# eISSN: 09748369, www.biolmedonline.com

# Role of secondary metabolites in defense mechanisms of plants

# \*Mazid M<sup>1</sup>, Khan TA<sup>2</sup>, Mohammad F<sup>1</sup>

<sup>1</sup> Plant Physiology Division, Department of Botany, Faculty of Life Sciences, AMU, Aligarh, India. <sup>2</sup> Department of Biochemistry, Faculty of Life Sciences, AMU, Aligarh, India.

#### \*Corresponding Author: mazidmohd699@gmail.com

#### Abstract

In all natural habitats, plants are surrounded by an enormous number of potential enemies (biotic) and various kinds of abiotic environmental stress. Nearly all ecosystems contain a wide variety of bacteria, viruses, fungi, nematodes, mites, insects, mammals and other herbivorous animals, greatly responsible for heavy reduction in crop productivity. By their nature, plants protect themselves by producing some compounds called as secondary metabolites. Secondary metabolites, including terpenes, phenolics and nitrogen (N) and sulphur (S) containing compounds, defend plants against a variety of herbivores and pathogenic microorganisms as well as various kinds of abiotic stresses. This review presents an overview about some of the mechanisms by which plants protect themselves against herbivory, pathogenic microbes and various abiotic stresses as well as specific plant responses to pathogen attack, the genetic control of host-pathogen interactions.

**Keywords:** Secondary metabolites; defense mechanism; phytoalexins; terpenes; alkaloids; phenolics; cynogenic glucosides.

**Abbreviations:** GST, glucosinolate synthase transferase; SIR, systematic induced resistance; GSL, glucosinolates; GSH, glutathione; HCN, hydrogen cyanide.

#### Introduction

In natural systems, plants face a plethora of antagonists and thus posses a myriad of defense and have evolved multiple defense mechanisms by which they are able to cope with various kinds of biotic and abiotic stress (Ballhorn et al., 2009). The most significant biotic and abiotic and man-made stress factors are summarized in figure 1. Generally, it is difficult to assign a change in the physiology of metabolism of the crop to a specific stress factor as normally a complex variety of various stress factors affects the plant simultaneously. However, there are inter-connections that exist between distinct and opposing signaling response pathways for defense against pathogens and insect herbivores and there also appear to be multiple response pathways invoked, depending on the specific stress context (Agosta, 1996; Barbour et al., 1987; Bostock, 1999; Bostock et al., 2001; Hell, 1997; Thomma et al., 1998; Vijayan et al., 1998; Whittaker and Feeny, 1971; Kusnierczyk et al., 2007). Besides antimicrobial nature, some of which are performed and some of which induced by infection. There are various other modes of defense include the construction of polymeric barriers to pathogen penetration and the synthesis of enzymes that degrade pathogen cell wall (Hammond et al., 1996). In addition, plants employ specific recognition and signaling systems enabling the rapid detection of pathogen invasion and initiation of vigorous defensive responses

(Schaller *et al.*, 1996). Once infected, some plants also develop immunity to subsequent microbial attacks (Putnam and Heisey, 1983; Putnam and Tang, 1986; Bernays, 1989; Elakovich, 1987).

Plants produce a high diversity of natural products or secondary metabolites with a prominent function in the protection against predators and microbial pathogens on the basis of their toxic nature and repellence to herbivores and microbes and some of which also involved in defense against abiotic stress (e.g. UV-B exposure) and also important for the communication of the plants with other organisms (Schafer et al., 2009), and are insignificant for growth and developmental processes (Rosenthal et al., 1991). There are three major groups of secondary metabolites viz terpenes, phenolics and N and S containing compounds. Terpenes composed of 5-C isopentanoid units, are toxins and feeding deterrents to many herbivores. Phenolics synthesized primarily from products of the shikimic acid pathway, have several important defensive role in the plants. Members of the third major group i.e. N and S containing compounds are synthesized principally from common amino acids (Rosenthal et al., 1992; Van Etten et al., 2001). Recent in vitro experiments using plants whose secondary metabolites expression has been altered by modern molecular methods have to confirm their defensive roles (Mes et al., 2000; Mansfield, 2000). Although the situation is still

unclear, it is believed that most of the 100,000 known secondary metabolites are to be involved in plant chemical defense systems, which are formed throughout the millions of years during which plants have co-existed with their attackers (Wink, 1999). Although higher concentrations of secondary metabolites might result in a more resistant plant, the production of secondary metabolites is thought to be and reduces plant growth and costly reproduction (Simms, 1992; Karban and Baldwin, 1999; Simens et al., 2002). The cost of defense has also been invoked to explain why plants have evolved induced defense, where concentrations generally increase only in stress situations (Harvell and Tollrian, 1999).

During the last several years, it has been discovered that hundreds of compounds that plants make have significant ecological and chemical defensive roles, opening a new area of scientific endeavour, often called ecological biochemistry (Harborne, 1988, 1989).

## Secondary metabolites

Plants produce a large and diverse array of organic compounds that appear to have no direct functions in growth and development i.e. they have no generally recognised roles in the process of photosynthesis, respiration, solute transport, translocation, nutrient assimilation and differentiation (Hartmann, 1991). They have a very restricted distribution than primary metabolites in the whole plant kingdom i.e. they are often found only in one plant species or a taxonomically related group of species. High concentrations of secondary metabolites might result in a more resistant plant. Their production is thought to be costly and reduces plant growth and reproduction (Simms, 1992; Karban and Baldwin, 1997; Harvell and Tollrian, 1999; Stotz et al., 1999; Siemens et al., 2002). Therefore, defense metabolites can be divided in to constitutive substances, also called prohibitins or phytoanticipins and induced metabolites formed in response to an infection involving de novo enzyme synthesis, known as phytoalexins (Van Etten et al., 1994; Grayer and Harborne, 1994). Phytoanticipins are high energy and carbon consuming and exhibit fitness cost under natural conditions (Mauricio, 1998), but recognized as the first line of chemical defense that potential pathogens have to overcome. In contrast, phytoalexin production may take two or three days, as by definition first the enzyme system needs to be synthesized (Grayer and Harborne, 1994).

### Principal groups

Plant secondary metabolites can be divided into three chemically distinct groups viz: Terpenes, Phenolics, N and S containing compounds.

# (i) Terpenes

Terpenes constitute the largest class of secondary metabolites and are united by their common biosynthetic origin from acetyl-coA or glycolytic intermediates (Gerhenzon *et al.*, 1991; Grayson, 1998; Fraga, 1988; Croteas, 1988; Loomis and Croteas, 1980; Robinson, 1980). A vast majority of the different terpenes structures produced by plants as secondary metabolites that are presumed to be involved in defense as toxins and feeding deterrents to a large number of plant feeding insects and mammals (Gershenzon and Croteau, 1991). Below, several examples will draw from the 5 major subclasses:

(a) *Monoterpenes* ( $C_{10}$ ): Many derivatives are important agents of insect toxicity. For example, the pyrethroids (monoterpenes esters) occur in the leaves and flowers of *Chrysanthemum* species show strong insecticidal responses (neurotoxin) to insects like beetles, wasps, moths, bees, etc. and a popular ingredient in commercial insecticides because of low persistence in the environment and low mammalian toxicity (Turlings *et al.*, 1995).

In Gymnosperms (conifers) like Pine and Fir, monoterpenes accumulate in resin ducts found in the needles, twings and trunks mainly as  $\alpha$ -pinene,  $\beta$ -pinene, limonene and myrecene, all are toxic to numerous insects including bark beetles, serious pest of conifer species throughout the world (Turlings *et al.*, 1995).

(b) Sesquiterpenes  $(C_{15})$ : A number of sesquiterpenes have been till now reported for their role in plant defense such as costunolides are antiherbivore agents of family composite characterized by a five membered lactone ring (a cyclic ester) and have strong feeding repellence to many herbivorous insects and mammals (Picman, 1986).

ABA is also a sesquiterpene plays primarily regulatory roles in the initiation and maintenance of seed and bud dormancy and plants response to water stress by modifying the membrane properties (Van-steveninck, 1983) and act as a transcriptional activator (McCarty *et al.*, 1991; Giraudat *et al.*, 1992). In addition, it increases the cytosolic calcium concentration and causes alkalinisation of the cytosol (Irving *et al.*, 1992; Blatt and Armstrong, 1993; Thiel *et al.*, 1992). The level of UV-B absorbing flavonols, quercetin and kaempferol were significantly increased when ABA was applied. The concentration of two hydroxy-cinnamic acids, caeffic and ferulic acids were also increased by ABA. All of above changes in the protective compounds, anti-oxidant enzymatic activities and sterols were correlated with lessened membrane harm by UV-B. Thus, defense system of plants against UV-B is activated in which ABA acts downstream in the signaling pathway (Berli *et al.*, 2010).

(c) Diterpenes  $(C_{20})$ : Abietic acid is a diterpene found in pines and leguminous trees. It is present in or along with resins in resin canals of the tree trunk. When these canals are pierced by feeding insects, the outflow of resin may physically block feeding and serve as a chemical deterrent to continued predation (Bardley et al., 1992). Another compound phorbol (diterpene ester), found in plants of Euphorbiaceae and work as skin irritants and internal toxins to mammals. Moreover, phytol a highly hydrophobic 20-C alcohol found in chlorophyll as a side chain help to anchor certain molecules in membranes and therefore increase the efficiency of chlorophyll during the photosynthesis (Knaff, 1991), a strategy for maximum  $CO_2$ fixation and biomass production (Jagendorf, 1967). Furthermore, gibberellins, a group of plant hormones are also diterpenes, that play various detrimental roles in numerous plant developmental processes such as seed germination, leaf expansion, flower and fruit set (Davies, 1995), dry mass and bio mass production (Gupta et al., 2001), stomatal conductance (Bishnoi et al., 1992), CO<sub>2</sub> fixation, phloem loading, assimilate translocation (Ouzounidou et al., 2005) and also known to exert their numerous physiological effects via specific enzymes, the synthesis of which they induce by influencing the basic process of translocation and transcription (Hutty et al., 1995).

(d) Triterpenes (C<sub>30</sub>): Several steroid alcohols (sterols) are important component of plant cell membranes, especially in the plasma membrane as regulatory channels and maintain permeability to small molecules by decreasing the motion of the fatty acid chains. The milkweeds produce several better tasting glucosides (sterols) that protect them against herbivory by most insects and even cattle (Lewis and Elvin-Lewis, 1977). Phytoecdysones have some defensive role against insects by disrupting moulting and developmental and other physiological processes with lethal consequences (Heftmann, 1975; Slama, 1979, 1980). Another triterpene, limnoid, a group of bitter substances in citrus fruits and act as antiherbivore compounds in members of family Rutaceae and some other families also. For example, Azadirechtin, a complex limnoid from *Azadirachta indica*, acts as a feeding deterrent to some insects and exerts various toxic effects (Mordue and Blackwell, 1993).

(e) Polyterpenes  $(C_5)_n$ : Several high molecular weight polyterpenes occur in plants. Larger terpenes include the tetraterpenes and the polyterpenes. The principal tetraterpenes are carotenoids family of pigments. Other one is rubber, a polymer containing 1500-15000 isopentenyl units, in which nearly all the C-C double bonds have a cis (Z) configuration while in gutta rubber has its double bond in trans (E) configuration. Rubber found in long vessels called laticifers, provide protection as a mechanism for wound healing and as a defense against herbivores (Eisner *et al.*, 1995; Klein, 1987).

# (ii) Phenolic compounds

Plants produce a large variety of secondary products that contain a phenol group, a hydroxyl functional group on an aromatic ring called Phenol, a chemically heterogeneous group also. They could be an important part of the plants defense system against pests and diseases including root parasitic nematodes (Wuyts *et al.*, 2006). Elevated ozone (mean 32.4ppb) increased the total phenolic content of leaves and had minor effects on the concentration of individual compounds (Saviranta *et al.*, 2010).

(a) *Coumarin:* They are simple phenolic compounds, widespread in vascular plants and appear to function in different capacities in various plant defense mechanisms against insect herbivores and fungi. They derived from the shikimic acid pathway (Murray *et al.*, 1982), common in bacteria, fungi and plants but absent in animals. Also, they are a highly active group of molecules with a wide range of anti-microbial activity against both fungi and bacteria (Brooker *et al.*, 2008). It is believed that these cyclic compounds behave as natural pesticidal defense compounds for plants and they represent a starting point for the exploration of new derivatives possessing a range of improved antifungal activity (figure 2).

Halogenated coumarin derivatives work very effectively *in vitro* to inhibit fungal growth. For example, 7-hydroxylated simple coumarins may play a defensive role against parasitism of *Orobanche cernua*, by preventing successful germination, penetration and connection to the host vascular system (Serghini *et al.*, 2001). Some coumarin derivatives have higher anti-fungal activity against a range of soil borne plant pathogenic fungi and exhibit more stability as compared to the original coumarin compounds alone (Brooker *et al.*, 2008).

(b) Furano-coumarins: Also a type of coumarin with special interest of phyto-toxicity, abundant in members of the family Umbelliferae including celery parsnip and parsley. Normally, these compounds are not toxic, until they are activated by light (UV-A), causes some furanocoumarins to become activated to a high energy electronic state, which can insert themselves into the double helix of DNA and bind to the pyramidine bases and thus blocking transcription and repair and eventually leading to cell death (Rice, 1987). Psoralin, a basic linear furacoumarin, known for its use in the treatment of fungal defense and found very rarely in SO<sub>2</sub> treated plants (Ali et al., 2008) (figure 3).

(c) Ligin: It is a highly branched polymer of phenyl-propanoid groups, formed from three different alcohols viz., coniferyl, coumaryl and sinapyl which oxidized to free radicals (ROS) by a ubiquitous plant enzyme-peroxidase, reacts simultaneously and randomly to form lignin. The reactive proportions of the three monomeric units in lignin vary among species. plant organs and even layers of a single cell wall (Lewis and Yamamoto, 1990). Its physical toughness deters feeding by herbivorous animals and its chemical durability makes it relatively indigestible to herbivores and insects pathogens (Mader and Amberg-Fisher, 1982). Lignifications block the growth of pathogens and are a frequent response to infection or wounding (Gould, 1983).

(d) *Flavonoids:* One of the largest classes of plant phenolic, perform very different functions in plant system including pigmentation and defense (Kondo *et al.*, 1992). Two other major groups of flavonoids found in flowers are flavones and flavonols function to protect cells from UV-B radiation because they accumulate in epidermal layers of leaves and stems and absorb light strongly in the UV-B region while letting visible (PAR) wavelengths throughout uninterrupted (Lake *et al.*, 2009). In addition, exposure of plants to increased UV-B light has been demonstrated to increase the synthesis of flavones and flavonols suggesting that flavonoids may offer a measure of protection

by screening out harmful UV-B radiation (Caldwell *et al.*, 1983; Saviranta *et al.*, 2010).

(e) Isoflavonoids: Isoflavonoids are derived from a flavonone intermediate, naringenin, ubiquitously present in plants and play a critical role in plant developmental and defense response. They secreted by the legumes and play an important role in promoting the formation of nitrogen-fixing nodules by symbiotic rhizobia (Sreevidya et al., 2006). Moreover, it seems that synthesis of these flavonoids is an effective strategy against reactive oxygen species (ROS) (Posmyk et al., 2009). The analysis of activity of antioxidant enzymes like SOD, CAT, POX, APX. GPX and GR suggested that peroxidases were the most active enzymes in red cabbage seedlings exposed to Cu<sup>2+</sup> stress. It could result from the fact that phenolic compounds (Phc), which could be also substrate for different peroxidases were the first line of defense against various environmental stress like metal stress (Posmyk et al., 2009; Novak et al., 2009) (figure 3).

(f) Tanins: It included under the second category of plant phenolic polymers with defensive properties. Most tannins have molecular masses between 600 and 3000. Tannins are general toxins that significantly reduce the growth and survivorship of many herbivores and also act as feeding repellents to a great diversity of animals. In mammalian herbivores, they cause a sharp, astringent sensation in the mouth as a result of their binding of salivary proteins. Mammals such as cattle, deer and apes, characteristically avoid plant with high tannin contents (Oates et al., 1980). The defensive properties of tannins are generally attributed to their ability to bind proteins. Protocatechllic and chlorogenic acids probably have a special function in disease resistance of certain plants. They prevent smudge in onions, a disease caused by the fungus Colletotrichum circinans and prevent spore germination and growth of other fungi as well (Vickery, 1981; Butt and Lamb, 1981; Mayer, 1987). It is thought by some that chlorogenic acid and certain other related compounds can be readily formed and oxidised into potent fungistatic quinones by certain disease resistant cultivars but less readily so by susceptible ones.

# (iii) Sulphur containing secondary metabolites

They include GSH, GSL, phytoalexins, thionins, defensins and allinin which have

been linked directly or indirectly with the defense of plants against microbial pathogens (Hell, 1997; Crawford *et al.*, 2000; Leustek *et al.*, 2000; Saito, 2004; Grubb and Abel, 2006; Halkier and Gershenzon, 2006), and a number of them thought to be involved in the SIR (ElkeBloem *et al.*, 2005).

(a) GSH: It is one of the major forms of organic S in the soluble fraction of plants and has an important role as a mobile pool of reduced S in regulation of plant growth the and development, and as an cellular anti-oxidant in stress responses (Kang et al., 2007; Noctor et al., 1998), reported as a signal of plant S sufficiency that down regulates S-assimilation and S-uptake by roots (Lappartient and Touraine, 1997; Lappartient et al., 1999). Specialized cells such as Trichomes exhibit high activities of enzymes for synthesis of GSH and other phytochelatins necessary for detoxification of heavy metals (Gutierez-Alcala et al., 2000; Choi et al., 2001). The GSH content varies between 3 to 10 mM and is present in the major cellular compartments of the plant (Leustek and Saito, 1999). To mitigate oxidative stress, GSH functions as a direct anti oxidant and also as a reducing agent also for other anti oxidants such as ascorbic acid (Nocito et al., 2002) as well as an integral weapon in the defense against ROS generated by O<sub>3</sub> (Pasqualini et al., 2002; Conklin et al., 2004) or as a reaction to biotic and abiotic stress.

Additionally, GSH is also involved in the detoxification of xenobiotics and cytotoxins by targeting them in to vacuole (Rea *et al.*, 1998). GSH is rapidly accumulated after fungal attack, may act as systemic messenger carrying information concerning the attack to non-infested tissues (Foyer and Rennenberg, 2000; Edwards *et al.*, 1991).

(b) GSL: A group of low molecular mass N and S containing plant glucosides that produced by higher plants in order to increase their resistance against the unfavourable effects of predators, competitors and parasites because their break down products are release as volatiles defensive substances exhibiting toxic or repellent effects (Mithen, 1992; Wallsgrove et al., 1999; De Vos et al., 2009), for example, mustard oil glucosides in cruciferae and allyl cys sulfoxides in allium (Leustek, 2002). The smelling volatiles from GSL catalyzed by myrosinase, cleave glucose from its bond with the S atom. The resulting aglycon rearranges with loss of the sulphate to give pungent and chemically reactive products, including nitriles, function in isothiocyanates and defense as herbivorous toxins and feeding repellent (Renwick, 1992; Grubb and Abel, 2006; Halkier and Gershenzon, 2006; Talalay and Fahey, 2001). This became obvious in 1986 when the switch from single low to double low oil seed rape varieties led to increasing infestation of oil seed rape with fungal diseases such as light leaf spot (Pyrenopeziza brassicae), sclerotina stem rot (Sclerotinia sclerotiorum) and Alterneria (Alternaria brassicae). This phenomenon was attributed drastically to the reduced glucosinolate content in double low varieties (Dornberger et al., 1975; Koch, 1989; Mithen et al., 1986; Pedras and Sorensen, 1998). The potency of GSL arises when the plant tissue is damaged and GSL come in to contact with the plant enzyme myrosinase removes the βglucose moiety leading to formation of an unstable intermediates i.e. isothiocyanates (R-N=C=S) and nitriles function in defense as herbivore toxins and feeding repellents (Geu-Flores et al., 2009; Poultron and Moller, 1993; Ratzka et al., 2002; Zukalova and Vask, 2002). They are metabolized and absorbed as isothiocyanates that can affect the activity of enzymes involved both in the antioxidant defense system and in the detoxification from xenobiotics and significantly affect GST activity and cell protection against DNA damage (Lipka et al., 2010; Porrini, 2008) whereas toxicity of glucosinolatic products is well documented but their mode of action has not been elucidated and results from vet experiments with Brassica plants modified in GSL content generated doubts about their contribution to plant defense. Studies of Mithen and Magrath (1992) were able to show that the level of alkenyl GSL within the leaves of a Brassica line that was resistant to Leptosphearia maculans was always consistently higher than that of a susceptible line but they could not find a correlation between the level of alkenyl GSL and disease resistance. Lazzeri et al. (1993) demonstrated the action of several isothiocyanates against the nematode Heterodera schachti. Pedras and Sorensen (1998) discovered that the germination and radial growth of spores of a virulent Phoma lingam pathotype was inhibited hiaher concentrations of different bv isothiocyanates. In studies of crosses of Brassica lines with different glucosinolate level resistance to fungal attacks failed to correlate with high and low glucosinolate level (Mithen and Magrath, 1992; Giamoustaris and Mithen, 1995; Wretblad and Dixelius, 2000).

The presence of GSL may help explain the characteristic patchy distribution of most cruciferous plants. This result challenges one end of the continuum of the long-standing plant apparency hypothesis, which essentially states the opposite causation, that low molecular weight toxins like GSL are evolutionary responses of patchy distribution and correlated life history traits necessary for better productivity (Siemens *et al.*, 2009).

(c) Phytoalexins: Phytoalexins are synthesized in response to bacterial or fungal infection or other forms of stress that help in limiting the spread of the invading pathogens by accumulating around the site of infection. appears to be a common mechanism of resistance to pathogenic microbes in a wide range of plants (Van Etten et al., 1994: Graver and Harborne, 1994; Bailey and Mansfield, 1982; Darvill and Albersheim, 1984). Many of these changes are linked to a rapid apoptotic response, resulting in the death of one or a few invaded plant cells, known as the hypersensitive response (HR) (Hammond-Kosack and Jones, 1996). Most plant families produce organic phytoalexins of diverse chemistry; these groups are often associated with a family, for example sesquiterpenoids of Solanaceae, isoflavonoids of Leguminosae, while phytoalexins from Brassica have an indole or related ring system and one S atom as common structural features. Crucifereae appears to be the only plant family producing these S metabolites (Gross, 1993; Pedras et al., 1997, 1998), which are clearly different from the other well-known GSL (Harborne, 1999). Cruciferous crops are cultivated worldwide because they are extremely valuable and for the last decades, various research groups have investigated cruciferous phytoalexins (Harborne, 1999; Gross et al., 1994; Monde et al., 2000) as well as their biological activity (Mehta *et al.*, 1995). Typically, there are multiple responses involving several related derivatives such as up to nine wyerone (Furano-acetylenic derivatives) forms in Vicia fava and several forms of phaseollin in Phaseolus vulgaris and glyceollins in Glycine max, pistin in Pisum sativum pods, ipomeamarone in sweet potato, orchinol in orchid tubers, trifolirhizin in red clover (Keen and Kennedy, 1974).

(d) *Defensins, thionins and lectins:* All these are S-rich non-storage plant proteins synthesize and accumulate after microbial attack and such related situations (Van Loon *et al.*, 1994). All of which inhibits the growth of a broad range of fungi (Thomma *et al.*, 2002). Some defensins are antifungal or occasionally anti-bacterial activity (Thomma *et al.*, 2002). Additionally defensins genes are partly pathogen-inducible (Chiang and Hadwiger, 1991; Gu et al., 1992) and others that are involved in resistance can be expressed constitutively (Terras et al., 1995; Parashina et al., 2000). The components seem to be involved in the natural defense system of plants as they can be highly toxic to microrganisms, insects and mammals. Accumulation of thionins in the cell wall of infected wheat spikes of resistant wheat cultivars indicating that the accumulation of thionins may be involved in defense responses to infections and in spreading of Fusarium culmorum (Kang and Buchenauer, 2003). Some plant species produce lectins as defensive proteins that bind to carbohydrate or carbohydrate containing proteins. After being ingested by herbivores, lectins bind to epithelial cell lining of the digestive tracts and interfere with nutrient absorption (Peumans and Van Damme, 1995) (figure 2).

# (iv) Nitrogen containing secondary metabolites

They include alkaloids, cyanogenic glucosides, and non-protein amino acids. Most of them are biosynthesized from common amino acids. All are of considerable interest because of their role in the anti herbivore defense and toxicity to humans.

(a) *Alkaloids:* A large family of N containing secondary metabolites found in approximately 20% of the species of vascular plants (Hegnauer *et al.*, 1988), most frequently in the herbaceous dicot and relatively a few in monocots and gymnosperms. Generally, most of them, including the pyrrolizidine alkaloids (PAs) are toxic to some degree and appear to serve primarily in defense against microbial infection and herbivoral attack. They are usually synthesized from one of the few common amino acids, in particular, aspartic acid, lysine, tyrosine and tryptophan (Pearce *et al.*, 1991).

Now most alkaloids are believed to function as defensive elements against predators, especially mammals because of their general toxicity and deterrence capability (Robinson, 1980; Harborne, 1988; Hartmann et al., 1991). Large number of livestocks death is caused by the ingestion of alkaloids containing plants. In US, a significant % of all grazing livestock are poisoned each year by consumption of large quantities of alkaloid containing plants such as lupines (Lupinus) and larkspur (Delphinium) (Keeler, 1975). On a cellular level, the mode of action of alkaloids in animals is quite variable. Some interfere with components of the nervous system, especially the chemical transmitters, other affect

membrane transport, protein synthesis and miscellaneous enzyme activities (Creelman and Mullet, 1997) (figure 2).

(b) *Cyanogenic glucosides:* They constitute a group of N-containing protective compounds other than alkaloids, release the poison HCN and usually occur in members of families viz., Graminae, Rosaceae and Leguminosae (Seigler, 1991). They are not in themselves toxic but are readily broken down to give off volatile poisonous substances like HCN and  $H_2S$  when the plant is crushed; their presence deters feeding by insects and other herbivores such as snails and slugs (Taize and Zeiger, 1995).

Amygdalin, the common cyanogenic glucoside found in the seeds of almonds, apricot, cherries and peaches while Dhurrin, found in Sorghum bicolar. Normally, both are not broken down in the intact plant because the glucosides and degradative enzymes are separated in different compartments (Poulton, 1990). Under ordinary conditions, this compartmentalization prevents decomposition, however, on damaging as during herbivore feeding, the cell contents of different tissues mix and form HCN, a toxin of cellular respiration by binding to the Fe-containing heme group of cytochrome oxidase and other respiratory enzymes. Similarly, the presence of cyanogenic glucosides in cassava, make it suitable for long time storage without being attacked by pests (Pearce et al., 1991). Lima bean (Phaseolus lunatus L.) is a model plant for studies of inducible indirect anti herbivore defences including the production of volatile organic compounds (VOCs) (Ballhorn et al., 2009). The cyanogenic potential (HCNp; concentration of cyanogenic glucosides) as a crucial parameter determining Lima bean cyanogenesis and quantitative variability of cyanogenesis in natural population of wild Lima bean in Mexico, was significantly correlated with missing leaf area and therefore, cyanogenesis has to be considered as an important direct defensive trait affecting Lima beans' overall defence in nature (Ballhorn et al., 2009).

(c) *Non-protein amino acids:* Many plants also contain unusual amino acids called non-protein amino acids that incorporated into proteins but are present as free forms and act as protective defensive substances (Johnson *et al.*, 1989). For examples, canavanine and azetidine-2-carboxylic acid are close analogs of arginine and proline respectively. They exert their toxicity in various ways. Some block the synthesis of or uptake of protein amino acid

while others can be mistakenly incorporated in to proteins. After ingestion, canavanine is recognized by herbivore enzyme that normally binds arginine to the arginine transfer RNA molecule and so become incorporated in to proteins in place of arginine. The usual result is a non-functional proteins because either its tertiary structure or it catalytic site is disrupted (Rosenthal, 1991). Plants that synthesize nonprotein amino acids are not susceptible to the toxicity of these compounds but gain defense herbivorous animals. insects to and pathogenic microbes. Also, a number of plants including Arabidopsis uses Arginine as a storage and transport form of N and proline as a compatible solute in the defense against abiotic stresses causing water deprivation (Funck et al., 2009) (figure 3).

# Recognition of some pathogenic substances necessary to initiate defense responses

Within a species, individual plants often differ greatly in their resistance to microbial pathogens. These differences often lay in the speed and intensity of plants reactions (Shulaev *et al.*, 1997; Ryals *et al.*, 1996). Resistant plants respond to pathogens than susceptible plants. Hence, it is important to learn how plants sense the presence of pathogens and initiate defense.

Until in the last few years, researchers have isolated numerous plant resistance genes, recognised as R genes, function in defense against fungi, bacteria and nematodes. Most of the R-genes are thought to encode receptors that recognise and bind specific molecules originating from pathogens and alert the plant to the pathogens presence. The specific pathogen molecules recognised are referred to as elicitors include proteins, peptides, lipids etc. arising from the pathogen wall, the outer membrane or a secretion process (Bollar, 1995) can induce phytoalexins production and activate other defense reactions (Ebel, 1986; Boller, 1989). Some are polysaccharides produced when pathogenic fungi and bacteria attack on the plant cell wall (Templeton and lams, 1988; Bollers, 1989; Stone, 1989) while others are oligosaccharides produced by degradation of fungal cell wall by plant enzymes that the fungus causes the plants to secrete. Apparently, such exogenous elicitors are recognized by certain proteins in membrane, which then signal increases transcription of mRNA molecules that code for enzymes that synthesize the phytoalexins (figure 4). For example, in case of glyceollin production by infected soya bean roots, the signal may be calcium. The hypersensitive

response (HR) of cell death and pathogenesis related (PR) gene expression of these gene induced by various elicitors which induced HR and activation of the genes. Some protein elicitors are boehmerin, harpin or INF1 (Zhang *et al.*, 2009). All R-gene products themselves are nearly proteins with a leucine rich domain that is repeated in exactly several times in the amino acids sequence (Hammond-Kosack and Jones, 1997). Similarly,  $\beta$  amino-butyric acid can induce disease resistance in *Arabidopsis* against the fungal pathogen *Hyaloperonospora arabidopsis* (Vander Ent *et al.*, 2009) and bacterial pathogen (*Pseudomonas syringae*).

#### Conclusion and future prospects

Plants have evolved multiple defense mechanisms against microbial pathogens and various types of environmental stress. Besides anti-microbial secondary metabolite, some of which are performed and some of which are induced by infection. Today, advanced tools are demanded to investigate the correct correlation between N and S fertilization and crop resistance management. In a number of previous research articles and review papers, it have shown that the N and S containing secondary metabolites are influenced by optimum supply of N and S and their good nutrition can enhance the capability of a plant to cope with biotic and abiotic stress. The identification of the mechanisms causing SIR will be an important milestone for sustainable agricultural production, as the use of funaicides could then be minimized or eliminated. Thus, SIR may become an important strategy for efficiently combating with pathogens in organic forming system. Therefore, additional research in area of natural pesticides development is needed in current scenario. In the long term, it will probably be possible to generate gene cassettes for complete pathways, which could then be used for production of valuable secondary defensive metabolites in bioreactors or for metabolic engineering of crop plants. This will improve their resistance against herbivores and microbial pathogens as well as various environmental stresses.

#### Acknowledgement

The authors are highly thankful for the facilities provided at AMU, Aligarh. Financial support from the Department of Science and Technology, New Delhi in the form of project (SR/FT/LS-087/2007) is gratefully acknowledged.

#### References

Agosta W, 1996. Bombardier beetle and fever trees: A close up look at chemical warfare and signals in animals and plants. Addison-Wesley, Reading, MA.

Ali ST, Mahmooduzzafar-Abdin MZ, Iqbal M, 2008. Ontogenetic changes in foliar features and psoralen content of *Psoralea corylifolia* Linn. exposed to SO<sub>2</sub> stress. Journal of Environmental Biology, 29(5): 661-668.

Bailey JA, Mansfield JW, 1982. Phytoalexins. Wiley, New York.

Ballhorn DJ, Kautz S, Heil M, Hegeman AD, 2009. Cyanogenesis of wild lima bean (*Phaseolus lunatus* L.) is an efficient direct defence in nature. Plant Signaling and Behavior, 4(8): 735-745.

Barbour MG, Burk JH, Pitts WD, 1987. Terrestrial plant ecology, Second edition, Benjamin/Cummings, Menlo Park, Calif.

Berli FJ, Moreno D, Piccolo P, Hespanhol-Viana L, Silva MF, Bressan-Smith R, cavarnaro JB, Bottini R, 2010. Abscisis acid is involved in the response of grape (Vitis vinifera L.) cv. Malbec leaf tissues to ultraviolet-B radiation by enhancing ultravioletabsobing compounds, antioxidant enzymes and membrane sterols. Plant Cell & Environment, 33(1): 1-10.

Bernays EA, 1989. Insect-plant interaction CRC Press, Boca Raton.

Bishnoi NR, Krishnamoorthy HN, 1992. Effect of waterlogging and gibberellic acid on leaf exchange in peanut (Arachis hypogaea L.). Journal of Plant Physiology, 139: 503-505.

Blatt MR, Armstrong F, 1993. Potassium channels of stomatal guard cells: Abscisic acid-evoked control of the outward rectifier mediated by cytoplasmic pH. Planta, 191: 330-341.

Bloem E, Haneklaus S, Schnug E, 2005. Significance of sulphur compounds in the protection of plants against pests and diseases. Journal of Plant Nutrition, 28: 763-784.

Boller T, 1989. Primary signals and second messengers in the reaction of plants to pathogens. Boss WF, Morre DJ, Second messengers in plant growth and development, (eds) Alan R Liss, New York, pp: 227-255.

Boller T, 1995. Chemoperception of microbial signals in plant cells. Annual Review of Plant Physiology and Plant Molecular Biology, 46: 189-214.

Bostock RM, 1999. Signal conflicts and synergies in induced resistance to multiple attackers. Physiological and Molecular Plant Pathology, 55: 99-109.

Bostock RM, Karban R, Thaler JS, Weyman PD, Gilchrist D, 2001. Signal interactions in induced resistance to pathogens and insect herbivores. European Journal of Plant Pathology, 107: 103-111.

Bradley DJ, Kjellborn P, Lamb CJ, 1992. Elicitor and wound induced oxidative cross linking of a proline rich plant cell protein: A novel rapid defence response. Cell, 70: 21-30.

Brooker N, Windorski J, Blumi E, 2008. Halogenated coumarins derivatives as novel seed protectants. Communication in Agriculture and Applied Biological Sciences, 73(2): 81-89.

Butt VS, Lamb CJ, 1981. Oxygenase and the metabolism of plant products. Stumpf PK, Conn EE, The Biochemistry of plants, Vol 7. Secondary Plant products. (eds) Academic Press, New York, pp: 627-665.

Caldwell MM, Robberechts R, Flint SD, 1983. Internal filters: prospects for UV-acclimation in higher plants. Physiologia Plantarum, 58: 445-450.

Chiang CC, Hadwiger LA, 1991. The *Fusarium solani*-induced expression of a pea gene family encoding high cysteine content proteins. Molecular Plant Microbe Interaction, 4: 324–331.

Choi YE, Harada E, Wada M, Tsuboi H, Morita Y, Kusano T, Sano H, 2001. Detoxification of cadmium in tobacco plants: formation and active excretion of crystals containing cadmium and calcium through trichomes. Planta, 213: 45–50.

Crawford NM, Kahn ML, Leustek T, Long SR, 2000. Nitrogen and sulfur. In Biochemistry and Molecular Biology of Plants, Buchanan BB, Gruissem W, Jones RL, eds, (Rockville, MD: American Society of Plant Biologists), pp: 824–849.

Creelman RA, Mullet JE, 1997. Biosynthesis and action of jasmonates in plants. Annual Review of Plant Physiology and Plant Molecular Biology, 48: 355-381.

Croteau RB, 1988. Metabolism of plant monoterpenes. ISI Atlas of Science. Biochemistry, 1: 182-187.

Darvill AG, Albersheim P, 1984. Phytoalexins and their alicitors-a defence against microbial infection in plants. Annual Review of Plant Physiology, 35: 243-275.

Davies PJ, 1995. The plant hormone concept: Concentration, sensitivity and transport. In plant hormones: physiology, biochemistry and molecular biology, P.J. Davies, ed., Kluwer, Boston, pp: 13-38.

DeVos M, Jander G, 2009. Myzus persicae (Green peach aphid) salivary components induce defence responses in *Arabidopsis thaliana*. Plant Cell & Environment, 32(11): 1548-1560.

Dornberger K, Boeckel V, Heyer J, Schoenfeld C, Tonew M, Tonew E, 1975. Untersuchungen ueber die Isothiocyanate Erysolin und Sulforaphan aus *Cardaria draba* L. (Investigations about the isothiocyanate erysolin and sulforaphan from *Cardaria draba* L.). Pharmazie, 30(12): 792–796.

Ebel J, 1986. Phytoalexin synthesis: The biochemical analysis of the induction process. Annual Review of Phytopathology, 24: 235-264.

Edwards R, Blount JW, Dixon RA, 1991. Glutathione and elicitation of the phytoalexin response in legume cultures. Planta, 184: 403–409.

Eisner T, Meinwald J, 1995. Chemical ecology: The chemistry of biotic interaction. Eds, National Academy Press, Washington, DC.

Elakovich SD, 1987. Sesquiterpenes as phytoalexins and allelopathic agents. (eds) Ecology and metabolism of plant lipids. Fuller G and Nes WD. American chemical society, Washington, D.C. pp: 93-108.

Fraga BM, 1988. Natural sesquiterpenoids. Natural Product Reports. 5: 497-521.

Funck D, Stadelhofer B, Koch W, 2009. Ornitinedelta-aminotransferase is essential for arginine catabolism but not for proline biosynthesis. BMC Plant Biology, 8: 40-45.

Gershenzon J, Croteau R, 1991. Terpenoids. In Herbivores their interaction with secondary plant metabolites, Vol I: The chemical participants, 2<sup>nd</sup> ed., G.A. Rosenthal and M.R. Berenbaum, eds, Academic press, San Diego, pp: 165-219.

Geu-Flores F, Olsen CE, Halkier BA, 2009. Towards engineering glucosinolates into noncruciferous plants. Planta, 229(2): 261-270.

Giamoustaris A, Mithen R, 1995. The effect of modifying the glucosinolate content of leaves of oilseed rape (*Brassica napus ssp oleifera*) on its interaction with specialist and generalist pests. Annals of Applied Biology, 126: 347–363.

Giraudat J, Hauge BM, Valon C, Smalle J, Parcy F, Goodman HM, 1992. Isolation of the Arabidopsis *AB13* gene by positional cloning. Plant cell, 4: 1251-1261.

Gould JM, 1983. Probing the structure and dynamics of lignin in situ. What's New in Plant Physiology, 14: 25-91.

Grayer RJ, Harborne JB, 1994. A survey of antifungal compounds from higher plants 1982–1993. Phytochemistry, 37: 19-42.

Grayson DH, 1998. Monoterpenoids. Natural Product Reports, 5: 497-521.

Gross D, Porzel A, Schmidt J, 1994. Phytoalexine mit Indolstruktur aus Kohlrabi [Phytoalexins with

indole structure from Kohlrabi]. Zeitschrift fur Naturforschung, 49(5–6): 281–285.

Grubb C, Abel S, 2006. Glucosinolate metabolism and its control. Trends in Plant Science, 11: 89–100.

Gu Q, Kawata EE, Morse MJ, Wu HM, Cheung AY, 1992. A flower specific cDNA encoding a novel thionin in tobacco. Molecular and General Genetics, 234: 89–96.

Gupta VN, Datta SK, 2001. Influence of gibberellic acid on growth and flowering in chrysanthemum (Chrysanthemum morifolium rahmat) cv. Jayanti. Indian Journal of Plant Physiology, 6: 420-422.

Halkier BA, Gershenzon J, 2006. Biology and biochemistry of glucosinolates. Annual Review of Plant Biology, 57: 303-333.

Hammond-Kosack KE, Jones JDG, 1996. Resistance gene dependent plant defence responses. Plant Cell, 8: 1773-1791.

Hammond-kosack KE, Jones JDG, 1997. Plant disease resistance genes. Annual Review of Plant Physiology and Plant Molecular Biology, 48: 575-607.

Harborne JB, 1988. Introduction to ecological biochemistry, Third edition. Academic press, New York.

Harborne JB, 1989. Recent advances in chemical ecology, Natural Products Reports, 6: 85-109.

Harborne JB, 1999. The comparative biochemistry of phytoalexin induction in plants. Biochemical Systematics and Ecology, 27: 335–367.

Hartmann T, 1991. Alkaloids. In herbivores; their interaction with secondary plant metabolites, Vol. I, The chemical participants, 2<sup>nd</sup> ed., G.A. Rosenthal and M.R. Berenbaum, eds Academic press, San Diego, pp: 33-85.

Harvell CD, Tollrian R, 1999. Why inducible defenses? In The ecology and evolution of inducible defenses, eds. Tollrian R and Harvell CD, Princeton, New Jersey: Princeton University Press, pp: 3–9.

Heftmann E, 1975. Function of steroids in plants. Phytochemistry, 14: 891-901.

Hegnauer R, 1988. Biochemistry, distribution and taxonomic relevance of higher plant alkaloids. Phytochemistry, 27: 2423-2427.

Hell R, 1997. Molecular physiology of plant sulfur metabolism. Planta, 202: 138-148.

Huttly AK, Phillips AL, 1995. Gibberellin regulated plant genes. Physiologia Plantarum, 95: 310-317.

Irving HR, Gehring CA, Parish RW, 1992. Changes in cytosolic pH and calcium of guard cells precede stomatal movements. Proceedings of the National Academy of Sciences of the USA, 89: 1790-1794.

Jagendorf AT, 1967. Acid base transition and photophosphorylation by chloroplast. Federation Proceedings, 26: 1361-1369.

Johnson R, Narvaez J, An G, Ryan C, 1989. Expression of proteinase inhibitors I and II in transgenic tobacco plants: Effects on natural defence against Manduca Sexta larvae. Proceedings of the National Academy of Sciences of the USA, 86: 9871-9875.

Kang SY, Kim YC, 2007. Decursinol and decursin protect primary cultured rat cortical cells from glutamate-induced neurotoxicity. Journal of Pharmacy and Pharmacology, 59(6): 863-870.

Kang Z, Buchenauer H, 2003. Immonocytochemical localization of cell wall-bound thionins and hydroxyproline-rich glycoproteins in *Fusarium culmorum*-infected wheat spikes. Journal of Phytopathology, 151(3): 120-129.

Karban R, Baldwin IT, 1997. Induced responses to herbivory. Chicago: University of Chicago Press.

Keeler RF, 1975. Toxins and teratogens of higher plants. Lloydia, 38: 56-86.

Keen NT, Kennedy BW, 1974. Hydroxyphaseollin and related isoflavonoids in the hypersensitive resistance reaction of soybeans to *Pseudomonas glycinea*. Physiological Plant Pathology, 4: 173-185.

Klein RM, 1987. The green world: An introduction to plants and people. New York: Harper and Row.

Knoff DB, 1991. Regulatory phosphorylation of chloroplast antenna proteins, Trends Biochemical Sciences, 16: 82-83.

Koch J, 1989. Breeding developments in double low oilseed rape: Looking to the future. In Proceedings of the Double Low Oilseed Rape for the 1990's Conference. Cambridge: Semundo/BASF.

Kondo T, Yoshida K, Nakagawa A, Kawai T, Tamura H, Goto T, 1992. Structural basis of bluecolor development in flower petals from commelina communis. Nature, 358: 515-518.

Kusnieczyk A, Winge P, Midelfart H, Armbruster WS, Rossiter JT, Bones AM, 2007. Transcriptional responses of Arabidopsis thaliana ecotypes with different glucosinolate profiles after attack by polyphagous myzus persicae and oligophagous brevicoryne brassicae. Journal of Experimental Botany, 58(10): 2537-2552.

Lake JA, Field KJ, Davey MP, Beerling DJ, Lomax BH, 2009. Metabolomic and physiological responses reveal multi-phasic acclimation of Arabidopsis thaliana to chronic UV radiation. Plant, cell & envirnment, 32(10): 1377-1389.

Lappartient AG, Touraine B, 1997. Gltathionemediated regulation of ATP Sulphurylase activity,  $SO_4^{2^-}$  uptake and oxidative stress response in intact canola roots. Plant physiology, 114: 177-183.

Lappartient AG, Vidmar JJ, Leustek T, Glass ADM, Toraine B, 1999. Inter-organ signalling in plants: Regulation of ATP sulphurylase and sulphate transporter gnes expression in roots mediated by phloem-translocated compound. The Plant Journal, 18: 89-95.

Lazzeri L, Tacconi R, Palmieri S, 1993. In vitro activity of some glucosinolates and their reaction products toward a population of the nematode *Heterodera schachtii.* Journal of Agricultural and Food Chemistry, 41: 825–829.

Leustek T, 2002. Sulfate metabolism. Somerville CR, Meyerowitz EM, eds, The Arabidopsis Book. American Society of Plant Biologists, Rockville, MD, doi/10.1199/tab.0009.

Leustek T, Martin MN, Bick JA, Davies JP, 2000. Pathways and regulation of sulphur metabolism revealed through molecular and genetic studies. Annual Review of Plant Physiolgy and Plant Molecular Biology, 51: 141–165.

Leustek T, Saito K, 1999. Sulfate transport and assimilation in plants. Plant Physiology, 120: 637–643.

Lewis NG, Yamamoto E, 1990. Lignin: Occurrence, biogenesis and biodegradation. Annual Review of Plant Physiolgy and Plant Molecular Biology, 41: 455-496.

Lewis WH, Elvin-Lewis MPF, 1977. Medical Botany; plants affecting mans health. Wiley, New York.

Lichtenthaler HK, 1996. Vegetation stress: An introduction to the stress concepts in plants. Journal of plant Physiology, 148: 4-14.

Lipka U, Fuchs R, Kuhns C, Petutschnig E, Lipka V, 2010. Live and let die-Arabidopsis non-host resistance to powdery mildews. European Journal of Cell Biology, 89(2): 194-199.

Loomis WD, Croteau R. 1980. Biochemistry of terpenoids. Stumpf PK (ed.), Lipids, structures and functions. The Biochemistry of Plants, Vol.4. Academic Press, New York, pp: 363-418.

Mader M, Amberg-Fisher V, 1982. Role of peroxidase in lignifications of tobacco cells. Oxidation of nicotinimide adenine dinucleotide and formation of hydrogen peroxide by cell wall peroxidises. Plant physiology, 70: 1128-1131.

Mansfield JW, 2000. Antimicrobial compounds and resistance. The role of phytoalexins and phytoanticipins. In: Slusarenko A, Fraser R, Van

Loon L, eds. Mechanisms of resistance to plant diseases, Netherlands: Kluwer Academic Publishers. pp: 325–370.

Mauricio R, 1998. Costs of resistance to natural enemies in field populations of the annual plant *Arabidopsis thaliana*. The American Naturalist, 151(1): 20–28.

Mayer AM, 1987. Polyphenols oxidase in plantsrecent progress. Phytochemistry, 26: 11-20.

Mccarty D, Hattori T, Carson CB, Vasil V, Lazar M, Vasil IK, 1991. The vivporous-1 development gene of maize encodes a novel transcription activator. Cell, 66: 895-905.

Mehta RG, Liu J, Constantinou A, Thomas CF, Hawthorne M, You M, Gerhauser C, Pezzuto JM, Moon RC, Moriarty RM, 1995. Cancer chemopreventive activity of brassinin, a phytoalexin from cabbage. Carcinogenesis, 16(2): 399–404.

Mes JJ, Van Doorn AA, Wijbrandi J, Simons G, Cornelissen BJC, Haring MA, 2000. Expression of the Fusarium resistance gene I-2 colocalizes with the site of fungal containment. The Plant Journal, 23: 183-193.

Mithen R, 1992. Leaf glucosinolate profiles and their relationship to pest and disease resistance in oilseed rape. Euphytica, 63: 71–83.

Mithen R, Lewis BG, Fenwick GR, 1986. In vitro activity of glucosinolates and their products against *Leptosphaeria maculans*. Transactions of British Mycological Society, 87: 433-440.

Mithen R, Magrath R, 1992. Glucosinolates and resistance to *Leptosphaeria maculans* in wild and cultivated *Brassica* species. Plant Breeding, 108: 60–68.

Monde K, Osawa SM, Harada N, Takasugi M, Suchy M, Kutschy P, Dzurilla M, Balentova E, 2000. Synthesis and absolute stereochemistry of a cruciferous phytoalexin, (-)-Spirobrassinin. Chemistry Letters, 598: 886-887.

Monde K, Takasugi M, 1992. High-performance liquid chromatographic analysis of cruciferous phytoalexins using complex ternary mobile phase gradients. Journal of chromatography, 598: 147-152.

Mordue AJ, Blackwell A, 1993. Azadirachtin: an update. Journal of Insect Physiology, 39: 903-924.

Murray RDH, Mendez J, Brown SA, 1982. The natural coumarins, Wiley, New York.

Nocito FF, Pirovano L, Cocucci M, Sacchi GA, 2002. Cadmium-induced sulfate uptake in maize roots. Plant Physiology, 129: 1872–1879.

Noctor G, Foyer CH, 1998. Ascorbate and glutathione: keeping active oxygen under control.

Annual Review of Plant Physiology and Plant Molecular Biology, 49: 249-279.

Novak K, Lisa L. Skrdleta V, 2004. Rhizobial nod gene-inducing activity in pea nodulation mutants: Dissociation of nodulation and flavonoid response. Physiologia Plantarum, 120(4): 546-555.

Oates JF, Waterman PG, Choo GM, 1980. Food selection by the south Indian leaf-monkey, Presbytis johnii, in relation to leaf chemistry. Oecologia, 45: 45-56.

Ouzouidou G, Llias I, 2005. Hormone induced protection of sunflower photosynthetic apparatus against copper toxicity. Plant Biology, 49: 223-228.

Parashina EV, Serdobinskii LA, Kalle EG, Lavorova NA, Avetisov VA, Lunin VG, Naroditskii BS, 2000. Genetic engineering of oilseed rape and tomato plants expressing a radish defensin gene. Russian Journal of Plant Physiology, 47: 417–423.

Pearce G, Strydom D, Johnson S, Ryan CA, 1991. A polypeptide from tomato leaves induces wound inducible protienase inhibitor proteins. Science, 253: 895-898.

Pedras MSC, Khan AQ, Taylor JL, 1997. Phytoalexins from Brassicas: Overcoming plants defenses. Phytochemicals for pest control. ACS Symposium Series, 658: 155–166.

Pedras MSC, Khan AQ, Taylor JL, 1998. The phytoalexin camalexin is not metabolized by *Phoma lingam*, *Alternaria brassicae*, or phytopathogenic bacteria. Plant Science, 139: 1–8.

Pedras MSC, Sorensen JL, 1998. Phytoalexin accumulation and antifungal compounds from the crucifer wasabi. Phytochemistry, 49: 1959-1965.

Peumans WJ, Van Damme EJM, 1995. Lectins as plant defence proteins. Plant Physiology, 109: 347-342.

Picman AK, 1986. Biological activities of sesquiterpene lactones. Biochemical systematics and Ecology, 14: 255-281.

Porrini M, 2008. Functional foods: from theory to practice. International Journal of Vitamin and Nutrition Research, 78(6): 261-268.

Posmyk MM, Kontek R, Janas KM, 2009. Antioxidant enzymes activity and phenolic compounds content in red cabbage seedlings exposed to copper stress. Ecotoxicology and Environmental Safety, 72(2): 596-602.

Poulton JE, 1990. Cyanogenesis in plants. Plant Physiology, 94: 401-405.

Poulton JE, Moller BL, 1993. Glucosinolates. In Enzymes of secondary metabolism, series: Methods in plant biochemistry; eds, Lea PJ, Chapter 8. London: Academic Press, pp: 209–237. Putnam AR, Heisey RM, 1983. Allelopathy chemical interaction between plants. What's New in Plant Physiology, 14: 21-24.

Putnam AR, Tang CS, 1986. The science of allelopathy, Wiley New York.

Ratzka A, Vogel H, Kliebenstein DJ, Mitchell-Olds T, Kroymann J, 2002. Disarming the mustard oil bomb. Proceeding of the National Academy of Sciences, 99(17): 11223–11228.

Rea PA, Li ZS, Lu YP, Drozdowicz YM, Martinoia E, 1998. From vacuolar GS-X pumps to multispecific ABC transporters. Annual Review of Plant Physiology and Plant Molecular Biology, 49: 727– 760.

Reigosa MJ, Pedrol N, Sanchez-Moreiras AM, Gonalez L, 2002. Stress and allelopathy. In allelopathy: from molecules to ecosystems, eds. Reigosa M, Pedrol N, Enfield, New Hampshire: Science Publishers, pp: 231-256.

Reigosa MJ, Sanchez-Moreiras AM, Gonalez L, 1999. Ecophysiological approach in allelopathy. Critical Review of Plant Sciences, 18: 577-608.

Renwick JAA, Radke CD, Sachdev-gupta K, Staedler E, 1992. Leaf surface chemicals stimulating oviposition by pieris rapae (Lepidoptera: Pieridae) on cabbage. Chemoecology, 3: 33-38.

Rice EL, 1984. Allelopathy, second edition. Academic Press, New York.

Robinson T, 1980. The organic constituents of Higher Plants, Fourth Edition. Cordus Press.

Rosenthal GA, 1991. The biochemical basis for the deleterious effects of L-canavanine. Phytochemistry, 30: 1055-1058.

Rosenthal GA, Berenbaum MR, 1992. Herbivores: Their interaction with secondary plant metabolites, Vol II Ecological and evolutionary processes, 2<sup>nd</sup> edition academic press, San Diego.

Ryals JA, Neuenschwander UH, Willits MG, Molina A, Steiner HY, Hunt MD, 1996. Systematic acquired resistance. Plant cell, 8: 1809-1819.

Saito K, 2004. Sulfur assimilatory metabolism. The long and smelling road. Plant Physiology, 136: 2443–2450.

Savirnata NM, Jukunen-Titto R, Oksanen E, karjalainen RO, 2010. Leaf phenolic compounds in red clover (Trfolium Pratense L.) induced by exposure to moderately elevated ozone. Environmental Pollution, 158(2): 440-446.

Schafer H, Wink M, 2009. Medicinally important secondary metabolites in recombinant microorganisms or plants: progress in alkaloid biosynthesis. Biotechnology Journal, 4(12): 1684-1703.

Schaller A, Ryan CA, 1996. Systemin- a polypeptide signal in plants. Bioessays, 18: 27-33.

Seigler DS, 1981. Secondery metabolites and plant systematic. Conn EE (ed), The biochemistry of plants, Vol 7. Secondery plant products. Plenum, New York and London, pp: 139-176.

Serghini K, Perez De Lugue A, Castejon MM, Garcia TL, Jorrin JV, 2001. Sunflower (Helianthus annuus L.) response to broomraoe (Orobanche cernua loefl.) parasitism: induced synthesis and excretion of 7-hydroxylated simple coumarins. Journal of Experimental Botany, 52: 227-234.

Shulaev V, Silverman P, Raskin I, 1997. Airborne signalling by methyl salicylate in plant pathogen resistance. Nature, 385: 718-721.

Siemans DH, Haugen R, Matzner S, Vanasma N, 2009. Plant chemical defence allocation constrains evolution of local range. Molecular Ecology, 18(23): 4974-4983.

Siemens DH, Garner SH, Mitchell-Olds T, Callaway RM, 2002. Cost of defense in the context of plant competition: *Brassica rapa* may grow and defend. Ecology, 83(2): 505–517.

Simms EL, 1992. Costs of plant resistance to herbivory. In Plant resistance to herbivores and pathogens. eds. Ecology, evolution and genetics, Fritz RS and Simms EL, Chicago: University of Chicago Press, pp 392-425.

Slama K, 1979. Insect hormone and antihormones in plants pages 683-700 in G.A. Rosenthal and D.H. Janzen (eds), herbivores: their interaction with secondary plant metabolites, Academic press, New York.

Slama K, 1980. Animal hormone and antihormones in plants. Biochemie Physiologie Pflanzen, 175: 177-193.

Sreevidya VS, Srinivasa RC, Rao C, Sullia SB, Ladha JK, Reddy PM, 2006. Metabolic engineering of rice with soyabean isoflavone synthase for promoting nodulation gene expression in rhizobia. Journal of Experimental Botany, 57(9): 1957-1969.

Stone B, 1989. Cell wall in plant microanism association. Australian Journal of Plant Physiology, 16: 5-17.

Stotz HU, Kroymann J, Mitchell-Olds T, 1999. Plantinsect interactions. Current Opinion in Plant Biology, 2: 268-272.

Taiz L, Zeiger E, 1995. Plant Physiology Edition. Panima Publishing Corporation, New Delhi, Bangalore.

Talalay P, Fahey JW, 2001. Phytochemicals from cruciferous plants protect against cancer by modulating carcinogenic metabolism. Journal of Nutrition, 131: 3027–3033.

Templeton MD, Lamb CJ, 1988. Elicitors and defence gene activation. Plant, cell and Environment, 11: 395-401.

Terras FR, Eggermont K, Kovaleva V, Raikhel NV, Osborn RW, Kester A, Rees SB, Torrekens S, Van-Leuven F, Vanderleyden J, 1995. Small cysteinerich antifungal proteins from radish: their role in host defense. Plant Cell, 7: 573–588.

Thiel G, Macrobbie EAC, Blatt MR, 1992. Membrane transport in stomatal guard cells, The importance of voltage control. Journal of Membrane Biology, 126: 1-18.

Thomma BPHJ, Cammue BPA, Thevissen K, 2002. Plant defenses. Planta, 216(2): 193–202.

Thomma BPHJ, Eggermont K, Penninckx IAMA, Mauch-Mani B, Vogelsang R, Cammue BPA, Broekaert WF, 1998. Separate jasmonatedependent and salicylate-dependent defenseresponse pathways in Arabidopsis are essential for resistance to distinct microbial pathogens. Proceedings of the National Academy of Science, 95: 15107-15111.

Turlings TCJ, Loughrin JH, Mccall PJ, Roese USR, Lewis WJ, Tumlinson JH, 1995. How caterpillardamaged plants protect themselves by attracting parasitic wasps. Proceeding of the National Academy of Sciences of the USA, 92: 4169-4174.

Van Etten H, Temporini E, Wasmann C, 2001. Phytoalexin (and phytoanticipin) tolerance as a virulence trait: why is it not required by all pathogens? Physiological and Molecular Plant Pathology, 59: 83-93.

Van Etten HD, Mansfield JW, Bailey JA, Farmer EE, 1994. Two classes of plant antibiotics: Phytoalexins versus "phytoanticipins". Plant Cell, 6: 1191-1192.

Van Loon LC, Pierpoint WS, Boller T, Conejero V, 1994. Recommendations for naming plant pathogenesis-related proteins. Plant Molecular Biology Reporter, 12: 245–264.

Van Steveninck RFM, Van Steveninck ME, 1983. Abscisic acid and membrane transport. In Abscisic Acid, Addicott FT, ed., Praeger, New York, pp: 171-235.

Vander-Ent S, Van-Hulten M, Pozo MJ, Czechowski T, Udvardi MK, Pieterse CM, Ton J, 2009. Priming of plant innate immunity by rhizobacteria and aminobutyric acid: differences and similarities in regulation. New Phytologist, 183: 419-431.

Vickery B, Vickery ML, 1981. Secondery plant metabolism. University Park Press, Baltimore.

Vijayan P, Shockey J, Levesque CA, Cook RJ, Browse J, 1998. A role for jasmonate in pathogen defense of Arabidopsis. Proceedings of the National Academy of Science, 95: 7209-7214. Wallsgrove R, Benett R, Kiddle G, Bartlet E, Ludwig-Mueller J, 1999. Glucosinolate biosynthesis and pest disease interactions. In Proceedings of the 10th International Rapeseed Congress, Canberra, Australia.

Whittaker RH, Feeny PP, 1971. Allelochemicals: chemical interaction between species. Science, 171: 757-770.

Wink M, 1999. Functions of plant secondary metabolites and their exploitation in biotechnology. In Annual plant reviews, Vol. 3. Boca Raton, Florida: CRC Press.

Wretblad S, Dixelius C, 2000. B-genome derived resistance to *Leptosphaeria maculans* in near isogenic *Brassica napus* lines is independent of

0

glucosinolate profile. Physiologia Plantarum, 110: 461–468.

Wuyts N, De waele D, Swennen R, 2006. Extraction and partial characterization of polyphenol oxidase from banana (Musa acuminate grandr naine) roots. Plant Physiology and Biochemistry, 44: 308-314.

Zhang H, Fang Q, Zhang Z, Wang Y, Zheng X, 2009. The role of respiratory burst oxidase homologues in elicitor-induced stomatal closure and hypersensitive response in nicotiana benthamiana. Journal of Experimental Botany, 60: 3109-3122.

Zukalova H, Vasak J, 2002. The role and effects of glucosinolates of Brassica species: A review. Rostlinna Vyroba, 48(4): 175–180.

Figures follow.....

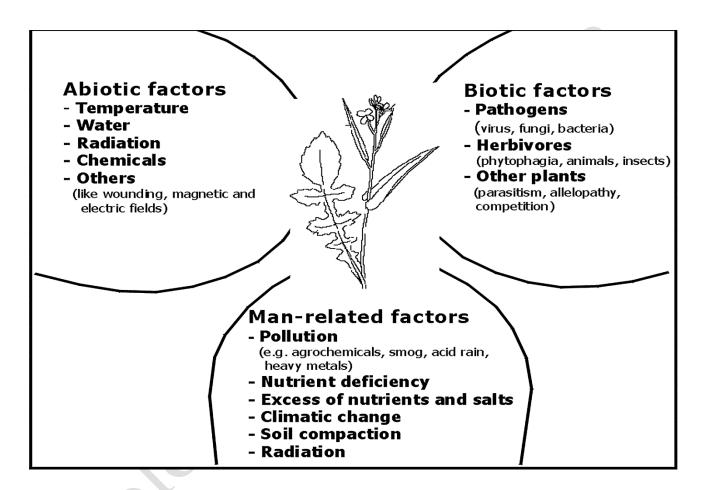


Figure 1. Biotic, abiotic and man related factors that can induce stress related reactions in terrestrial plants (Lichtenthler, 1996; Reigosa et al, 1999, 2002; Bloem, 2005).

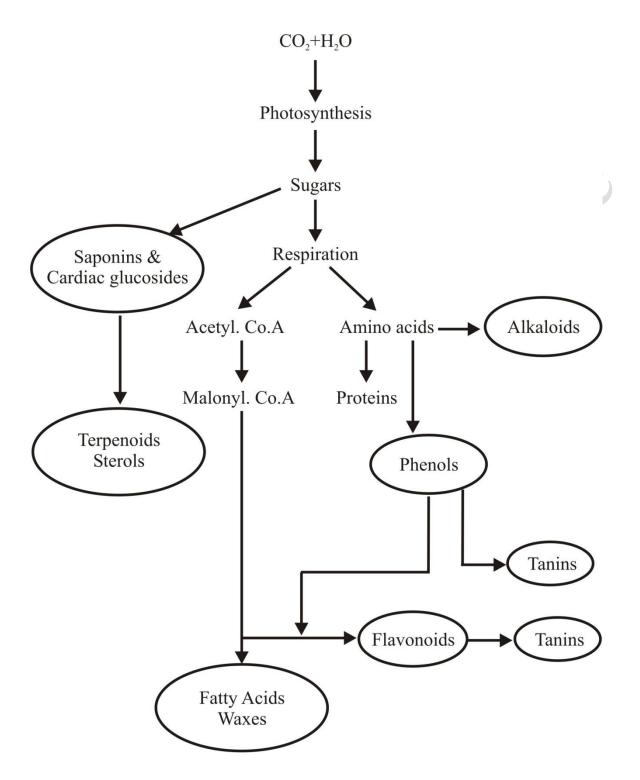


Figure 2. Biosynthetic relationship among some primary and secondary metabolites. The principal group of secondary metabolites are circled.

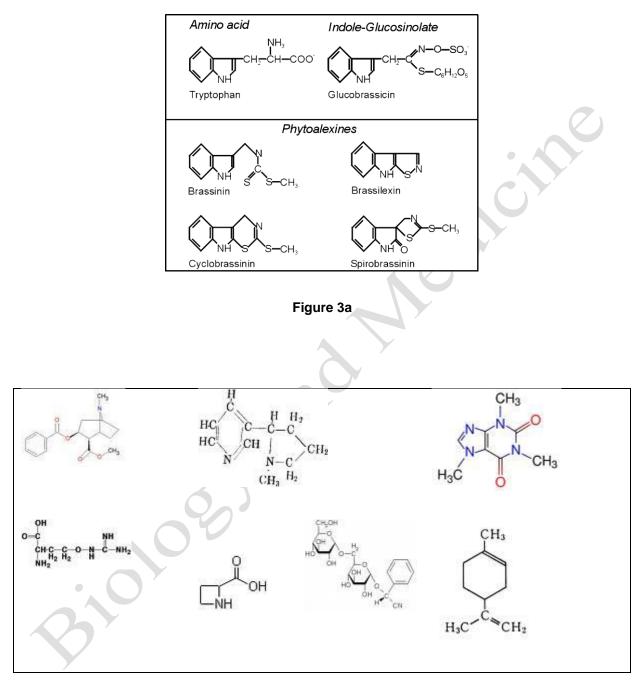


Figure 3b

Figure 3. Structure of N and S containing secondary metabolites and their precursor amino acids. (a) Structure of some typical cruciferous phytoalexins (Monde and Takasugi, 1992) (b) Structures of cocaine, nicotine, caffeine, canavanine, azetidine-2-carboxlicac, amygdalin, limonene.

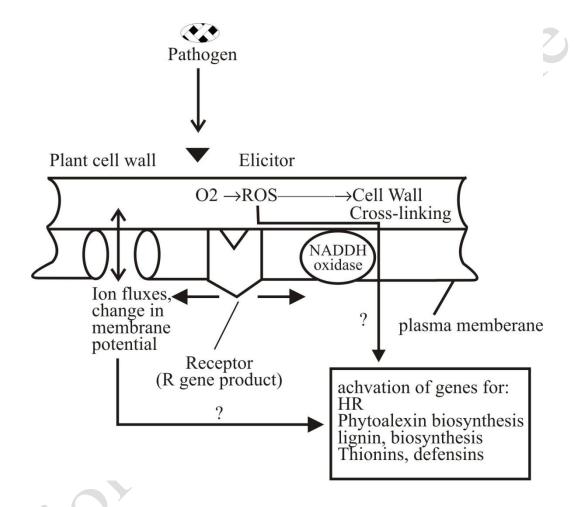


Figure 4. Many modes of anti-pathogenic defence are induced by infection. Fragments of pathogen molecules called elicitors initiate a complex signalling pathway leading to the activation of defence responses.