR (KBr) vmax: 3450 sm, 3055 db, sm, 2969 db, sm, 1718 s db, 1448 m, 1264 sm, 1150-1098 trp, 840 sm, 749 sm, 708 sm, 618 sm. 1H NMR (MeOD, 400 MHz) δ : 7.11, 7.06 (s, J=7.21Hz), (s, J=0.63 Hz) aromatic, 1.46, 1.52 –CH₃ 11.0 COOH,

2.19, 2.92 CH₂-N, 1.34, 1.59 -CH₂-CH₂-N, (CH₂)₃-N open chain at 2.36, 1.39, 1.77, 4.50 (t, J=0.5 Hz) (J=4.8Hz) 3.49 (piperidine)-CH-OH, 2.0 (s) (J=11.9Hz) alcoholic. Calculated for $C_{36}H_{51}FeNO_4$: C, 70.01; H, 8.32; N, 2.27. Found: C, 70.06; H, 8.35; N, 2.26.

Fexofenadine cobalt complex: Color blue Yield 20%, m.p. 118°C, UV nm (ε): 0.6(202nm), IR (KBr) vmax: 3420 m, 2969 db, m, 1717 m db, 1447 m, 1380 db, sm, 1151 sm, 1180-1100 db, m, 790 sm, 750 sm, 706 m. 1H NMR (MeOD, 400 MHz) δ: 7.06 (s, J=2.31Hz) 7.28 (s, J=2.12Hz) aromatic, 1.46, 1.64 $-(CH_3)_2$ -C-COOCo, 2.24 (t, J=7.5 Hz) CH₂-N-Co, 1.50 (t, J=7.5 Hz) -CH₂-CH₂-N, 1.39 (t, J=7.0 Hz), 3.39, 1.77 (t, J=1.0 Hz) protons, 4.5 (J=1.0Hz) (CH₂)₃-N-Co open chain, 2.0 (s) alcoholic. Calculated for C₃₆H₅₁CoNO₄: C, 69.66; H, 8.28; N, 2.26. Found: C, 69.61; H, 8.228; N, 2.30.

Fexofenadine nickel complex: Color light green Yield 33%, m.p. 212°C, UV nm (ε): 0.78(204nm), IR (KBr) vmax: 3390 m, 2969 db, m, 1718 s db, 1447 m, 1380 db, sm, 1300 sm, 1150-1098 db, 950-990 trp, sm, 850 sm, 750 sm, 707 m, 637-619 db, m. 1H NMR (MeOD, 400 MHz) δ: 7.19 (s, J=7.12Hz), 7.08 (s, J=0.5 Hz), 7.38, 7.06, 7.11 aromatic, 1.64 $-(CH_3)_2$ -C-COONi, 2.29 2.19 (t, J=2.0 Hz), CH₂-N, 1.59, 1.34 $-CH_2$ -CH₂-N, 3.2, 1.39, 1.77 (t, J=7.0 Hz) 4.5(CH₂)₃-N open chain, 4.49 (J=4.8Hz) (piperidine)(Ph)-CH-OH, 2.0 (s) alcoholic. Calculated for C₃₆H₅₁NNiO₄: C, 69.68; H, 8.28; N, 2.26. Found: C, 69.64; H, 8.23; N, 2.29.

Fexofenadine copper complex: Color light blue Yield 44%, m.p. 110°C, UV nm (ε): 1.16(244nm), IR (KBr) vmax: 3390sm, 2968db,m, 1717 sm db, 1447 m, 1388 db, sm, 1300 sm, 1150 sm, 1150-1100 db, m, 1000- 1050 db, sm, 850 sm, 790 m, 706 sm, 750 sm. 1H NMR (MeOD, 400 MHz) : 6.99, 7.07 aromatic, 4.45 -CH₂-COOCu, 2.29, 2.19 (t, J=3.5 Hz) CH₂-N, 1.59, 1.34 -CH₂-CH₂-N, 2.36 (t, J=14.0 Hz) 1.39 (t, J=7.0 Hz) 1.77 (t, J=6.3 Hz) 4.5 (CH₂)₃-N open chain, 3.49 (J=4.8Hz) Ph-CH₂-OH, 3.9 (s) alcoholic. Calculated for $C_{36}H_{51}$ CuNO₄: C, 69.14; H, 8.22; N, 2.24. Found: C, 69.10; H, 8.29; N, 2.27.





Fexofenadine zinc complex: Color dirty white Yield 46.64%, m.p. 102°C, UV nm (ε): 0.86(217nm), IR (KBr) vmax: 3618 sm, 3376 db, 2924 m, db, 1717, 1697 db m, 1605 sm, 1596-1508 trp, m, 1480 s, 1310 db, sm, 1220 s, 1115 db, sm, 1033-1007 db, 753 m, 537 sm, 468 sm. 1H NMR (MeOD, 400 MHz) δ: 6.96, 7.08, 7.21 aromatic, 3.49 -CH₂-COOZn, 2.29, 2.19 (t, J=2.0 Hz), CH₂-N, 1.59, 1.34 (t, J=1.34 Hz), -CH₂-CH₂-R, 3.9 (t, J=14.0 Hz), 1.39 (t, J=8.0 Hz), 1.77 (t, J=7.0 Hz) 4.5 (CH₂)₃-N open chain, 2.51 (J=4.8Hz) (piperidine)(Ph)-CH-OH, 5.5 (s) alcoholic. Calculated for $C_{36}H_{51}NO_4Zn$: C, 68.94; H, 8.20; N, 2.23. Found: C, 68.91; H, 8.26; N, 2.28.

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Fexofenadine cadmium complex: Colorless Yield 42.5%, m.p. 132°C, UV nm (ε): 1.95(221nm), IR (KBr) vmax: 3629 sm, 3450 s, 3070 db, m, 1710-1725 db, sm, 1491-1972, db, sm, 1447 m, 1410 db, sm, 1270 db sm, 1154 m, 1099-1017 db, m, 950 db, sm, 860 sm, 940 sm, 750 sm. 1H NMR (MeOD, 400 MHz) δ: 6.99, 7.30 aromatic, 3.6 –CH₂-COOCd, 2.29, 2.19 (t, J=3.75 Hz)(t, J=7.69 Hz) CH₂-N, 1.59, 1.34 (t, J=5.75 Hz), -CH₂-CH₂-N, 2.36 (t, J=14.0 Hz), 1.39 (t, J=8.0 Hz), 1.77 (t, J=7.0 Hz) 4.5 (CH₂)₃-N open chain, 3.49 (J=5.75Hz) piperidine-CH-OH, 4.4 (s) alcoholic. Calculated for $C_{36}H_{51}$ CdNO₄: C, 64.13; H, 7.62; N, 2.08. Found: C, 64.17; H, 7.67; N, 2.11.

Results and Discussion

Fexofenadine is a zwitterion having good solubility in stomach and small intestinal fluids. Grapefruit juice decreases the dissolution of fexofenadine because of its interaction with excipients included in formulation to impair this process [19]. In the light of this study was initiated to explore fexofenadine interactions with metals of biological interest.

Transition metal complexes of fexofenadine were synthesized in mole ratios as described earlier in methanol. Prior to synthesis complexation studies between fexofenadine and metals were investigated by conductometric titrations. The effects on conductivity caused by the increased volume of reaction medium were compensated by correcting it for dilution by correction term [(v + V)/V] and equivalence point were calculated. Molar conductivity values (Λ_m) were then calculated by correcting conductivity values (μ S) using the formula $\Lambda_m = K/C$, where, *K* is the measured conductivity and *C* is the electrolyte concentration.

From the graph it is found that magnesium, calcium, chromium and manganese metal ions bind to ligand in the 2:1 ratio, whereas nickel, copper, zinc, cadmium and iron complexes are formed in equimolar ratio (Figure 2).

Metal complexes of fexofenadine were synthesized by refluxing fexofenadine with metal salt solutions in a mixture of methanol and water in equimolar ratios and then crystallizing them at room temperature. Solubility and melting points were noted; all these complexes are 50% soluble in methanol and 25% soluble in ethanol, ethyl acetate and acetone.

Stability of these complexes was studied by taking their melting points at an interval of 24 hours for seven days according to Q1A (R2) compliant [20]. Samples were stored at 25°C/60% RH, 30°C/60% RH, 30°C/65% RH, 40°C/75% RH, 30°C/75% RH and 25°C/40% RH. Melting points of the complexes were recorded after 24 hours for 7 days. No appreciable changes in the melting points were observed, and the estimated error was \pm 1°C. It was concluded that the complexes were stable according to ICH guidelines [21]. Their structures were established from the elemental analyses, which agree well with their proposed formulae.





Infrared studies

Fexofenadine metal complexes were characterized by studying the most typical vibrations in the IR spectrum in the region 400-4000 cm⁻¹. In the IR spectra of metal complexes, some very prominent peak shifting has been observed along with a change in intensities of several important peaks indicating complexation of fexofenadine with metals (Figures 3 and 4).

The major infrared absorption bands of fexofenadine for -OH stretching occurred in the region 3300-3370 cm⁻¹ and out of plane bending at 965 cm⁻¹, C-H stretching at 3058.04 cm⁻¹ due to strained molecule because of the substitution of aromatic rings, a sharp band of C=O stretching at 1700 cm⁻¹, at 1600 and 1475 cm⁻¹, C=C aromatic medium weak bands, at 1447.60 cm⁻¹ bending absorptions of $-CH_2$ C-O medium stretching vibration at 1279 cm⁻¹, at 1099 cm⁻¹C-N stretching region and at 1167 and 747 cm⁻¹ four or more CH₂ groups.

On comparing the IR spectrum of fexofenadine with its magnesium,

manganese, iron and cobalt complexes it is found that the major doublet band of -OH group shifted to 3420 cm⁻¹ as the singlet sharp band which was due to -O-metal and N-metal stretching [22,23]. In calcium complex two very weak bands appeared at 3650 cm⁻¹ and another at 3411 cm⁻¹ as sharp band, in chromium complex as a sharp band it was at 3400 cm⁻¹, in case of manganese complex a very weak band was at 3647 cm⁻¹. In case of iron ammonium citrate this band appeared as broadband in the region 3440-3380 cm⁻¹ which was due to the N-H stretching of ammonium ion. In iron sulfate complex it was at 3400-3390 cm⁻¹ as a medium band. In nickel and copper complex a sharp stretching band appeared at 3390 cm⁻¹. Fexofenadine zinc complex showed three consecutive small singlet at 3618 and sharp doublet at 3376. In cadmium complex a small band at 3629 cm⁻¹ and 3450 cm⁻¹ was recorded. Water bands are also appeared above 3000 cm⁻¹ (Figures 5 and 6).

Fexofenadine showed C=O stretching at 1700 cm⁻¹ due to carboxylic group which shifted to 1680 cm⁻¹ as doublet strong band in magnesium





complex whereas in cadmium, calcium, chromium, manganese, iron chloride, iron sulfate, iron ammonium citrate, cobalt, nickel, copper and zinc complexes this band appeared in the region 1715-1720 cm⁻¹ [24,25] complexes indicating the coordination of this moiety to the metal ions [26] (Figure 7).

band of medium stretching, while in magnesium, calcium and cobalt complex this band appeared at 1300 cm⁻¹, in chromium it was in the form of broadband, whereas in manganese and iron chloride it was in the region of 1300-1280 cm⁻¹, iron sulfate it was at 1264 cm⁻¹ as weak band, in iron ammonium citrate it appeared at 1253 cm⁻¹ as a medium intensity band. In nickel complex at 1300 cm⁻¹ as medium band whereas

Fexofenadine C-O stretching appeared at 1279 cm⁻¹ as single

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in copper it was a doublet of weak intensity, in zinc complex a weak doublet appeared at 1248 cm⁻¹ and in cadmium complex at 1260 cm⁻¹. The change in the C-O stretching showed that the carboxylate ion also coordinated to a metal [27].

The C-N stretching in the reference compound was recorded at 1167 cm⁻¹ as medium stretching but in magnesium, chromium, manganese, ferric chloride, cobalt, copper, zinc and cadmium complex this appeared in the region 1149-1151 cm⁻¹, a sharp doublet appeared at 1100-1140 cm⁻¹ in calcium, whereas in nickel complex it was at 1120 cm⁻¹ as a medium band. In iron sulfate a doublet band was found at 1114 cm⁻¹ because of metal to nitrogen bonding and was absent in iron ammonium citrate [28-30].

The medium intensity bands of metal nitrogen absorptions were recorded in the region $1370-1395 \text{ cm}^{-1}$ as medium bands whereas in the range 670-580 cm⁻¹ as low intensity bands. From the above stretching modes of metal nitrogen stretching in fexofenadine metal complexes, there is no coordination of metal with nitrogen as there are no strong bands in these regions in the spectrum of fexofenadine [26,31].

Metal oxygen stretching was recorded at 1510-1490, weak bands of metal oxygen appeared in low intensity region in between 500-600 cm⁻¹ as sharp and weak singlets and medium doublets [24,29]. When atoms of higher atomic masses were attached to the oxygen of carboxylate ion the stretching frequency lowered and another reason is that if metal oxygen bond was weaker the stretching frequency will also lower. That's why weaker bonds are recorded at lower intensity.

The metal chloride stretching frequencies were recorded in the region 690-640 cm⁻¹ as weaker bands in magnesium, chromium and manganese but prominent bands in the rest of the complexes [22].

¹H NMR studies

The NMR spectra of fexofenadine and its metal complexes were recorded in deuterated methanol on 400 MHz Avance 500 spectrophotometer.

On comparing main peaks of fexofenadine with its complexes, it is observed that all the signals of the free ligand are present in the 1HNMR spectra of the complexes. Fexofenadine showed the aromatic proton resonating at δ 7.19 (s, J=1.16 Hz), 7.11 (s, J=0.5 Hz) and 7.06. The proton resonating at δ 2.0 is assigned to N-H proton, at δ 1.52 are assigned for two methyl on tertiary carbon. Protons of piperidine ring CH₂-N and -CH₂-CH₂-N resonate at δ 2.29, 2.19 and 2.34 (t, J=12.2 Hz), -CH₂ open chain at 2.36 (t, J=14.0 Hz), 1.39, 1.77 and 4.5 (s, J=1.0 Hz) respectively, at δ 4.95 (s) are alcoholic protons.

Two tertiary methyl groups have their chemical shift at 1.52 ppm in reference compound which shifted to 1.64 ppm in iron sulfate, cobalt and nickel complexes. The protons of methyl substituent with carboxylic group were found at 1.61 ppm in chromium complex, 1.46 ppm and 3.12 ppm in manganese complex, 1.60 ppm in copper complex, 1.63 ppm in zinc complex and 1.62 ppm in cadmium complex respectively. The protons of adjacent carbon of carboxylic group in magnesium complex shifted to 1.65 ppm, 1.64 ppm in calcium complex and in iron ammonium citrate this shift appeared at 2.8 and 2.55 ppm.

In iron sulfate, nickel, copper, zinc and cadmium complexes $-CH_2$ adjacent to piperidine nitrogen have peaks at 2.29 ppm and 2.19 ppm (t, J=2.0 Hz), (t, J=3.5 Hz) (t, J=3.75 Hz) (t, J=7.69 Hz). The methylene proton peak in magnesium complex was found at 1.94 ppm (t, J=0.99 Hz), in calcium complex at 2.92 ppm (t, J=3.5 Hz), in chromium complex at 1.91 ppm 1.88 ppm (t, J=12.2 Hz), in manganese complex at 2.27 ppm, 1.79 ppm and 1.69 ppm, in iron chloride at 2.93 ppm,

2.92 ppm, in iron sulfate 2.19 ppm, 2.92 ppm and in cobalt complex the peak was observed at 2.24 ppm (t, J=7.5 Hz). In iron ammonium citrate the methylene protons were at 4.79, 2.8, 2.55, 3.49 ppm and aromatic protons were at 6.99 and 7.07 ppm.

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The set of signals of most of the groups were almost similar except for piperidine nitrogen, its adjoining protons and a carboxylic group because both these functional groups were taking part in the coordination.

The methylene protons adjacent to carboxylic group of magnesium, calcium, chromium and manganese complex the methyl protons of tertiary carbon shifted in the region 1.46-3.12 ppm. The methylene protons of piperidine ring shifted in all these complexes which confirmed the coordination of metal with nitrogen of piperidine ring and carboxylic group as evidenced from ¹H NMR data.

At 1.42, 1.52 and 1.64 ppm shifting of methyl protons of tertiary carbon adjacent to carboxylic group were observed in fexofenadine iron chloride and iron sulfate complex, whereas there was a shift to 2.92 ppm for iron chloride and 2.19 for iron sulfate complex which confirms that the coordination of metal with nitrogen of piperidine ring and carboxylic group was evidenced from ¹H NMR data. The shifting of methylene protons adjacent to carboxylic group to 2.8 and 2.55 ppm confirmed that the coordination of metal with carboxylic group was evidenced from ¹H NMR data.

The methyl protons of tertiary carbon adjacent to carboxylic group shifted in the region 1.46-1.64 ppm and methylene protons of piperidine ring shifted to 2.24 ppm confirmed that the coordination of metal with nitrogen of piperidine ring and carboxylic group was evidences from ¹H NMR data of cobalt complex.

Spectroscopic studies as UV, IR, ¹H NMR, and CHN analysis fully supports the formation of fexofenadine metal complexes in which fexofenadine is coordinated to the metal through piperidine nitrogen, oxygen of carboxylic group and oxygen of alcoholic group (Figures 2 and 3).

Conclusion

Fexofenadine metal complexes with magnesium, calcium, chromium, manganese, iron (ferric and ferrous); cobalt nickel, copper, zinc and cadmium were synthesized and characterized by FT-IR, ¹H-NMR and elemental analysis. Fexofenadine coordinate to metal ions to form stable monodentate complexes. The coordination being through piperidine nitrogen in case of magnesium, calcium, chromium and manganese complexes and with oxygen of carboxylato group in complexation with iron, cobalt nickel, copper, zinc and cadmium.

Disclosure Statement

There are no actual or potential conflicts of interest in this paper.

References

- O'Neil, Patricia E Heckelman, Cherie B Koch, Kristin J. Roman (2006) The Merck Index. An encyclopedia of Chemicals Drugs, and Biologicals, Fourteenth, Merck & Co., Inc., NJ, USA.
- Caballero E, Ocaña I, Azanza JR, Sádaba B (1999) [Fexofenadine: a antihistaminic. Review of its practical characteristics]. Rev Med Univ Navarra 43: 93-97.
- Amon U, Amon S, Gibbs BF (2000) In vitro studies with fexofenadine, a new nonsedating histamine H1 receptor antagonist, on isolated human basophils. Inflamm Res 49 Suppl 1: S13-14.

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- Stone BM, Turner C, Mills SL, Nicholson AN (1999) Studies into the possible central effects of the H-1 receptor antagonist, fexofenadine. Int Arch Allergy Immunol 118: 338.
- Turncliff RZ, Hoffmaster KA, Kalvass JC, Pollack GM, Brouwer KL (2006) Hepatobiliary disposition of a drug/metabolite pair: Comprehensive pharmacokinetic modeling in sandwich-cultured rat hepatocytes. J Pharmacol Exp Ther 318: 881-889.
- Greenblatt DJ (2009) Analysis of drug interactions involving fruit beverages and organic anion-transporting polypeptides. J Clin Pharmacol 49: 1403-1407.
- Shimizu M, Fuse K, Okudaira K, Nishigaki R, Maeda K, et al. (2005) Contribution of OATP (organic anion-transporting polypeptide) family transporters to the hepatic uptake of fexofenadine in humans. Drug Metab Dispos 33: 1477-1481.
- Triggiani M, Gentile M, Secondo A, Granata F, Oriente A, et al. (2001) Histamine induces exocytosis and IL-6 production from human lung macrophages through interaction with H1 receptors. J Immunol 166: 4083-4091.
- Castillo O, Luque A, Román P, Lloret F, Julve M (2001) Syntheses, crystal structures, and magnetic properties of one-dimensional oxalato-bridged Co(II), Ni(II), and Cu(II) complexes with n-aminopyridine (n = 2-4) as terminal ligand. Inorg Chem 40: 5526-5535.
- Cuevas A, Kremer C, Suescun L, Russi S, Mombrú AW, et al. (2007) Synthesis, crystal structure and magnetic properties of novel heterobimetallic malonatebridged MIIReIV complexes (M = Mn, Fe, Co and Ni). Dalton Trans 7: 5305-5315.
- Sultana N, Arayne MS, Rizvi SBS, Haroon U, Mesaik MA (2012) Synthesis, spectroscopic and biological evaluation of some levofloxacin metal complexes. Medicinal Chemistry Research 22: 1371-1377.
- Sultana N, Humza E, Arayne MS, Haroon U (2011) Effect of metal ions on the in vitro availability of enoxacin, it's in vivo implications, kinetic and antibacterial studies. Quim Nova 34: 186-189.
- Arayne S, Sultana N, Haroon U, Mesaik MA (2009) Synthesis, characterization, antibacterial and anti-inflammatory activities of enoxacin metal complexes. Bioinorg Chem Appl.
- Sultana N, Arayne MS, Gul S, Shamim S (2010) Sparfloxacin–metal complexes as antifungal agents – their synthesis, characterization and antimicrobial activities Journal of Molecular Structure 975: 285-291.
- Sultana N, Arayne MS, Naz A, Mesaik MA (2010) Synthesis, characterization, antibacterial, antifungal and immunomodulating activities of gatifloxacin-metal complexes. Journal of Molecular Structure 969: 17-24.
- Arayne MS, Sultana N, Mirza AZ (2009) Preparation and spectroscopic characterization of metal complexes of gliquidone. Journal of Molecular Structure 927: 54-59.
- Sultana N, Arayne MS, Iftikhar B, Nawaz M (2008) A new RP-HPLC method for monitoring of atenolol: Application to atenolol metal interaction studies. J Chem Soc Pak 30: 113-118.

- Lingane JJ (1958) Electroanalytical Chemistry Second edition, Interscience Publishers, New York, USA.
- 19. Bailey DG (2010) Fruit juice inhibition of uptake transport: a new type of fooddrug interaction. Br J Clin Pharmacol 70: 645-655.
- 20. Ford R, Schwartz L, Dancey J, Dodd LE, Eisenhauer EA, et al. (2009) Lessons learned from independent central review. Eur J Cancer 45: 268-274.
- 21. (2003) Stability testing of new drug substances and products. International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use.
- Iwamoto R, Matsuda T (2004) Characterization of infrared and near-infrared absorptions of free alcoholic OH groups in hydrocarbon. Appl Spectrosc 58: 1001-1009.
- Kasumov VT, Medjidov AA, Yayli N, Zeren Y (2004) Spectroscopic and electrochemical characterization of di-tert-butylated sterically hindered Schiff bases and their phenoxyl radicals. Spectrochim Acta A Mol Biomol Spectrosc 60: 3037-3047.
- Korshin EE, Leitus G, Shimon LJ, Konstantinovski L, Milstein D (2008) Silanolbased pincer Pt(II) complexes: synthesis, structure, and unusual reactivity. Inorg Chem 47: 7177-7189.
- Wille U, Tan JC, Mucke EK (2008) A computational study of multicomponent orbital interactions during the cyclization of silyl, germyl, and stannyl radicals onto C-N and C-O multiple bonds. J Org Chem 73: 5821-5830.
- García-Terán JP, Castillo O, Luque A, García-Couceiro U, Román P, et al. (2004) One-dimensional oxalato-bridged Cu(II), Co(II), and Zn(II) complexes with purine and adenine as terminal ligands. Inorg Chem 43: 5761-5770.
- Garrone E, Bulánek R, Frolich K, Otero Aréan C, Rodríguez Delgado M, et al. (2006) Single and dual cation sites in zeolites: theoretical calculations and FTIR spectroscopic studies on CO adsorption on K-FER. J Phys Chem B 110: 22542-22550.
- Cossio FP, Arrieta A, Sierra MA (2008) The mechanism of the ketene-imine (staudinger) reaction in its centennial: still an unsolved problem? Acc Chem Res 41: 925-936.
- Martin R, Buchwald SL (2008) Palladium-catalyzed Suzuki-Miyaura crosscoupling reactions employing dialkylbiaryl phosphine ligands. Acc Chem Res 41: 1461-1473.
- Tye JW, Weng Z, Johns AM, Incarvito CD, Hartwig JF (2008) Copper complexes of anionic nitrogen ligands in the amidation and imidation of aryl halides. J Am Chem Soc 130: 9971-9983.
- Martínez-Lillo J, Delgado FS, Ruiz-Pérez C, Lloret F, Julve M, et al. (2007) Heterotrimetallic oxalato-bridged RelV2MII complexes (M=Mn, Co, Ni, Cu): synthesis, crystal structure, and magnetic properties. Inorg Chem 46: 3523-3530.