

Study on Resistance of *Culex pipiens* (Diptera: Culicidae) Populations to Fenitrothion in Northern Tunisia

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Abstract

Four populations of *Culex pipiens* were collected as larvae in Northern Tunisia to evaluate their resistance status against two insecticides: fenitrothion and propoxur. At LC₅₀, the sample # 1 was susceptible, whereas all the other samples were resistant. The RR₅₀ ranged from 1.08 in sample # 1 to 550 in sample # 3. The A2-B2, A4-B4 (and/or A5-B5), B12 and C1 esterases were found in collected samples and the frequencies ranged from 0.02 to 0.42. Propoxur caused a mortality of 0% in samples # 3 which showed the highest resistance levels to fenitrothion insecticide and 87% in sample # 1 which was susceptible hence the involvement of AChE 1 in the recorded resistance. Our results are essential for the development of such strategies of vector control.

Keywords: *Culex pipiens*; Fenitrothion; Propoxur; Resistance; Esterases; AChE1; Central Tunisia

Introduction

Formerly, insect populations resistant to pesticides were controlled either by increasing the quantity of product used, either by applying new active ingredients. Both strategies are now over. The use of increasing amounts of insecticides is a danger to the environment and is very costly; moreover, the discovery and development of new insecticides is clearly decreasing. There are thus few alternatives to control insects such as *Culex pipiens* which are resistant to insecticides, whether organophosphates, carbamates or pyrethroids [1-10]. Fenitrothion is one of the most popular organophosphorus (OP) insecticides used worldwide, which inhibits arthropod [11].

All of these considerations prompt urgent action, based on the development of appropriate strategies for the use of pesticides. Data inherent to insecticides, their toxicity and their interactions with arthropod action sites, to the knowledge of biochemical resistance mechanisms, are essential for the development of such strategies. It is in this context that this document is inscribed. We reported in this paper a study on fenitrothion resistance of *Culex pipiens* populations collected in four breeding sites of Northern Tunisia (Figure 1 and Table 1).

Materials and Methods

Mosquito strains

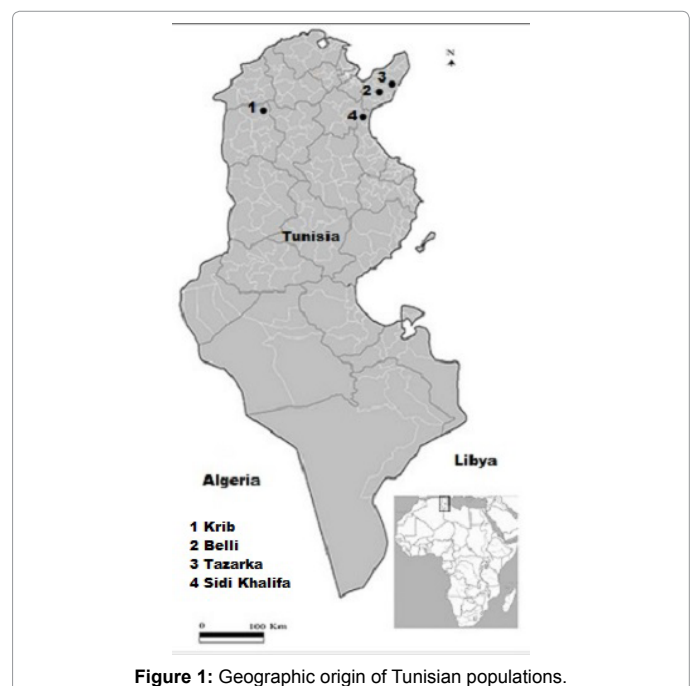
Four populations of *Culex pipiens* were collected as larvae and pupae in Northern Tunisia between August 2003 and October 2005. Collected samples were reared in the laboratory for further bioassays. Three strains were used as references: S-Lab was a sensitive strain, SA2, and SA5 were resistant strains with A2-B2 and A5-B5 esterases, respectively.

Insecticides

Assays were performed using two insecticides: fenitrothion (98.5% [AI]), brought from laboratory Dr Ehrenstorfer, Germany), and propoxur (99.9% [AI], Bayer AG, Leverkusen, Germany), organophosphates and carbamates compounds, respectively. We used two synergists in order to detect detoxification enzymes involved in resistance: S, S, S {ributyl phosphorothioate (DEF), an esterase inhibitor, and piperonyl butoxide (pb), an inhibitor of mixed function oxidases.

Bioassay procedures and data analysis

Bioassays were realized on late third and early fourth instar larvae according to procedures of WHO [12]. Results were analysed for the



median lethal concentration (LD₅₀) and LD₉₅ by probit analysis using a Basic program [13].

Esterase's detection

Esterase phenotypes were established by starch electrophoresis (TME 7.4 buffer system) as described by Pasteur et al. [14,15] using homogenates of thorax and abdomen.

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Code	Locality	Breeding site	Date of collection	Mosquito control (used insecticides)	Agricultural pest control
1	Krib	River	Oct. 2005	Occasional (P)	Yes
2	Belli	River	Aug. 2003	Rare (C,D)	Yes
3	Tazarka	River	May 2005	Very frequent (C, T, Pm, F, P, D)	Yes
4	Sidi khalifa	Water pond	July 2004	None	None

C : Chlorpyrifos ; T : Temephos ; Pm : Pirimiphos methyl ; F : Fenitrothion ; P : Permethrin ; D : Deltamethrin

Table 1: Geographic origin of Tunisian populations, breeding site characteristics and insecticide control.

Population	Fenitrothion			Fenitrothion +DEF					Fenitrothion +Pb					
	LC ₅₀ in µg/l (a)	Slope ± SE	RR ₅₀ (a)	LC ₅₀ in µg/l (a)	Slope ± SE	RR ₅₀ (a)	SR ₅₀ (a)	RSR	LC ₅₀ in µg/l (a)	Slope ± SE	RR ₅₀ (a)	SR ₅₀ (a)	RSR	
S-Lab	3.3 (1.7-6.3)	3.19 ± 0.94	-	1.3 (1.0-1.6)	2.43 ± 0.26	-	2.5 (1.2-5.2)	-	-	2.8 (0.18-44)	1.44 ± 0.93	-	1.1 (0.34-3.9)	-
Krib	3.6 (1.9-6.6)	2.36 ± 0.52	1.08 (0.47-2.4)	-	-	-	-	-	-	-	-	-	-	-
Belli	8.7 (5.0-14)	1.13 ± 0.15	2.6 (1.3-5.1)	7.5 (3.2-17)	0.94 ± 0.19	5.6 (3.7-8.4)	1.1 (0.78-1.7)	0.46	14 (9.8-20)	1.58* ± 0.2	4.9 (1.9-12.5)	0.61 (0.43-0.84)	0.52	
Tazarka	1840 (1710-1980)	6.38 ± 0.61	550 (241-1250)	1990 (1790-2220)	3.52 ± 0.25	1497 (1140-1965)	0.92 (0.71-1.1)	0.37	1040 (953-1130)	4.55 ± 0.35	360 (119-1087)	1.7 (1.3-2.3)	1.5	
Sidi khalifa	126 (74-215)	1.43 ± 0.21	37.7 (18.6-76.5)	26 (8.0-51)	0.71 ± 0.17	19.8 (14.3-27.4)	4.7 (3.1-7.2)	1.9	37 (26-54)	1.56* ± 0.24	13.0 (4.2-40.3)	3.3 (2.2-5.0)	2.9	

(a), 95% CI; * The log dose-probit mortality responses is parallel to that of S-Lab. RR₅₀, resistance ratio at LC₅₀ (RR₅₀ = LC₅₀ of the population considered / LC₅₀ of Slab); SR₅₀, synergism ratio (LC₅₀ observed in absence of synergist / LC₅₀ observed in presence of synergist). RR and SR considered significant (P<0.05) if their 95%CI did not include the value 1. RSR, relative synergism ratio (RR for insecticide alone / RR for insecticide plus synergist).

Table 2: Fenitrothion resistance characteristics of Tunisian *Culex pipiens* in presence and absence of synergists DEF and Pb.

Results

Fenitrothion resistance

The linearity of the dose-mortality response was accepted ($p < 0.05$) for S-Lab and field samples # 3. At LC₅₀, the sample # 1 was susceptible, whereas all the other samples were resistant (Table 2). The RR₅₀ ranged from 1.08 in sample # 1 to 550 in sample # 3. High resistance levels were manifested by sample # 3 (>500 folds). The samples # 1 and 2 showed low resistance levels, not exceeding 10-fold. At LC₉₅, RR₉₅>100 in samples # 3 and 4.

The addition of DEF to fenitrothion bioassays decreased significantly the tolerance in S-Lab (SR₅₀=2.5, $p < 0.05$) and sample # 4 (Table 2). The SR was not significantly higher than that recorded in S-Lab in all samples. These results indicate that the increased detoxification by the EST (and/or GST) did not play any role in the resistance. The Pb had not a significant effect on the fenitrothion resistance in S-Lab (SR₅₀ = 1.16, $p < 0.05$). The resistance decreased significantly in sample # 4, but the SR₅₀ was not significantly higher than that recorded in S-Lab in any samples (Table 2). These mechanisms did not account any portion of the fenitrothion resistance for all samples.

Cross-resistance of fenitrothion/propoxur

Propoxur caused a mortality of 0% in samples # 3 which showed the highest resistance levels to fenitrothion insecticide. The highest percentage of mortality was recorded in sample # 1 (87%) which showed a susceptibility to fenitrothion. Mortalities due to propoxur were 39% and 68% in resistant samples # 2 and 4, respectively. A strong correlation were found between mortality due to propoxur and the LC₅₀ of fenitrothion (Spearman rank correlation, $r = 0.69$ ($P < 0.01$)).

Esterase's activities

The A2-B2, A4-B4 (and/or A5-B5), B12, and C1 esterases were found in collected samples and the frequencies ranged from 0.02 to 0.42. The A1 esterase was not detected in any used sample.

Discussion

Our study on resistance of *Culex pipiens* to fenitrothion showed high levels compared to other studies on different mosquitoes in the world [16-18]. In addition to the treatments carried out in vector control, arthropod vectors are also subjected, depending on their ecology, to the insecticidal pressure resulting from agriculture or domestic uses, thus accelerating the appearance of the phenomenon and the spread of resistant alleles in the vector populations, resulting in a loss of effectiveness of the treatments.

Fenitrothion is one of the most popular organophosphorus insecticides used worldwide [11]. Intensive insecticide applications often result in accelerated biodegradation of the insecticide in the environment [19-21]. Many studies confirmed drastic increase of fenitrothion-degrading *Pseudomonas*, *Flavobacterium*, and *Burkholderia* in agricultural field soils [22-24].

Our synergist study showed that the increased detoxification by EST (and/or GST) and oxydases were not involved in the recorded resistance. Starch electrophoresis detected many esterases in all studied samples. This confirms the hypothesis that some esterases, GSTs, and cytochrome P450 enzymes may be insensitive to the action of DEF and Pb. The involvement of EST and the GST in the OPs resistance was confirmed by many previous studies [2,3,25-32]. Our study is in agreement with previous publication on correlation between cytochrome P450 enzymes and resistance to pyrethroids [33].

We also showed that the resistance to the studied OP was correlated with the propoxur resistance hence the involvement of AChE 1, where mutations changed the sensitivity of AChE, in the recorded resistance. Our results were in agreement with many previous studies that have shown the role offered by the resistant allele, Ace-1, in many areas of the world [34-37].

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References

1. Ben Cheikh H, Marrakchi M, Pasteur N (1995) Mise en évidence d'une très forte résistance au chlorpyrifos et à la perméthrine chez les moustiques *Culex pipiens* de Tunisie. *Arch. Institut Pasteur Tunis* 72: 7-12.
2. Ben Cheikh H, Haouas-Ben Ali Z, Marquine M, Pasteur N (1998) Resistance to organophosphorus and pyrethroid insecticides in *Culex pipiens* (Diptera: Culicidae) from Tunisia. *J Med Entomol* 35: 251-260.
3. Hemingway J, Hawkes NJ, McCarroll L, Ranson H (2004) The molecular basis of insecticide resistance in mosquitoes. *Insect Biochemistry and Molecular Biology* 34: 653-665.
4. Hemingway J, Hawkes N, Prapanthadara L, Jayawardenal KG, Ranson H (1998) The role of gene splicing, gene amplification and regulation in mosquito insecticide resistance. *Phil Trans R Soc Lond B Biol Sci* 353: 1695-1699.
5. Karunaratne SH, Hemingway J (2001) Malathion resistance and prevalence of the malathion carboxylesterase mechanism in populations of mosquito vectors of disease in Sri Lanka. *Bull. World Health Org* 79: 1060-1064.
6. Ranson H, Claudianos C, Ortelli F, Abgrall C, Hemingway J, et al. (2002) Evolution of supergene families associated with insecticide resistance. *Science* (Washington, DC) 298: 179-181.
7. Weill M, Lutfalla G, Mogensen K, Chandre F, Berthomieu A, et al. (2003) Insecticide resistance in mosquito vectors. *Nature* (Lond.) 423: 136-137.
8. Russel RJ, Claudianos C, Campbell PM, Home I, Sutherland TD, et al. (2004) Two major classes of target site insensitivity mutations confer resistance to organophosphate and carbamate insecticides. *Pestic Biochem Physiol* 79: 84-93.
9. Alout H, Berthomieu A, Hadjivassilis A, Weill M (2007) A new amino-acid substitution in acetylcholinesterase 1 confers insecticide resistance to *Culex pipiens* mosquitoes from Cyprus. *Insect Biochem Mol Biol* 37: 41-47.
10. Daaboub J, Ben Cheikh R, Lamari A, Ben Jha I, Feriani M, et al. (2008) Resistance to pyrethroid insecticides in *Culex pipiens* pipiens (Diptera: Culicidae) from Tunisia. *Acta Trop* 107: 30-36.
11. Stenersen J (2004). *Chemical pesticides: Mode of action and toxicology*. CRC Press, Boca Raton, Florida.
12. WHO (1963) Insecticide resistance and vector control: 13th Report of the WHO Expert Committee on Insecticides. *WHO Tech Rep Ser* 265.
13. Raymond M, Fournier D, Bergé JB, Cuany A, Bride JM, et al. (1985) Single-mosquito test to determine genotypes with an acetylcholinesterase insensitive to inhibition to propoxur insecticide. *J Am Mosq Control Assoc* 1: 425-427.
14. Pasteur N, Iseki A, Georghiou GP (1981) Genetic and biochemical studies of the highly active esterases A' and B associated with organophosphate resistance in mosquitoes of the *Culex pipiens* complex. *Biochemical Genetics* 19: 909-919.
15. Pasteur N, Pasteur G, Bonhomme F, Britton-Davidian J (1988) Practical isozyme genetics. Ellis Horwood, Chichester, UK.
16. Bracco JE, Barata JMS, Marinotti O (1999) Evaluation of insecticide resistance and biochemical mechanisms in a population of *Culex quinquefasciatus* (Diptera: Culicidae) from São Paulo, Brazil. *Mem Inst Oswaldo Cruz, Rio de Janeiro* 94: 115-120.
17. Rodriguez MM, Bisset J, Fernandez DMD, Lauzan L, Soca A (2001) Detection of insecticide resistance in *Aedes aegypti* (Diptera: Culicidae) from Cuba and Venezuela. *J. Med. Entomol* 38: 623-628.
18. Sathantriphop S, Paeporn P, Supaphathom K (2006) Detection of insecticide resistance status in *Culex quinquefasciatus* and *Aedes aegypti* to four major groups of insecticides. *Trop Biomed* 23: 97-101.
19. Felsot AS (1989) Enhanced biodegradation of insecticides in soil: Implications for agroecosystems. *Annu Rev Entomol* 34: 453-476.
20. Arbeli Z, Fuentes CL (2007) Accelerated biodegradation of pesticides: An overview of the phenomenon, its basis and possible solutions and a discussion on the tropical dimension. *Crop Prot* 26:1733-1746.
21. Singh BK, Walkera A, Wright DJ (2005) Cross-enhancement of accelerated biodegradation of organophosphorus compounds in soils: Dependence on structural similarity of compounds. *Soil Biol Biochem* 37: 1675-1682.
22. Tago K (2006) Diversity of fenitrothion-degrading bacteria in soils from distant geographical areas. *Microbes Environ* 21: 58-64.
23. Singh BK (2009) Organophosphorus-degrading bacteria: Ecology and industrial applications. *Nat Rev Microbiol* 7: 156-164.
24. Hayatsu M, Hirano M, Tokuda S (2000) Involvement of two plasmids in fenitrothion degradation by *Burkholderia* sp. strain NF100. *Appl Environ Microbiol* 66: 1737-1740.
25. Devonshire AL (1991) Role of esterases in resistance of insects to insecticides. *Biochem Soc Trans* 19: 755-759.
26. Liu H, Xu Q, Zhang L, Liu N (2005) Chlorpyrifos resistance in mosquito *Culex quinquefasciatus*. *J. Med. Entomol* 42: 815-820.
27. Huang HS, Hu NT, Yao YE, Wu CY, Chiang SW, et al. (1998) Molecular cloning and heterologous expression of a glutathione-S-transferase involved in insecticide resistance from the diamondback moth *Plutella xylostella*. *Insect Biochem Mol Biol* 28: 651-658.
28. Wei SH, Clark AG, Syvanen M (2001) Identification and cloning of a key insecticide-metabolizing glutathione -S- transferase (MdGST-6A) from a hyper insecticide-resistant strain of the house fly *Musca domestica*. *Insect Biochem Mol Biol* 31: 1145-1153.
29. Yang Y, Wu Y, Chen S, Devine GJ, Denholm I, et al. (2004). The involvement of microsomal oxidases in pyrethroid resistance in *Helicoverpa armigera* from Asia. *Insect Biochemistry and Molecular Biology* 34: 763-773.
30. Wu G, Jiang S, Miyata T (2004) Seasonal changes of methamidophos susceptibility and biochemical properties in *Plutella xylostella* (Lepidoptera: Yponomeutidae) and its parasitoid *Cotesia plutellae* (Hymenoptera: Braconidae). *Journal of Economic Entomology* 97: 1689-1698.
31. Cui F, Qu F, Cong J, Liu X, Qiao L (2007) Do mosquitoes acquire organophosphate resistance by functional changes in carboxylesterases. *FASEB J*. 21: 3584-3591.
32. Yang ML, Zhang JZ, Zhu KY, Xuan T, Liu XJ, et al. (2009) Mechanisms of organophosphate resistance in a field population of oriental migratory locust, *Locusta migratoria manilensis* (Meyen). *Archives of Insect Biochemistry and Physiology* 71: 3-15.
33. Wan-Norafikah O, Nazni WA, Lee HL, Zainol-Arifin P, Sofian-Azirun M (2010) Permethrin resistance in *Aedes aegypti* (Linnaeus) collected from Kuala Lumpur, Malaysia. *J Asia-Pacific Entomol* 13: 175-182.
34. Quistad GB, Sparks SE, Casida JE (2001) Fatty acid amide hydrolase inhibition by neurotoxic organophosphorus pesticides. *Toxicol Appl Pharmacol* 173: 48-55.
35. Boume Y, Taylor P, Radic Z, Marchot P (2003) Structural insights into ligand interactions at the acetylcholinesterase peripheral anionic site. *EMBO J*. 22: 1-12.
36. Labbé P, Berthomieu A, Berticat C, Alout H, Raymond M, et al. (2007) Independent duplications of the acetylcholinesterase gene conferring insecticide resistance in the mosquito *Culex pipiens*. *Mol Biol Evol* 24: 1056-1067.
37. Kady GA, Kamel NH, Mosleh YY, Bahgt IM (2008) Comparative toxicity of two bio-insecticides (Spinotoram and Vertemic) compared with Methomyl against *Culex pipiens* and *Anopheles multicolor*. *World Journal of Agricultural Science* 4: 198-205.