Optimization of *Streptomyces* sp.A11 Medium Cultivation on Cyclo(Tyrosyl-Prolyl) Production Using the Response Surface Methodology

Rofiq Sunaryanto

Center of Biotechnology, Agency of The Assessment and Application of Technology (BPPT), gd.630 Kawasan Puspiptek Serpong, Tangerang Selatan, Banten 15314
Email : rofiqsn@yahoo.com

Abstract - The Response Surface Methodology is a three factorial design which provides the relationship between one or more measured dependent response and a number of input (independent) factors. Response surface methodology was used for optimization of fermentation medium for antibacterial agent cyclo(tyr-pro) production by *Streptomyces* sp. A11. The optimal response region of the significant factor was predicted by using a second order polynomial model fitted to the results obtained by applying the Central Composite Design (CCD) statistical design. Determination of the best production medium of cyclo(tyr-pro) showed among 6 carbon sources used, dextrin produced the highest cyclo(tyr-pro). Among the five nitrogen sources used, peptone produced the highest cyclo(tyr-pro), and the mineral salts group I produced the highest cyclo(tyr-pro) compared with others. The results of optimization showed that three variables (dextrin as a carbon source, peptone as nitrogen source, and mixture of mineral salts) used showed significantly affected to the activity of cyclo(tyr-pro). The mathematical model used, obtained by the composition of the three variables most optimum were concentration of dextrin was 32.55 $g.L^{-1}$, peptone concentration was 11.22 $g L^{-1}$, and 8.65 mL of mineral salts. By using the composition of mathematic model, cyclo(tyr-pro) was obtained 51.54 mg L^{-1} (model expectations) and 50.04 mg L^{-1} from experiments. The differences of experiment with expectation response value was 2.91%. Using the optimized parameters, the studies demonstrated an increase in the cyclo(tyr-pro) was 2.5 fold from 20 mg L^{-1} to 50 mg L^{-1}.

*[**Keywords** : response surface methodology; cyclo(tyrosyl-prolyl); Streptomyces sp. A11; optimization]*

Received: September 3, 2012   *Accepted: October 15, 2012*

I. INTRODUCTION

Cyclo(tyrosyl-prolyl) or Cyclo(tyr-pro) is a diketopiperazine formed by the fusion of tyrosine and proline which has been reported as a secondary metabolite of fungi and bacteria. Cyclo(tyr-pro), generically named maculosin, was identified as a host-specific toxin produced by *Alternaria alternate* on *spotted knapweed* [1]. Cyclo(tyr-pro) appears to be highly toxic only to *spotted knapweed* and is being researched for potential field efficacy and bioherbicide. Cyclo(tyr-pro) was also produced by *Streptomyces* sp. A11, actinomycetes which isolated from marine sediment [2] and also produced by oleh *Pseudomonas fluorescens* GcM5-1A [3].

Response Surface Methodology (RSM) is a powerful technique for testing multiple process variables because fewer experimental trials are needed compared to the study of one variable at a time [4]. Also, interactions between variables can be identified and quantified by such a technique. Otherwise, the conventional method of optimization involves varying one parameter at a time and keeping the others constant. This often does not bring about the effect of interaction of various parameters as compared to factorial design [5]. RSM is a useful model for studying the effect of several factors influencing the responses by varying them simultaneously and carry out a limited number of experiments [6].

In this study, we applied response surface methodology (RSM), especially central composite design (CCD), to further attempt to enhance the cyclo(tyr-pro) production. The CCD is conducted in the optimum vicinity to locate the true optimum values of the multiple variables. This investigation is aimed at optimization of medium components (glucose, peptone and mineral salt), which have been predicted to play a very significant role in enhancing the production the cyclo(tyr-pro). Hence the use of experimental factorial design and response surface methodology, already successfully applied in other fields [7,8,9,10], is well suited to the study of the main and interaction effects of the factors on the production of the cyclo(tyr-pro). Selection of the best carbon source, nitrogen source, and mineral salt were conducted by the classical methods involves changing one independent variable while maintaining all others at fixed level. This selection were conducted before optimization using Response Surface Methodology.

II. MATERIALS AND METHODS

A. Microorganism

Streptomyces sp.A11, isolated in our laboratory was used as producer of Cyclo(tyr-pro). It was grown at 30°C for 4 days and maintained on marine agar slants at 4°C and was subcultured at four week intervals.

B. Inoculum Preparation

Inoculum was prepared by transferring 5 mL suspension prepared from a slant culture, into 250 mL Erlenmeyer flask containing 50 mL of sterile inoculum medium. The composition of inoculum medium consisted of bacto peptone 15 g L^{-1}, yeast extract 3 g L^{-1}, malt extract 3 g L^{-1}, glucose 10 g L^{-1} and dissolved into water 12.5 mL, and sea water 37.5 mL, pH was adjusted at 7.6.

C. Production Culture for Determination of the Best of Carbon Source.

Five millilitres of 48 hours aged inoculum *Streptomyces* sp.A11 was added to 50 mL production medium in 250 mL erlenmeyer flask. The composition of inoculum medium consisted of 10 g L^{-1} carbon source, peptone 5 g L^{-1}, yeast extract 1 g L^{-1}, Fe (III) citrate hydrate 0.3 g L^{-1}, demineral water 12.5 mL, and sea water 37.5 mL, pH was adusted at 7.6 [11]. Six kinds of carbon source were investigated for selected the best of carbon source, namely: glucose, maltose, lactose, sucrose, mollase, and dextrine.

D. Production Culture for Determination of the Best of Nitrogen Source.

Five millilitres of 48 hours aged inoculum *Streptomyces* sp. A11 was added to 50 mL production medium in 250 mL Erlenmeyer flask. The composition of inoculum medium consisted of 10 g L^{-1} glucose, nitrogen source 5 g L^{-1}, yeast extract 1 g L^{-1}, Fe (III) citrate hydrate 0.3 g L^{-1}, demineral water 12.5 mL, and sea water 37.5 mL, pH was adusted at 7.6 [11]. Five kinds of nitrogen source were investigated for selected the best of nitrogen source, namely: peptone, yeast extract, glutamic acid, casein, ammonium sulphate.

E. Production Culture for Determination of the Best Mineral Source.

Five millilitres of 48 hours aged inoculum *Streptomyces* sp. A11 was added to 50 mL production medium in 250 mL erlenmeyer flask. The composition of inoculum medium consisted of 10 g L^{-1} glucose, nitrogen source 5 g L^{-1}, yeast extract 1 g L^{-1}, Fe (III) *citrate hydrate* 0.3 g L^{-1}, and added salt solution, demineral water 12.5 mL, and sea water 37.5 mL, pH was adusted at 7.6 [11]. The composition of each salt solution as followed:

No	Salt solution	Composition
1	(M1) The composition of salt solution by Sousa *et al.* [12].	K$_2$HPO$_4$ 1 g L^{-1}, MgSO$_4$.7 H$_2$O 0.025 g L^{-1}, ZnSO$_4$ 7 H$_2$O 0.025 g L^{-1}, CaCl$_2$.2 H$_2$O 0.025 g L^{-1}, FeSO$_4$ 7 H$_2$O 0.025 g L^{-1}.
2	(M2)The composition of salt solution by Furtado *et al.* [13].	KH$_2$PO$_4$ 0.6 g L^{-1}, Mg.SO$_4$.7 H$_2$O 5 g L^{-1}, Cu.SO$_4$.5 H$_2$O 0.001 g L^{-1}, FeSO$_4$.7 H$_2$O 0.003 g L^{-1}
3	(M3)The composition of salt solution by Dhananjeyan *et al.* [14].	K$_2$HPO$_4$ 0.1 g L^{-1}, Mg.SO$_4$.7 H$_2$O 0.5 g L^{-1}, CaCl$_2$.2H$_2$O 0.1 g L^{-1}, FeSO$_4$.5H$_2$O 0.05 g L^{-1}
4	(M4)The composition of salt solution by Voelker & Altaba [15].	CaCl$_2$.2H$_2$O 0.011 g L^{-1}, FeSO$_4$.5H$_2$O 0.007 g L^{-1}, MnCl$_2$.4 H$_2$O 0.002 g L^{-1}, ZnSO$_4$.7 H$_2$O 0.002 g L^{-1}, CuSO$_4$. H$_2$O 0.0004 g L^{-1}, CoCl$_2$.6 H$_2$O 0.0004 g L^{-1}
5	The composition of salt solution by Dharmaraj *et al.* [16].	NaCl 0.8 g L^{-1}, NH$_4$Cl 1 g L^{-1}, KCl 0.1 g L^{-1}, KH$_2$PO$_4$ 0,1 g L^{-1} 0.2 g L^{-1} MgSO$_4$. 7H$_2$O, 0.04 g L^{-1} CaCl$_2$2H$_2$O.

F. Experimental Design and Optimization by RSM

The best carbon source, nitrogen source, and salt solution were selected and used for optimized of fermentation medium using Response Surface Methodology. Boundaries of experimental domain and spacing of level expressed in code units are given in Table 1.

TABLE 1. BOUNDARIES OF EXPERIMENTAL DOMAIN AND SPACING OF LEVEL EXPRESSED IN CODE UNITS AND NATURAL UNITS.

Variable	Range and level of experimental				
	-1.68	-1	0	1	1.68
Carbon source (g L^{-1})	21.6	25	30	35	38.4
Nitrogen source (g L^{-1})	6.64	8	10	12	13.36
Salt solution (mL stock solution per volume (liter) of fermentation medium)	3.3	5	7.5	10	11.7

A factorial central composite design (CCD) for 3 factors with six replicates at the central point and six start point, resulting in 20 experiments was used to optimize the screened variables grouped as carbon source (X1), nitrogen source (X2), and mineral salts solution (X3). The behaviour of the system was explained by the following second order polynomial equation:

$$Y = bo + \Sigma \, bixi + \Sigma \, \Sigma \, bij \, xi \, xj + \Sigma \, bii \, x^2i + e \quad ....(1)$$

where : y = predicted response, bo = offset term, bi = linier effect, bii = squared effect, bij = interaction effect.

The Design Expert was used for regression analysis of the data obtained and estimate the coefficients of the regression equation. Response contour plots were also obtained by using Design Expert.

III. RESULT AND DISCUSSION

It is now recognized that the rate at which the carbon source is metabolized can often influence the formation of biomass or production of primary or secondary metabolites. This has been demonstrated the following Table 2.

TABLE 2. EFFECT OF DIFFERENT SUBSTRATES ON CYCLO(TYR-PRO) PRODUCTION BY *Streptomyces* sp. A11.

Carbon source	Concentration of Cyclo(tyr-Pro) (mg L^{-1})	(So-S)/So x 100%	Dry weight cell (g L^{-1})
Lactose	12.84	32.26	3.89
Sucrose	14.05	47.87	4.12
Mollase	15.5	83.47	6.2
Glucose	23	86.05	6.72
Maltose	25.29	79.45	5.63
Dextrin	28.41	78.08	5.45

So = Initial substrate of total sugar, S= final substrate of total sugar

Of the carbon sources tested, dextrin showed the best production of cyclo(tyr-pro). Even though the consume of dextrin lower than glucose, maltose and mollase, dextrin produce cyclo(tyr-pro) higher than others. More glucose consumption will be converted to be biomass. Higher glucose concentration will effect to metabolite repression, and production of secondary metabolite will decrease significant. The next study dextrin will selected as a carbon source for optimization.

As carbon sources, nitrogen also play an important role in cell metabolism. Nitrogen is used as a source of amino acid synthesis such as purine, pyridine, proteins, DNA and RNA. Nitrogen sources also play an important role in the formation of cell biomass in the growth phase, and formation of secondary metabolites, particularly the class of peptide antibiotic. In this investigation organic and inorganic nitrogen source were used. Effect of different nitrogen source on

cyclo(tyr-pro) production by *Streptomyces* sp. A11 is shown on Table 3.

Of the nitrogen source tested, peptone showed the highest cyclo(tyr-pro) production and almost the same with casein and yeast extract. Nitrogen consumption of peptone, casein and yeast extract also the same. Nitrogen source divide into 2 main part, organic nitrogen and inorganic nitrogen. Organic nitrogen usually rich of amino acid. Amino acids can act as a precursor in the formation of secondary metabolites. The next study peptone will be selected as a nitrogen source for optimization.

TABLE 3. EFFECT OF DIFFERENT NITROGEN SOURCE ON CYCLO(TYR-PRO) PRODUCTION BY *Streptomyces* sp. A11

Nitrogen source	Dry weight cell (g.L^{-1})	Concentration of Cyclo(tyr-pro) (mg.L^{-1})	Consumption of nitrogen (mg.mL^{-1})
Glutamic acid (C$_5$H$_9$NO$_4$)	5.21	9.99	0.32
Peptone	6.24	22.7	0.41
Casein	6.01	20.65	0.41
Yeast Extract	6.45	20.88	0.44
Ammonium sulphate (NH$_4$)$_2$SO$_4$	3.21	0	0.2

Salt solution or mineral has a crucial role in enzyme reactions as a cofactor in the metabolic process. Combination of mineral mixture also plays an important role in electrolytic and osmotic regulation in the cell. In this research, several mineral mixture were investigated. The different mixture effect of mineral salt on cyclo(tyr-pro) production by *Streptomyces* sp. A11 is shown on Table 4.

Table 4. showed that mineral I produced the highest cyclo(tyr-pro). The composition of mineral I as followed ;

K_2HPO_4 1 g L^{-1}, MgSO$_4$.7 H$_2$O 0.025 g L^{-1}, ZnSO$_4$ 7 H$_2$O 0.025 g L^{-1}, CaCl$_2$.2 H$_2$O 0.025 g L^{-1}, FeSO$_4$ 7 H$_2$O 0.025 g L^{-1}. According to Stanbury and Whitaker (1987) manganese, iron, and zinc were mineral that the most needed. Low concentration could increase cell growth and product formation, but the excess concentrations would be toxic and cause cell lysis.

TABLE 4. DIFFERENT MIXTURE EFFECT OF MINERAL SALT ON CYCLO(TYR-PRO) PRODUCTION BY *Streptomyces* sp. A11

Mineral salt	Concentration of cyclo(Tyr-Pro) (mg.L^{-1})	Dry weight cell (g.L^{-1})
Mineral I	35.5	6.56
Mineral II	31.68	6.25
Mineral III	33.1	6.34
Mineral IV	31.76	6.28
Mineral V	25.99	5.92

Three variable, dextrin, peptone, and mineral I were selected on basis of highest induced cyclo(tyr-pro) and optimized using central composite design. The experimental and predicted response of cyclo(tyr-pro) production after 5 days cultivation by *Streptomyces* sp. A11 are shown in Table 5. By applying multiple regression analysis on the experimental data, the following second order polynomial equation explain cyclo(tyr-pro) production as follow:

$$Y = 47.40 + 4.13X_1 + 6.74 X_2 + 4.59 X_3 + 2.17 X_1X_2 - 5.59 X_1^2 - 6.78 X_2^2 - 5.88 X_3^2.$$

Y = Cyclo(tyr-pro) (mg L^{-1}),
X_1 = concentration of dextrin (g L^{-1}),
X_2 = concentration of peptone (g L^{-1}),
X_3 = volume of mineral I (mL).

TABLE 5. OBSERVED AND PREDICTED RESPONSE OF PRODUCTION OF CYCLO(TYR-PRO)

No	X1	X2	X3	Observed response/ concentration of cyclo(tyr-pro) (mg L^{-1})	Predicted response
1	-1	-1	-1	19.13	17.19
2	1	-1	-1	19.96	20.15
3	-1	1	-1	23.19	24.64
4	1	1	-1	40.35	36.28
5	-1	-1	1	20.59	23.72
6	1	-1	1	30.99	28.61
7	-1	1	1	35.67	34.54
8	1	1	1	47.11	48.11
9	-1.68	0	0	26	24.65
10	1.68	0	0	35.88	38.56
11	0	-1.68	0	16.74	16.89
12	0	1.68	0	38.37	39.55
13	0	0	-1.68	20.9	23.05
14	0	0	1.68	39.31	38.49
15	0	0	0	47.56	47.4
16	0	0	0	47.71	47.4
17	0	0	0	47.88	47.4
18	0	0	0	46.94	47.4
19	0	0	0	47.19	47.4
20	0	0	0	47.36	47.4

Analysis of variance (ANOVA) for cyclo(tyr-pro) production by *Streptomyces* sp. A11 shown in Table 6.

TABLE 6. ANALYSIS OF VARIANCE (ANOVA) FOR CYCLO(TYR-PRO) PRODUCTION BY *Streptomyces* sp. A11.

Source	Sum of square	Degree of freedom	Mean square	F value	p-value (Prob > F)
Model	2533.49	**9**	281.50	49.83	< 0.0001
A-dextrin	233.30	1	233.30	41.30	< 0.0001
B-peptone	620.13	1	620.13	109.77	< 0.0001
C- mineral	287.79	1	287.79	50.94	< 0.0001
AB	37.71	1	37.71	6.68	0.0272
AC	1.85	1	1.85	0.33	0.5795
BC	5.0	1	5.70	1.01	0.3390
A^2	449.57	1	449.57	79.58	< 0.0001
B^2	662.88	1	662.88	117.33	< 0.0001
C^2	498.35	1	498.35	88.21	< 0.0001
Residual	56.50	**10**	5.65		
Total	2589.98	19			

$R^2 = 0.98$; adj $R^2 = 0.96$; CV= 6.8%

The analysis of variance of quadratic regression model demonstrates that the model is higly significant (p-value/Prob>F) <0.0001. The closer the value of R (multiple correlation coefficient) to 1, the better the correlation between the observed and predicted value. The coefficient of variation (CV) indicates the degree of precision with which the treatments are compared. Usually, the higher the value of CV, the lower is the reliability of experiment. In this experiment, a lower value of CV indicates a greater reliability of the experiments performed. The parameter estimate and the corresponding P value suggest that among the independent variables dextrin (X1), peptone (X2), and Mineral I (X3) have significant effect on cyclo(tyr-pro) production. The quadratic term of these three variables also have a significant effect. There are no interaction between dextrin to mineral and peptone to mineral but there is interaction between dextrin to peptone. Response surface plot obtained as function of dextrin concentration versus peptone concentration and other variable are at zero level. A maximum increase in cyclo(tyr-pro) yield with increase in dextrin concentration versus peptone concentration was observed (Fig. 1).
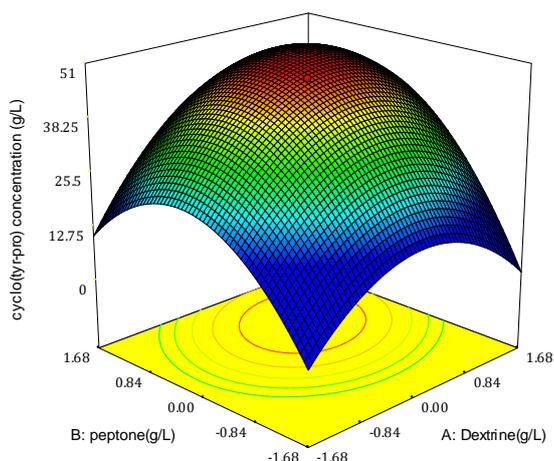

Fig. 1. Response surface plot showing the effect on dextrin concentration , peptone concentration and their mutual effect on the production of cyclo(tyr-pro). Other variables are held at zero level.

Dextrin and peptone concentration increases significantly and influence the production of cyclo(tyr-pro). Increasing the concentration of peptone looks more influence on the production of cyclo(tyr-pro) compared the increasing of the dextrin concentration. There is interaction between dextrin to peptone.

Response surface plot obtained as function of dextrin concentration versus mineral salt and other variable are held at zero level. A maximum increase in cyclo(tyr-pro) yield with increase in dextrin concentration versus mineral salt was observed (Fig. 2).
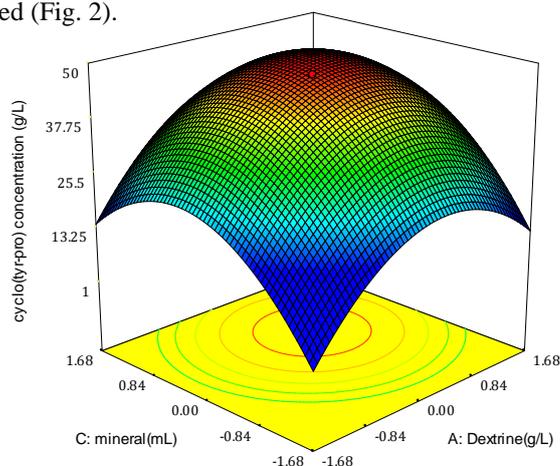

Fig. 2. Response surface plot showing the effect on dextrin concentration, mineral salt concentration and their mutual effect on the production of cyclo(tyr-pro). Other variables are held at zero level.

Dextrin and mineral salt concentration increases significantly and influence the production of cyclo(tyr-pro). Increasing the dextrin concentration looks the same influence on the production of cyclo(tyr-pro) compared the increasing of the mineral salt concentration, but there is no interaction between dextrin to mineral salt.

Response surface plot obtained as function of peptone concentration versus mineral salt and other variable are held at zero level. A maximum increase in cyclo(tyr-pro) yield with increase in peptone concentration versus mineral salt was observed (Fig. 3).
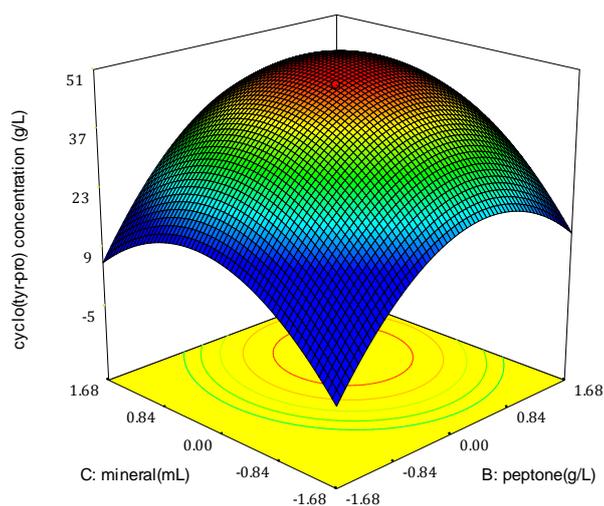
Fig. 3. Response surface plot showing the effect on peptone concentration, mineral salt concentration and their mutual effect on the production of cyclo(tyr-pro). Other variables are held at zero level.

Peptone and mineral salt concentration increases significantly and influence the production of cyclo(tyr-pro). Increasing the peptone concentration looks more influence on the production of cyclo (tyr-pro) compared the increasing of the mineral salt concentration, but there is no interaction between peptone to mineral salt.

The regression equation (equation 1) was solved by Design Expert. The optimal values of test variables in coded unit were $X_1 = 0.51$; $X_2 = 0.61$; $X_3 = 0.46$. At the these values, the concentration of dextrin and peptone were 32.55 g L^{-1}, 11.22 g L^{-1} respectively, and volume increase of mineral salt was 8.65 mL. The maximum predictes value of cyclo(tyr-pro) production was 51.54 mg L^{-1}. A repeated fermentation of cyclo(tyr-pro) under optimal condition was carried out in laboratory. The maximal of cyclo(tyr-pro) was obtained 50.04 mg L^{-1}. This value was found to be 2.9% less than the predicted value. The discrepancy might due to the slight variation in experimental conditions.

IV. CONCLUSION

Determination of the best production medium of cyclo(tyr-pro) showed dextrin, peptone, and the mineral salts group I produced the highest cyclo(tyr-pro). The combination concentration of dextrin, peptone were 32.55 g L^{-1}, 11.22 g L^{-1} respectively, and volume increase of mineral salt was 8.65 mL was obtained the highest cyclo(tyr-pro) concentration 50.04 mg L^{-1}. The differences of experiment with predicted

value was 2.91%. Using the optimized parameters, the studies demonstrated an increase in the cyclo(tyr-pro) was 2.5 fold from 20 mg L^{-1} to 50 g L^{-1}.

REFERENCE

[1] A. Stierle, J.H. Cardellina, G.A. Strobel, "Maculosin, a host-specific phytotoxin for spotted knapweed from *Alternaria alternata*," *Proc Nat Acad Sci*., vol. 85(21), pp. 8008-8011.1988.

[2] R. Sunaryanto, B. Marwoto, T.T. Irawadi, Z.A. Mas'ud, L. Hartoto, "Isolation and characterization of antimicrobial substance from marine *Streptomyces* sp, " *J Microbiol Indonesia*., vol.4(2), pp. 84-89, 2010.

[3] Q. Guo, G. Daosen, B. Zhao, J. Xu, R.Li, "Two cyclic dipeptides from *Pseudomonas fluorescens* GcM5-1A carried by the pine wood nematode and their toxicities to Japanese black pine suspension cells and seedlings in vitro," *J Nematol*., vol. 39(3), pp. 243–247, 2007.

[4] G.E.P. Box, W.G. Hunter, J.S. Hunter, *Statistics for Experimenters. an Introduction to Design, Data Analysis and Model Building*, New York: John Wiley and Sons,1978.

[5] D.C. Montgomery, *Design and Analysis of Experiments*. 4th ed., New York: John Wiley and Sons, 1997.

[6] G.E.P. Box and N.R. Draper, *Empirical Model Building and Response Surface,* New York: John Wiley & Sons,1987.

[7] O. Popa, B. Narcisa, A. Vamanu, E. Namanu, "The utilization of the response surface methodology for the optimization of cultivation medium and growth parameters in the cultivation of the yeast strain *S cerevisiae* 3.2 on ethanol," *Afr J Biotechnol*., vol. 6(23), pp. 2700-2707, 2007.

[8] G.Q. He, Q. Kong, L.X. Ding, "Response surface methodology for optimizing the fermentation medium of *Clostridium butyricum*," *Lett App Microbiol*., vol. 39. pp 363-368, 2004.

[9] J. Sathyanarayanan, J. Kunthala, K. Gurumurthy, "Optimization of MRS media components using response surface methodology for the riboflavin production by *Lactobacillus fermentum* isolated from yogurt sample," *Int Food Res J*., vol. 18, pp. 149-159, 2011.

[10] L.H. Man, S.K. Behera, H.s. Park, "Optimization of operational parameters for ethanol production from Korean food waste leachete," *Int J Environ Sci Tech*., vol. 7(1), pp. 157-164, 2010.

[11] K. Kanoh, Y. Matsuo, K. Adachi, K. Imagawa, M. Nishizawa, Y. Shizuri, "Mechercharmycins A and B, cytotoxic substances from marine-derived *Thermoactinomyces* sp. YM3-251," *J Antibiot.,* vol. 58(4), pp. 289–292, 2005.

[12] M.F.V.Q. Sousa, C.E. Lopes, N.P. Junior, "A chemically defined medium production of actinomycin D by S*treptomyces parvulus*," *Brazilian arch biol technol.,* vol. 44, pp. 227-231, 2001.

[13] N.A.J.C. Furtado, M.T. Pupo, I.Carvalhho, V.L. Campo, M.C.T. Duarte, J.K. Bastos, "Diketopiperazines produced by an *Aspergillus fumigatus* Brazillian strain," *Braz Chem Soc.,* vol. 16, pp. 1448-1453, 2005.

[14] V. Dhananjeyan, N. Selvan, K. Dhanapal, "Isolation, characterization, screening and antibiotic sensitivity of actinomycetes from locally (Near MCAS) collected soil samples, " *J Biol Sci*., vol. 10(6), pp. 514-519, 2010.

[15] Voelker and Altaba, "Nitrogen source governs the pattern of growth and prostinamycein production in *Streptomyces pristinaespiralis*,". *Microbiol*., vol. 147, pp. 2447-2459, 2001

[16] S. Dharmaraj, B. Ashokkumar, K. Dhevendaran, "Isolation of marine *Streptomyces* and the evaluation of its bioactive potential," *Afr J Microbiol Res*., vol. 4(4), pp. 240-248, 2010.