Research Article

POOL-SEQ Study of Bulgarian Centenarians Highlights the Relevance for Human Longevity of Gene Expression Pathways

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ABSTRACT

In human longevity studies a large number of genetic variants with small effects have been identified, but these are not easily replicable in different populations. We have performed whole-exome sequencing of two DNA pools 32 Bulgarian centenarians and 61 young healthy controls, respectively. A total of 59935 filtered variants were discovered, 216 of which were included in LongevityMap database which lists 2843 longevity associated variants. Using Fisher's exact test, 22 of these variants showed significantly higher allele frequency in the centenarian compared to the control pool and are thus positively associated with longevity. Other 24 variants had significantly higher frequency in the controls and could be considered as negatively associated with longevity. The risk C allele in rs429358 of the APOE gene was only detected in the control pool and with lower frequency compared to other populations. REACTOME analyses showed that over-represented pathways with positive longevity variants belong to expression/transcription network with leading role of TP53, interplaying with other genes (ATR, FANCD2, BAX, BRIP1), whereas those with negative longevity variants belong to the signal transduction network. Our results confirm the importance of studying centenarians in different populations to discover those combinations of variants that associate with longer health span.

Keywords: Centenarian; Exome; Molecular pathways

INTRODUCTION

Longevity is an exceedingly complex phenotype, a quantitative trait that depends on the cumulative action of many genes and the environment. The genetic component that determines longevity has been estimated to be around 30% [1], and this genetic contribution increases with age. Centenarians and supercentenarians, the extreme phenotypes of human lifespan, are thus unique cohorts to study the genetic basis of longevity and the factors determining the risk of various age-related disorders. The genomics of multifactorial diseases or conditions require sequencing a large number of genomes at high coverage; this is mandatory both in order to reach sufficient power for case-control analysis and to compare the patterns of genetic variations across populations. Yet, few studies employing whole-

exome or whole-genome sequencing in centenarian populations to identify risk or beneficial variants relevant to extreme longevity have been performed to date.

Genetic factors can contribute to longevity in at least two important ways: an individual may inherit certain genetic variants that confer disease resistance thereby prolonging lifespan, or may inherit genetic variants that predispose him or her to disease thus decreasing longevity. Human longevity studies have identified a large number of genetic variants; these however provide limited biological insights as they are of small effect and are not easily replicated in different populations. One of the very few variants that has been associated with longevity across a number of studies is rs429358 in the APOE gene with lower frequency of C allele in centenarians [2]. Clarifying the

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role of such variants is further complicated by the evidence that they operate in intricate network of interactions through biological pathways to influence the final phenotype. New insights may thus come from the combined analysis of different genes or SNPs, especially when grouped by metabolic pathways or gene region. Pathway-based analyses thus allow the interpretation of variants with respect to the biological processes in which the affected genes and proteins are involved. As such, pathway analyses are an important follow-up for genome-wide association studies (GWAS) to provide mechanistic insight about the underlying biology of longevity. By identifying wellannotated pathways that map to the lists of significant genes identified in a GWAS, biochemical hypotheses can be enumerated and tested. The effect on longevity of SNPs belonging to certain candidate pathways, such as the insulin/ insulin like growth factor signaling (IIS) and DNA repair, have repeatedly been demonstrated [3, 4].

Sequencing DNA pools (Pool-seq) is a cost-effective method widely used in genomic/exomic sequencing. Pool-seq methods have been repeatedly shown to provide reliable estimates of allele frequencies [5-7], making them particularly suitable for unravelling the genetic basis of complex traits such as longevity. In this study, we first obtain data on SNP and indel variants from whole-exome sequencing two DNA pools, one compiled of Bulgarian centenarians and the other with co-ethnic healthy young controls. Of all annotated variants, we select variants designated as being associated with longevity (LAVs) in the publicly accessible database LongevityMap [8]. The functional significance of the set of genes with variants that show significant allele frequency difference between the two pools was subsequently analyzed using REACTOME pathway platform.

MATERIALS AND METHODS

Ethics statement, centenarian and control subjects recruitment

The project was approved by the Ethics committee of the Medical University of Sofia and was found to be in accordance with the requirements for conducting research with the participation of human subjects, as well as with the national and international legislation. Each participant in the study was informed with the aims of the project. Written informed consent was obtained from participants before collecting samples for DNA extraction. Centenarian health status and medical history for major age-related diseases were based on interviews conducted with the subjects. Tissue samples (saliva or blood sample) were collected from a group of Bulgarian centenarians [100 to 106 year old] and ethnically matched control group composed of young healthy individuals aged 25-30 years.

DNA isolation, WES sequencing and statistical analyses

DNA was extracted using QIAamp DNA Blood Mini Kit (Qiagen) and equimolar amounts of DNA were used to prepare two pools: one with 32 Bulgarian centenarians and one with 61 young individuals as controls. These were whole-exome sequenced using BGI v4 chemistry on a BGISEQ-500 platform

(by BGI Genomics) at a mean 250x coverage. Such high coverage is required for pool-seq sequencing to ensure that alleles with low frequency are also detected [5]. The obtained .VCF files were annotated using the web-based service wANNOVAR [9]. Following 'best the recommendations for pool-seq data [7], we performed robust filtering on variant calling: genotype quality ≥ 99, mapping quality ≥ 60, number of reads per MAF>2, total depth of coverage above 30 and below 500. The total number of variants annotated in both pools after applying these filters was 59935 (52870 SNPs and 7065 indels). The number of allele reads for each variant in both pools was used to construct contingency tables and Fisher's exact test was then deployed to evaluate the significance of the allele frequency differences. The allele frequency estimates obtained were compared with values taken from the publicly available resource for exome sequencing data, The Genome Aggregation Database (gnomAD) [10]. False Discovery Rate (FDR) adjustment of Benjamini and Hochberg [11] was used to reduce the number of false positives. All statistical analyses were performed using R scripts [12].

LongevityMap

We used LongevityMap, a publicly accessible database of human genetic variants associated with longevity to compile a list of variants. This database lists a total of 2843 variants, 510 of which denoted to be significantly and 2333 denoted to be non-significantly associated with longevity. We then examined our pool-seq data for the presence and the frequency of these variants.

Pathway analysis

Pathway analysis methods use a variety of different strategies to aggregate or interpret individual marker or gene based phenotype association statistics to yield a single interpretable test statistic (or p-value) summarizing the strength of evidence of association between the pathway and the phenotype. Pathway analyses simplify the interpretation by placing findings in context of prior knowledge, as well as the analysis (by reducing the multiple comparisons burden inherent to genome-wide approaches). The functional significance of the assemblage of genes that have variants showing significant difference between the two pools was examined using the web-based platform REACTOME [13].

RESULTS

Assessment of the accuracy of allele frequency estimation

In order to verify the adequacy of the population allele frequency estimation using pool-seq data, we compared the estimated individual allele frequencies from the Bulgarian control group with allele frequencies from the gnomAD database. The obtained correlation coefficient (Pearson's r) for non-Finnish Europeans is 0.96, suggesting a high degree of accuracy of allele frequency estimation from pool-seq data.

Longevity associated variants (LAVs)

Of all 2843 variants designated to be associated with longevity in LongevityMap, 216 were discovered in both the centenarian and control pools. Forty six of these showed significant difference between the two pools and they were in 43

genes. Twenty two variants in 20 genes are positively associated with longevity (positive LAVs), i.e. their frequency is significantly higher in the centenarian pool (FDR adj. p-value<0.05) (Table 1).

Table 1: Longevity associated SNPs found in the pool-seq data from Bulgarian centenarians and a control group.

Chr (n)	Ref	Alt	Function	Gene	Exonic function	dbSNP	Frequency centenarians	Frequency controls	FDR p-Value
Variants de	esignated in	Longevity	Map as being si	ignificantly ass	sociated with longevity				
17	G	С	exonic	TP53	nonsynonymous	rs1042522	0.844	0.724	0.001
7	С	T	exonic	EGFR	synonymous	rs2072454	0.553	0.438	0.046
6	A	G	ncRNA exonic	СМАНР	•	rs303006	0.854	0.705	<0.0001
Variants de	esignated in	Longevity	Map as being n	on-significant	ly associated with long	evity			
2	С	G	exonic	CYP1B1	nonsynonymous	rs1056836	0.626	0.502	0.021
1	Т	С	intronic	IL10		rs1518111	0.741	0.565	0.005
11	A	G	exonic	GSTP1	nonsynonymous	rs1695	0.347	0.242	0.041
14	G	A	exonic	MLH3	nonsynonymous	rs175080	0.577	0.453	0.029
3	С	Т	exonic	ATR	synonymous	rs1802904	0.87	0.729	<0.0001
19	A	G	intronic	BAX		rs1805419	0.839	0.663	0.001
11	Т	С	exonic	ACTN3	unknown	rs1815739	0.766	0.599	<0.0001
11	Т	С	exonic	MCAM	synonymous	rs2249466	0.802	0.689	0.013
3	С	Т	UTR3	FANCD2		rs3172417	0.39	0.283	0.013
5	G	A	intronic	RASA1		rs3752862	0.588	0.387	0.048
8	С	Т	intronic	RECQL4		rs4251689	0.569	0.447	0.043
17	С	G	intronic	ACE	,	rs4295	0.783	0.557	0.013
17	Т	С	intronic	ACE		rs4311	0.665	0.488	0.012
17	A	G	exonic	ACE	synonymous	rs4331	0.529	0.351	0.009
17	A	G	exonic	BRIP1	nonsynonymous	rs4986764	0.585	0.472	0.018
11	G	С	UTR3	APOC3		rs5128	0.925	0.847	0.011
17	Т	С	exonic	SLC2A4	synonymous	rs5435	0.72	0.622	0.018
3	G	A	exonic	IMPG2	nonsynonymous	rs571391	0.633	0.48	<0.0001
7	С	T	exonic	TAS2R16	nonsynonymous	rs860170	0.688	0.458	<0.0001

Chr - chromosome number, Ref - referent allele, Alt-alternative allele

Three of these variants are designated as being significantly associated with longevity by LongevityMap, while the remaining 19 have been designated to be non-significantly associated.

Gene set analysis for molecular pathways based on genes with positive LAVs

Twelve out of 15 significant REACTOME pathways (FDR adj. p-value<0.05) with genes carrying positive LAVs are part of the gene expression/transcription network (GET) (Figure 1 and Table 2).

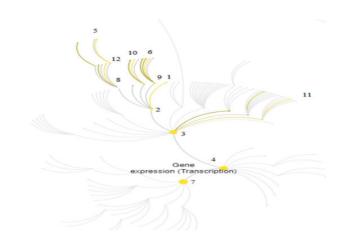


Figure 1: REACTOME Gene expression (Transcription) Network. The numbers denote the pathways (marked in yellow color) containing genes with variants positively associated with longevity: (1). TP53 Regulates Transcription of DNA Repair Genes; (2). Transcriptional Regulation by TP53; (3). Generic Transcription Pathway; (4). RNA Polymerase II Transcription; (5). Regulation of TP53 Expression; (6). TP53 Regulates Transcription of Genes Involved in Cytochrome C Release; (7). Gene expression (Transcription); (8). Regulation of TP53 Activity; (9). TP53 Regulates Transcription of Cell Death Genes; (10). TP53 Regulates Transcription of Genes Involved in G2 Cell Cycle Arrest; (11). TFAP2 (AP-2) family regulates transcription of growth factors and their receptors; (12). Regulation of TP53 Activity through Phosphorylation.

Table 2: Pathways, indicated by REACTOME to be significantly associated (FDR < 1.94e-2) with the set of genes with LAVs.

Nr.	Pathway name	vay name Entities found Entities total		Entities ratio	Entities p-value	Entities FDR	
Gene exp	pression/Transcription						
1	TP53 regulates transcription of DNA repair genes	7	90	0.007	1.145.00	2.68E-07	
	Input: ATR, FANCD2, TP53	- (09	89 0.006 1.14E-09			
2	Transcriptional regulation by TP53	11	407	0.024	5.160.00	(04F 07	
	Input: ATR, BAX, BRIP1, FANCD2, TP53	- 11	486	0.034	5.16E-09	6.04E-07	
2	Generic transcription pathway	12	1525	0.108	1.25E-05	9.75E-04	
3	Input: ATR, BAX, BRIP1, EGFR, FANCD2, TP53	- 13					
	RNA polymerase II transcription			2.110		. 055.00	
4	Input: ATR, BAX, BRIP1, EGFR, FANCD2, TP53	- 13	1664	0.118	3.21E-05	1.85E-03	
5	Regulation of TP53 expression					1.85E-03	
	Input: TP53	- 2	4	0	4.00E-05		

6	TP53 Regulates transcription of genes involved in Cytochrome C release Input: BAX, TP53	3	33	0.002	6.00E-05	2.33E-03		
7	Gene expression (Transcription)	- 13	1822	0.129	8.38E-05	2.77E-03		
	Input: ATR, BAX, BRIP1, EGFR, FANCD2, TP53							
8	Regulation of TP53 activity	- 4	178	0.013	6.79E-04	1.94E-02		
	Input: ATR, BRIP1, TP53	,						
9	TP53 regulates transcription of cell death genes	3 83	0.2	0.006	0 005 04	1.045.02		
9	Input: BAX, TP53	- 3	63	0.006	8.80E-04	1.94E-02		
10	TP53 Regulates Transcription of Genes involved in G2 cell cycle arrest	2	21	0.001	1.06E-03	1.90E-02		
	Input: BAX, TP53	-			0.001 1.06E-03			
11	TFAP2 (AP-2) family regulates transcription of growth factors and their receptors	f 2	21	0.001	1.06E-03	1.94E-02		
	Input: EGFR	-						
12	Regulation of TP53 activity through phosphorylation	3	95	0.007	1.30E-03	1.94E-02		
	Input: ATR							
Cell cyc	cle							
	G2/M DNA damage checkpoint	_						
13	Input: ATR	- 3	81	0.006	8.21e-4	1.94e-2		
Disease								
	Uptake and function of anthrax toxins			0.002				
14	Input: ATR	- 2	22		1.16e-3	1.94e-2		
Immun	ne system							
	Interleukin-4 and Interleukin-13 signaling							
15	Input: IL10, TP53	- 4	211	0.015	1.27e-3	1.94e-2		

Other significant pathways (FDR adj. p-value =1.94e-2) that REACTOME states to be constituted by genes from our positive LAVs gene set are: G2/M DNA damage checkpoint (part of the Cell cycle network); Interleukin-4 and Interleukin-13 signaling (part of the Disease network); uptake and function of anthrax toxins (part of the Immune system network).

Twenty four variants in 23 genes have significantly lower variant frequency in the centenarian pool and it can be speculated that these have disadvantageous effect on longevity (negative LAVs) (Table 3).

Table 3: Variants from LongevityMap with significantly lower frequency in BG centenarians.



Chr	Ref	Alt	Function	Gene	Exonic function	dbSNP	Frequency centenarians	Frequency controls	FDR p-value
Variants	s designated	in Longevi	tyMap as being sign	nificantly associate	d with longevity, with n	negative associat	tion in BG cente	narians	
14	С	А	intergenic	AKT1		rs2498804	0.264	0.392	0.024
1	A	G	exonic	EXO1	nonsynonymous	rs735943	0.411	0.612	0.001
Variants	s designated	in Longevi	tyMap as being no	n-significantly asso	ciated with longevity, w	rith negative ass	ociation in BG o	entenarians	
16	G	A	intronic	PDPK1		rs1005273	0.327	0.512	0.036
11	A	G	exonic	HRAS	synonymous	rs12628	0.29	0.484	<0.000
15	A	Т	exonic	PIF1	nonsynonymous	rs17802279	0.358	0.494	0.028
1	Т	G	exonic	MTHFR	nonsynonymous	rs1801131	0.28	0.396	0.017
1	G	A	exonic	MTHFR	nonsynonymous	rs1801133	0.276	0.384	0.04
16	A	G	intronic	PRKCB		rs198145	0.209	0.373	0.003
7	G	Т	exonic	TAS2R5	nonsynonymous	rs2227264	0.485	0.628	0.006
17	G	A	intronic	RPA1		rs2270412	0.169	0.261	0.022
16	С	G	exonic	CLEC3A	synonymous	rs2293776	0.213	0.334	0.006
19	Т	G	exonic	ERCC2	synonymous	rs238406	0.419	0.559	0.023
2	С	G	intronic	MSH6		rs3136367	0.619	0.773	0.001
19	С	A	exonic	CD3EAP	nonsynonymous	rs3212986	0.183	0.438	0.001
16	A	G	exonic	IGFALS	synonymous	rs3751893	0.688	0.812	0.004
15	Т	С	intronic	BLM		rs3815003	0.201	0.287	0.041
3	A	G	intronic	ADCY5		rs4482616	0.733	0.968	<0.000
7	A	G	exonic	IGFBP1	nonsynonymous	rs4619	0.151	0.347	<0.000
16	Т	G	ncRNA intronic	MT1JP		rs4784701	0.476	0.692	<0.000
1	С	Т	exonic	NTRK1	synonymous	rs6337	0.676	0.785	0.02
2	С	G	intronic	XDH		rs761926	0.208	0.361	<0.000
7	G	Т	intronic	NOS3		rs7830	0.174	0.327	0.046
19	G	С	intronic	GPX4		rs8178977	0.199	0.323	<0.000
2	С	Т	exonic	FSHR	nonsynonymous	rs6166	0.477	0.586	0.013

Gene set analysis for molecular pathways based on genes with negative LAVs

Ten out of 20 significant REACTOME pathways (FDR adj. p-value <0.05) with genes carrying negative LAVs are members of the signal transduction network (Figure 2 and Table 4).

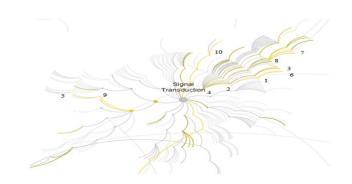


Figure 2: REACTOME Signal Transductions Network. The numbers denote the pathways (marked in yellow color) containing genes with variants negatively associated with longevity: 1. VEGFA-VEGFR2 Pathway; 2. Signaling by VEGF; 3. VEGFR2 mediated vascular permeability; 4. Signaling by Receptor Tyrosine Kinases; 5. G beta:gamma signaling through PI3Kgamma; 6. VEGFR2 mediated cell proliferation; 7. TRKA activation by NGF; 8. Signaling to ERKs; 9. G-protein beta:gamma signaling; 10. Non-genomic estrogen signaling.

Table 4: Pathways, indicated by REACTOME to be significantly associated (FDR adj. p-value < 1.94e-2) with the set of genes with variants negatively associated with longevity.

Nr.	Pathway name	Entities found	Entities total	Entities ratio	Entities p-value	Entities FDR
Sign	al transduction network					
	VEGFA-VEGFR2 Pathway		125	2.222		1.41.4
1	Input: AKT1, HRAS, NOS3, PDPK1, PRKCB	6	125	0.009	4.28e-7	1.41e-4
	Signaling by VEGF		124	2.222	(40F 07	1.415.04
2	Input: AKT1, HRAS, NOS3, PDPK1, PRKCB	——	134	0.009	6.40E-07	1.41E-04
	VEGFR2 mediated vascular permeability			0.003	3.55E-06	5 10F 04
3	Input: AKT1, NOS3, PDPK1	4	44			5.19E-04
	Signaling by receptor tyrosine kinases	0	521	0.037	2.05E-05	1.650.00
4	Input: AKT1, HRAS, NOS3, NTRK1, PDPK1, PRKCB	—— 8	521			1.65E-03
	G beta:gamma signaling through PI3Kgamma		20	0.002	4.48E-05	2.1/2.22
5	Input: AKT1, PDPK1	3	29			2.16E-03
	VEGFR2 mediated cell proliferation				5.00E-05	2.1/2.22
6	Input: HRAS, PDPK1, PRKCB	3	30	0.002		2.16E-03
	TRKA activation by NGF	2		2	(545.05	2.1/5.22
7	Input: NTRK1	2	5	0	6.54E-05	2.16E-03
	Signaling to ERKs	2		0.003	1.07E-04	2 (05.22
8	Input: HRAS, NTRK1	3	39			2.68E-03



9	G-protein beta:gamma signaling	3	39	0.003	1.07E-04	2.68E-03	
9	Input: AKT1, PDPK1)	39	0.003	1.07E-04	2.00E-U3	
10	Non-genomic estrogen signaling	4	100	0.009	1 205 04	2.0(5.02	
10	Input: AKT1, HRAS, NOS3, PDPK1	4	109	0.008	1.20E-04	2.86E-03	
DNA	A repair						
11	Mismatch repair (MMR) directed by MSH2:MSH6 (MutSalpha)	2	22	0.003	2.00.05	1 (55 02	
11	Input: EXO, MSH6, RPA1)	22	0.002	2,00e-05	1.65E-03	
12	Mismatch Repair	2	23	0.002	2 26E 05	1.65E-03	
12	Input: EXO, MSH6, RPA1)	23	0.002	2.206-05	1.03E-03	
13	HDR through single strand annealing (SSA)	2	39	0.002	1.075.04	2.68E-03	
13	Input: BLM, EXO1, RPA1)	39	0.003	1.20E-04 2,00e-05 2.26E-05 1.07E-04 1.53E-04 3.24E-05 1.07E-04	2.00E-03	
14	Presynaptic phase of homologous DNA pairing and strand exchange	3	41	0.003	1.24E-04	2.86E-03	
	Input: BLM, EXO1, RPA1						
15	Homologous DNA pairing and strand exchange	2	44	0.003	1 53E 04	3.36E-03	
13	Input: BLM, EXO1, RPA1		TT	0.003	1.5512-07	J.50E-05	
Imm	une system						
16	CD28 dependent PI3K/Akt signaling	3	26	0.002	3 24E 05	2.01E-03	
10	Input: AKT1, PDPK1		20		3.24E-03	2.016-03	
17	CD28 co-stimulation	3	39	0.003	1 07F 04	2.68E-03	
11	Input: AKT1, PDPK1			0.003	1.071204	2.002-03	
Gen	e expression/Transcription						
18	Regulation of TP53 activity	5	178	0.013	5 56E 05	2.16E-03	
10	Input: AKT1, BLM, EXO1, PDPK1, RPA1	, 	170	0.013	3.30E-03	2.10E-03	
19	RNA polymerase I transcription termination	3	33	0.002	6 56E 05	2.16E.02	
17	Input: CD3EAP, ERCC2, RPA1			0.002	0.301203	2.16E-03	
Dise	ase						
20	Constitutive signaling by AKT1 E17K in cancer	3	32	0.002	6.00E-05	2.16E-03	
20	Input: AKT1, PDPK1	3	34	0.002	U.UUE-UJ	2.10E-03	

Other significant pathways that REACTOME indicates to be constituted by genes from this set are CD28 dependent PI3K/Akt signaling and CD28 co-stimulation (both part of the Immune system network); Regulation of TP53 activity and RNA

polymerase I transcription termination (both part of the Gene expression/Transcription); Constitutive signaling by AKT1 E17K in cancer (Disease network). The remaining 170 variants failed to replicate as longevity-associated variants in Bulgarian

centenarians as there are no significant differences in allele frequencies between the centenarian and control pools.

DISCUSSION

Whole-exome sequencing on DNA pools from Bulgarian centenarians and a control group found 216 out of 2843 longevity associated variants (LAVs) listed in the LongevityMap database. Only 22 of these showed significantly higher allele frequency in the centenarian pool compared to the control pool and can thus be considered positively associated with longevity in Bulgarian population.

LongevityMap database designates only 3 of these 22 variants (in genes TP53, EGFR, CMAHP) to have significant association with longevity, while the remaining 19 variants, although determined to be non-significant LAVs, are significantly associated with longevity in Bulgarian population (see Table 1). The positive LAVs in our study are thus less than 1% of all variants listed in the LongevityMap database. A possible explanation for this low percentage is that longevity is a complex phenotype, being determined by interaction between genetic and environmental factors. LAVs should be studied in different populations as they must deal with different conditions and specific LAV combinations can determine longevity in different ethnic or geographical groups.

Pathway analysis of gene set with positive LAVs in Bulgarian population

Twelve out of 15 REACTOME overrepresented pathways in the set of genes with positive LAVs constitute the gene expression/transcription (GET) network (Fig. 1). Co-expressed genes within a pathway are either controlled by the same transcriptional factors, are functionally related or are members of the same protein complex. A transcriptional regulatory network plays a major role in response to environmental influences and genetic disturbances and it consists of many components [14]. Although there has been much progress in establishing interactions between the molecular pathways in GET network, their role in centenarians remains unclear.

REACTOME pathways are hierarchically arranged, the top level pathway being "Gene expression/transcription" (Fig. 1, pathway 7) in which participate the enzymes RNA polymerase I, II, III and mitochondrial RNA polymerase. It is followed by the pathways "RNA Polymerase II transcription" (Fig. 1, pathway 4) and "Generic transcription pathway" in which cell- or tissue-specific regulation of differential gene transcription is mediated (Fig. 1, pathway 3). These are fundamental pathways for gene expression and starting point for the other pathways in the network. The remaining significant pathways involve overlapping set of positive LAV genes (TP53, ATR, FANCD2, BAX and BRIP1) and here we discuss how they relate to longevity. The gene exhibiting most interactions in these pathways is TP53 and it seems to act as a driver gene in longevity of Bulgarian centenarians.

TP53 (rs1042522) is a variant designated to be significantly associated with longevity that our results confirm is the case in Bulgarian centenarians. The 2 alleles at the rs1042522 locus

encode protein isomorphs that differ in their capacities to induce target gene transcription, their ability to interact with p73 (another tumor suppressor protein) and their targeting of the proteasome. A Danish study finds that minor allele homozygotes, i.e. rs1042522 (C/C) genotypes, live on average 3 years longer than major allele (G/G) homozygotes [15]. The frequency of the C allele in the Bulgarian population is 0.724, similar to European exome frequency 0.738 [16], while the frequency in Bulgarian centenarians is significantly higher (0.844) (see Table 1). The increased longevity is speculated to be related to the increased apoptosis seen for this gain of function SNP [17]. TP53 encodes a tumor suppressor protein which responds to diverse cellular stresses by regulating the expression of genes involved in i.e. inducing cell cycle arrest, DNA repair, or changes in metabolism [18].

Our pathway analysis demonstrated that this gene is set to be involved in several pathways associated with longevity, e.g. regulation of TP53 activity, DNA repair and G2/M DNA damage checkpoint. TP53 directly stimulates transcription of several genes involved in DNA mismatch repair [19-21], in nucleotide excision repair [22], as well as in repairing DNA interstrand crosslinks [23]. Expression of several DNA repair genes is under indirect TP53 control, e.g. genes involved in the repair of DNA double strand breaks and/or the Fanconi anemia pathway [24-27]. Throughout the cell cycle, the genome is constantly monitored for damage, resulting either from errors of, by-products of metabolism or through extrinsic sources such as ultra-violet or ionizing radiation. The different DNA damage checkpoints act to inhibit or maintain the inhibition of the relevant cyclin-dependent kinase that will control the next cell cycle transition.

Our results show significant association between rs1802904 in the ATR gene and longevity, whereas LongevityMap doesn't. The ATR protein is a serine/threonine kinase which phosphorylates TP53 in stress cells and participates in the post-translational modification of TP53 activity [28]. The ATR gene was found to be involved in the regulation of DNA repair and G2/M DNA damage checkpoint pathways [29], and functions as an apoptotic activator [30]. Failure of the G2 DNA damage checkpoint leads to catastrophic attempts to segregate unrepaired chromosomes. The variant rs1802904 is however synonymous and with unknown functional significance, and further research is needed to clarify its effect on longevity.

The variant rs3172417 in FANCD2 gene is not designated as significantly associated with longevity by LongevityMap, nevertheless our results show significant association. The FANCD2 gene is also involved in the regulation of TP53 activity and DNA repair pathways [31]. Part of the Fanconi anemia complementation group, the protein encoded by this gene is monoubiquinated in response to DNA damage, resulting in its localization to nuclear foci with other proteins (BRCA1 and BRCA2) involved in homology-directed DNA repair [32]. The rs3172417 SNP is in a 3' untranslated region and little is known about its functional mechanisms, and its association with longevity should be confirmed in a larger case-control studies.

We find significant association with longevity of rs1805419 in the BAX gene, in line with Erdman, Nasibullin [33] who have found that the G allele of this variant is significantly associated with longevity. BAX is a transcriptional target for the TP53 and plays a role in mitochondrial apoptosis. The protein encoded by this gene forms a heterodimer with BCL2 (Bcl2-Bax) with antiapoptotic function, while Bax/Bax homodimer acts as apoptotic inducer. The factors that provide the fine balance between programmed cell death and cell survival processes probably regulate not only cell longevity but could also be important for human longevity.

Another variant with uncertain association in LongevityMap is rs4986764 in the BRIP1 gene, yet our results indicate significant association. BRIP1 is also involved in regulation of TP53 activity, DNA repair and G2/M DNA damage checkpoint pathways. The protein encoded is a component of a complex important in the normal double-strand break repair function of breast cancer (BRCA1) [34]. Meta-analysis indicates that rs6504074 may lead to decreased risk of gynecologic cancer in the overall population [35].

The EGFR variant rs2072454 has previously been associated with longevity in Koreans [36], and we corroborate this significant association. The interaction of EGRF protein with ligands triggers signaling pathways within the cell and leads to DNA synthesis and cell proliferation. Pathogenic mutations in this gene are associated with lung cancer with head and neck squamous cell carcinoma (HNSCC) and [37].

IL10 (rs1518111) is an intronic non-coding variant. Its functional consequence on the processing of the transcript and the protein is unknown. The protein produced by this gene is an anti-inflammatory cytokine that has pleiotropic effects in immunoregulation and inflammation. Other IL10 gene polymorphisms has previously been linked to longevity, e.g. the high IL10-producer genotype is increased among centenarians [38].

Pathway analysis of gene set with negative LAVs in Bulgarian population

Unlike the set of genes with positive LAVs which were assigned to be part of gene expression/transcription (GET) network, ten of 20 significant pathways of genes carrying putatively disadvantageous variants for longevity are part of a different significantly over-represented network - the signal transduction (ST) network. The ST network includes the following pathways: Signaling by VEGF resulting angiogenesis by mediating endothelial cell proliferation and vascular permeability; Signaling by receptor tyrosine kinases; G-protein beta:gamma signaling; Signaling by AKT1; Signaling to ERKs; Non-genomic estrogen signaling. These pathways include a combination of 6 genes from our input set: AKT1, HRAS, NOS3, PDPK1, PRKCB and NTRK.

All cells are sensitive to specific signals and can respond to changes in their environment [39]. Signal transduction is the transmission of chemical or physical signals from the cell membrane to the nucleus and coordinates the communication among cells and between cells and extracellular matrix. Fine-tuned positive and negative regulation of signaling networks is a critical feature of normal, physiological signaling balance in any

given cell. Despite the presence of cellular protective responses, pathologies (e.g. cancers) frequently develop in tissues due to somatic mutations within these key signal transduction networks [40, 41]. Dysregulation of signal transduction could lead cells to over-proliferating and to bypassing survival and migration mechanisms, thus promoting cancer development. During the aging process, changes in the signaling intensities of these networks could manifest in age-related pathologies [41].

Vascular endothelial growth factor (VEGF) and its receptors (VEGFR) are key regulators of the process of angiogenesis. VEGF/VEGFR signaling pathways include five genes from the set input in REACTOME with negative LAVs in Bulgarian centenarians (AKT1, HRAS, NOS3, PDPK1, and PRKCB). Disturbances in these pathways play role in disease pathology and may be an early step in the process of metastasis.

Signaling by receptor tyrosine kinases is a pathway in which function the same set of 5 genes. Receptor tyrosine kinases (RTKs) are a major class of cell surface proteins involved in common signaling pathways including RAF/MAP kinase cascades [42] and AKT signaling [43]. RTKs regulate many key cell processes and their dysregulation is established in a wide range of cancers.

G-proteins are involved in transmitting signals from outside the cell to the nucleus through connected signaling cascades, and changes in G proteins and their downstream signaling could initiate cancer. AKT1 is a known oncogene, recognized as a critical node in the PI3K/AKT/mTOR pathway. Reduced activity of this nutrient-sensing pathway is also one of the few physiological mechanisms repeatedly shown to be associated with longevity in humans [44].

The extracellular signal-regulated kinase (ERK) signaling pathway plays role in controlling diverse cellular processes such as proliferation, survival, differentiation and motility. The ERK pathway is often up-regulated in human tumors and as such represents a suitable target for the development of anticancer drugs.

Non-genomic estrogen signaling is well characterized in multiple carcinomas. Recent studies have established a potential immune regulatory role of estrogens in the tumor microenvironment, and promote estrogen as a potential mediator of tumor immunosuppression [45]. Estrogen signaling regulates the gene expression of certain chromatin-modifying enzymes and miRNAs and is connected to epigenetic mechanisms.

Other significant pathways indicated by the set of genes showing prevalence of allele frequency in the control pool constitute the DNA repair and immune system networks, both of which have been shown to impact longevity [46,47].

Variants in the APOE gene

Among variants that showed significant difference in allele frequencies between the centenarians and controls were absent variants in the APOE gene. As this gene has been repeatedly associated with longevity, we consider separately the variants in APOE. The risk C allele in rs429358 was only detected in the

control pool and with lower frequency compared to other populations (Table 5).

Table 5: Allele frequency of variants in APOE gene in Bulgarian population.

D 1	Ref	D (A 1.	E	11 CNID	D (11.1 ()	A1. 11.1 ()	Frequency	alt. allele	Other population source
Pool	Ker	Alt	Function	dbSNP	Ref. allele (n)	Alt. allele (n)	Bulgarian	Other populations		
centenarians	С	G	exonic	rs440446	32	50	0.61	0.641	gnomAD_genome_NFE	
controls	С	G	exonic	rs440446	31	38	0.551	0.641	gnomAD_genome_NFE	
centenarians	G	A	intronic	rs769449	737	25	0.033	0.115	gnomAD_genome_NFE	
controls	G	A	intronic	rs769449	534	62	0.104	0.1145	gnomAD_genome_NFE	
centenarians	С	Т	intronic	rs143063029	903	13	0.014	0.0012	gnomAD_genome_NFE	
centenarians	С	Т	exonic	rs121918393	90	2	0.022	0.0001	ExAC	
centenarians	G	A	exonic	rs756564996	90	2	0.022	6.49E-05	gnomAD_genome_ALL	
centenarians	G	A	exonic	rs767339630	132	2	0.015	NA	gnomAD_genome_NFE	
controls	G	A	exonic	rs746382742	1226	4	0.003	0.00001	ExAC	
controls	С	Т	exonic	rs752079771	282	2	0.007	0. 00002	gnomAD_exome_NFE	
controls	Т	С	exonic	rs429358	84	5	0.056	0.143	gnomAD_genome_NFE	

gnomAD_genome_NFE-Genome database with allele frequencies in non-Finnish European population; gnomAD_genome_ALL-Genome database with allele frequencies in all studied populations; gnomAD_exome_NFE - Exome database with allele frequencies in non-Finnish European population; ExAC - exome database of exome aggregation consortium

The absence of this variant in the centenarian pool is in line with previous studies that show this variant is negatively associated with longevity as it is associated with increased risk for cardiovascular disease and Alzheimer disease. Four variants with low frequencies were found only in the centenarian pool: rs143063029, rs121918393, rs756564996, rs767339630. Even though the minor allele frequencies is higher in Bulgarian centenarians compared to other populations, these rare variants are of unknown clinical significance and are not listed in LongevityMap database. More studies are needed to confirm or reject their association with longevity. Three exonic variants in APOE were established only in the control pool: rs746382742, rs752079771, rs429358.

CONCLUSION

By employing a WES approach we ensure that all gene variants listed in the LongevityMap database are analyzed. The pool-seq approach has proven to give accurate population allele frequency estimates, especially after appropriate filters are used. The downside of this approach is that it is not possible to determine individual genotypes. The positive and negative LAVs participate in different over-represented pathways. Variants showing significant enrichment in centenarians are in genes that constitute gene expression/transcription (GET) network with leading role of TP53, interplaying with other genes (ATR,

FANCD2, BAX, BRIP1). Genes carrying negative LAVs, on the other hand, are members of the signal transduction (ST) network.

The involvement of different networks in positive and negative LAVs should be confirmed with additional studies in other populations. The results of this work confirm the importance of studying truly rare survival to discover those combinations of variants associated with extreme longevity and longer health span in genetically different populations. Complex phenotypes such as longevity require not only population level genomic data, but future studies should also include GWAS studies of longevous families, functional analysis of interesting loci, somatic mutations, epigenetic studies and microbiome data.

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CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

AUTHORS CONTRIBUTION

Dimitar Serbezov, Lubomir Balabanski have contributed equally. Lubomir Balabanski, Desislava Nesheva and Olga Boyanova are supported by the Bulgarian Ministry of Education and Science under the National Program for Research "Young Scientists and Postdoctoral Students".

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