Nephroprotective activities of the aqueous seed extract of *Carica papaya* Linn. in carbon tetrachloride induced renal injured Wistar rats: a dose- and time-dependent study

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Abstract

In the present study, the dose related effect of the aqueous seed extract of Carica papaya Linn. extract (CPE) was evaluated by pre-treating three groups of rats (made up of six male rats per group) with 100 - 400 mg/kg body weight per oral of the extract for 7 days before challenging with 1.5 ml/kg body weight of 20% carbon tetrachloride in olive oil in addition to the untreated control and model control rats. Also, the time-course effect of 400 mg/kg per oral of the extract were determined at 3 hr pre-, 0 hr, 1 hr post-, 3 hr post-, and 6 hr post-CCl₄ induction, respectively, in addition to the untreated control and model control groups. After 72 hours, serum levels of uric acid, urea and creatinine of all study groups were measured using standard procedures. Histological studies of rat kidneys of all study groups were also done. Results showed that intraperitoneal injection of CCl₄ caused a significant (p<0.001) elevation in the serum levels of uric acid, urea and creatinine and induced histological features of severe tubulointerstitial necrosis. However, elevations in the measured biochemical parameters were significantly (p<0.05, p<0.01 and p<0.001) attenuated in rats pre-treated with the graded oral doses of the extract, in dose related fashion. Maximum nephroprotection was offered by the extract at 400 mg/kg/day CPE which lasted up to 3 hours post-CCl₄ exposure and these biochemical evidences were corroborated by improvements in the renal histological lesions induced by CCl₄ intoxication. In conclusion, our study showed that CPE has nephroprotective effect on CCl₄ renal injured rats, an effect which could be mediated by any of the phytocomponents present in it via either antioxidant and/or free radical scavenging mechanism(s).

Keywords: Carica papaya Linn.; Aqueous seed extract; Nephroprotection; Carbon tetrachloride; Wistar rats.

Introduction

Carica papaya Linn. (Family: Caricaceae) is a perennial, herbaceous plant, with copious milky latex reaching to 6 - 10 meters tall. Its erect stem is about 30 cm thick and roughened with leaf scars (Duke, 1984). The unripe fruit is used traditionally among the Yoruba tribe of Nigeria for the treatment of various human and veterinary diseases including malaria, hypertension, diabetes mellitus, hypercholesterolaemia, jaundice, intestinal helminthiasis (Gill, 1992) and for the management of sickle cell anaemia (Ogunyemi et al., 2008). Also, among the Yoruba herbalists, hot infusion of the seeds of the unripe, mature fruits of Carica papaya is employed in the local treatment of poison related renal and hepatic disorders. Based on the traditional use of Carica papaya seed in the treatment of poison related renal disease, the present study was designed to investigate the effect of graded oral doses (100 - 400 mg/kg/day) and the time-course effect of the aqueous seed extract of mature unripe *Carica papaya* (CP*E*) in CCl₄ treated rats as a way of validating this folkloric use. The choice of the extract dose range employed in this was based on the result obtained from the orientation study earlier conducted.

Materials and Methods

Plant extract

Ten mature, unripe fruits of *Carica papaya* were collected from a cultivated Pawpaw Plantation at Oke-Afa, Isolo, Lagos State, Nigeria, in the first week of May, 2008. Plant identification and authentication had earlier been done by Olagunju *et al.* (1995). The pawpaw

fruits were cut into pieces and the wet seeds were separated out. These were then gently but thoroughly rinsed in tap water for two times and completely airdried at room temperature for 4 weeks. The dried seeds were pulverized into fine powder using a new domestic mixer grinder (Kanchan Tycoon[®], Kanchan International Limited Unit III, Daman, India).

The dried aqueous seed extract of the unripe mature fruits of Carica papaya was obtained as follows: 40 g of the powdered dried seeds of Carica papaya was boiled in 500 ml of distilled water for 30 minutes, after which it was rapidly filtered through a piece of clean cotton gauze. The filtrate was allowed to cool at room temperature for 4 hours after which it was completely oven-dried at a preset temperature of 40 °C, producing a fine, aromatic, and chocolate colour solid residue. The procedure was repeated 4 times with this same quantity of solvent [vield: $22.0 \pm 0.5\%$ (w/w)]. The dry residues obtained were weighed, pooled and stored in air- and water-proof container kept in a refrigerator at 4 °C. From this stock, fresh preparation was made whenever required.

Experimental animals

Male Wistar rats weighing between 150 and 180 g were used for investigating the nephroprotective effect *in vivo* on CCl₄induced nephrotoxic rats. All animals were conditioned in standard metallic cages (6 rats per cage), fed standard laboratory diet (Ladokun Animal Feeds, Ibadan, Nigeria) *ad libitum* and allowed free access to drinking water. The animals were also kept in 12:12 hour light/dark cycle. The experimental rats were handled in strict compliance with international guidelines as prescribed by the Canadian Council on the Care and Use of Laboratory Animals in Biomedical Research (1984).

Determination of the dose-related protective effect of the extract

To determine the dose of CP*E* necessary for the maximum nephroprotection, three different groups of Wistar rats were separately treated with 100 mg/kg, 200 mg/kg and 400 mg/kg of CP*E* in addition to the untreated control and model control groups, for 7 days prior to single intraperitoneal CCl₄ treatment at a dose of 1.5 ml/kg body weight of 20% CCl₄ in olive oil. Seventy-two hours post-induction, the rats were sacrificed under inhaled diethyl ether anaesthesia.

Determination of the time-course effect of the extract

To determine the time required for the CPE to exhibit nephroprotection, seven groups of rats were separately treated with the extract at 400 mg/kg of CPE at 3 hours pre-, 0 hour, 1 hour post-, 3 hours post-and 6 hours post-CCl₄ induction, respectively.

Experimental induction of nephrotoxicity

CCl₄-induced acute renal injury was initiated by intraperitoneal injection of 1.5 ml/kg of 20% CCl₄ (Merck, Darmstadt, Germany) dissolved in olive oil (Roberts Laboratories Limited, Belton, England) as described by Lu *et al.* (2002).

CCL

For the oral graded dose model, CCl₄ was intraperitoneally injected into groups II - V fasted rats 24 hours after the last oral dose of CPE on the 7th day while group I rats (untreated control) received 10 ml/kg/oral route for 7 days before intraperitoneal injection of 1 ml/kg of distilled water. Rat feed and potable water were further withheld for additional 4 hours before the rats were allowed free access to feed and water. Similarly, for the time course model. same dose of the nephrotoxicant was administered intraperitoneally except that the toxicant was injected at specified time intervals of 3 hours pre-, 0 hour, 1 hour post-, 3 hours post- and 6 hours post-CCl₄ induction.

Assessment of serum uric acid, urea and creatinine

Blood samples, obtained directly from the heart chamber of the anaesthetized rats were kept at the temperature of 4 °C for 6 hours before they were centrifuged using Uniscope Laboratory Centrifuge (Model SM 902B, Surgifriend Medicals, England, U.K.) at 3000 rpm at same temperature for 20 minutes in order to separate the sera. The urea and creatinine levels in all the sample sera were estimated by modified methods based on diacetylmonoxime reaction (Marsh et al., 1965) and Jaffe's (Biod reaction and Sirota. 1948). respectively, on standard diagnostic test kits (Randox Laboratories, Crumlin, U.K.) on Automated Clinical System (Sychron Clinical System[®], model: CX5 PRO) (Beckman Coulter Inc., Galway, Ireland).

Histopathological studies of the rat kidneys After all the animals were sacrificed, postmortem examination was performed on all the identified and dissected rat kidneys. The kidneys were rinsed in normal saline and sections were taken from them. The renal tissue sections obtained were fixed in 10% neutral buffered formalin, dehydrated with 100% ethanol solution and embedded in paraffin. Tissue sections of 5 µm in thickness were prepared, stained with haematoxylin and eosin (H & E) and observed under a photomicroscope (Model N - 400ME, CEL-TECH Diagnostics, Hamburg, Germany).

Statistical analysis

Data were expressed as mean \pm SEM of six observations and statistically assessed by two-way analysis of variance and the group means were compared using Student's t-test on statistical software program, SYSTAT 10.6. A probability of p<0.05, p<0.01 and p<0.001 were considered as significant.

Results

Dose-dependent preventive activity of CPE in the CCl_4 renal injured rats

Table 1 shows the effect of CPE on the serum levels of urea and creatinine in the CCl₄ injured rats. As shown in table 1, intraperitoneal injection of 1.5 ml/kg of 20% CCl₄ in olive oil caused significant (p<0.001) rise in the serum levels of urea and creatinine as evident in the model control (group II rats). These elevations significantly (p<0.05, were p<0.01. p<0.001) attenuated in rats pretreated with the extract in dose related fashion. However, maximum nephroprotection was offered by the 400 mg/kg of CRE pretreatment.

Time-course protective activity of CPE in the CCl₄ renal injured rats

Table 2 shows the time-dependent preventive role of 400 mg/kg body weight/oral route of CPE against CCl₄ nephrotoxicity. As shown in table 2, CPE at the oral dose of 400 mg/kg body weight offered significant (p<0.05) protection for up to 3 hours post-exposure to CCl₄ injection.

Table 1: Dose-dependent activities of 100 – 400 mg/kg of CPE on serum uric acid, urea and creatinine in CCl₄ renal injured rats.

Treatment group	s	Serum concentration (mg/dl) of		
	Uric acid	Urea	Creatinine	
	2.9 ± 0.2	29.0 ± 3.8	0.9 ± 0.1	
	3.8 ± 0.2^{a}	$42.3 \pm 3.5^{\circ}$	1.4 ± 0.2^{a}	
	3.2 ± 0.2	33.3 ± 2.0^{d}	1.0 ± 0.1	
IV 🖉	3.2 ± 0.3^{d}	29.0 ± 1.1 ^e	1.1 ± 0.1	
V	2.3 ± 0.1^{f}	23.7 ± 1.4^{f}	0.9 ± 0.2^{d}	

^a and ^c represent significant increases at p<0.05 and p<0.01 when compared to group I (untreated control) values while ^{d, e} and ^f represent significant decreases at p<0.05, p<0.01 and p<0.001 when compared with group II (model control) values.

I = 10 ml/kg per oral of DW + 1 ml/kg/IP of DW $II = 10 \text{ ml/kg per oral of DW} + 1.5 \text{ ml/kg/IP of 20\% CCl}_4 \text{ in olive oil}$ $III = 400 \text{ mg/kg of CP}E + 1.5 \text{ ml/kg/IP of 20\% CCl}_4 \text{ in olive oil}$ $IV = 200 \text{ mg/kg of CP}E + 1.5 \text{ ml/kg/IP of 20\% CCl}_4 \text{ in olive oil}$ $V = 400 \text{ mg/kg of CP}E + 1.5 \text{ ml/kg/IP of 20\% CCl}_4 \text{ in olive oil}$ CPE = Aqueous seed extract of Carica papaya IP = intraperitoneal route $CCl_4 = \text{carbon tetrachloride}$ DW = distilled water

Treatment groups	Serum concentration (mg/dl) of			
	Uric acid	Urea	Creatinine	
	2.9 ± 0.2	29.0 ± 3.8	0.9 ± 0.1	
II	3.8 ± 0.2^{a}	$42.3 \pm 3.5^{\circ}$	1.4 ± 0.2^{b}	
VIII	4.1 ± 0.3^{a}	38.8 ± 3.3^{a}	1.1 ± 0.1 ^b	
IX	2.9 ± 0.3^{b}	28.8 ± 3.1 ^d	0.6 ± 0.2^{f}	
Х	3.4 ± 0.3	26.3 ± 1.5 ^e	1.2 ± 0.1^{d}	
XI	3.0 ± 0.3^{d}	$34.5 \pm 1.7^{\circ}$	1.0 ± 0.1^{d}	
XII	3.6 ± 0.6	$42.3 \pm 6.4^{\circ}$	1.5 ± 0.3⁵	

Table 2: Time-course activities of 400 mg/kg of CPE on serum uric acid, u	irea and
creatinine in CCl₄ renal injured rats.	

^a and ^c represent significant increases at p<0.05 and p<0.001, respectively, when compared to group I (untreated control) values while ^{d, e} and ^f represent significant decreases at p<0.05, p<0.01 and p<0.001, respectively, when compared to group II (model control) values.

I = 10 ml/kg per oral of DW + 1 ml/kg/IP of DW $II = 10 \text{ ml/kg per oral of DW} + 1.5 \text{ ml/kg/IP of 20\% CCl}_4 \text{ in olive oil}$ $VIII = 400 \text{ mg/kg of CP}E + 3 \text{ hr pre-CCl}_4 \text{ induction}$ $IX = 400 \text{ mg/kg of CP}E + 0 \text{ hr of CCl}_4 \text{ induction}$ $X = 400 \text{ mg/kg of CP}E + 3 \text{ hr post-CCl}_4 \text{ induction}$ $XI = 400 \text{ mg/kg of CP}E + 3 \text{ hr post-CCl}_4 \text{ induction}$ $XI = 400 \text{ mg/kg of CP}E + 6 \text{ hr post-CCl}_4 \text{ induction}$ CPE = Aqueous seed extract of Carica papaya IP = intraperitoneal route $CCl_4 = \text{ carbon tetrachloride}$ DW = distilled water

Histological studies of the dose- and timedependent effects of CPE on CCl₄-treated kidneys

1, 2, **3, 4** and 5 Figures are photomicrographs of sections of normal rat kidney, CCl₄-treated kidney, 100 mg/kg/day CPE, 200 mg/kg/day CPE and 400 mg/kg/day CPE pretreated rat kidneys, respectively. As shown in the photomicrographs, CCl₄ renal intoxication was associated with severe glomerular and tubulo-interstitial necrosis which was characterized by hydropic degeneration of the glomerular and tubular cells with complete obliteration of the tubular lumen (from hydropic degeneration and tubular casts) (figure 2) when compared to normal rat kidney (figure 1). However, oral pretreatments with 100 - 400 mg/kg/day CP*E* ameliorated renal histological lesions in dose related fashion (figures 3 - 5) with the least renal architecturally-affected being the 400 mg/kg/day CP*E* treated group (figure 5).

Similarly, figures 6 - 10 represent the timecourse effect of oral treatments in CCl₄treated rat kidnevs. As shown in the figures, 400 mg/kg/day CPE treatments at 0 hr, 1 hr post-, and 3 hr post- CCl₄ prevented marked renal intoxication histological lesions of hydropic degeneration of the glomerular and tubular cells (figures 7 - 9) when compared to those seen in the 3 hr pre-, and 6 hr post-CCl₄ induction treated groups (figures 6 and 10).



Figure 1







Figure 3



Figure 4









Figure 7







Figure 9



Figure 10

Figure 1: A sectional representation of normal rat kidney at x400 magnification (Haematoxylin and Eosin stain) showing normal glomeruli (**GL**) with an intact Bowman's capsule (**BM**), proximal convoluted (**CT**) and distal convoluted (**DT**) tubules

Figure 2: A representative section of CCl₄–intoxicated rat kidney at x400 magnification (Haematoxylin and Eosin) showing severe hydropic glomerular degeneration (**GLH**), obliterated proximal convoluted tubular lumen (**OPC**) and obliterated distal convoluted tubular lumen (**OPC**). The tubular lumens were completely obliterated and filled with fluid and casts

Figure 3: A representative section of 100 mg/kg/day CP*E* pretreated, CCl₄–intoxicated rat kidney at x400 magnification (Haematoxylin and Eosin) showing moderate hydropic glomerular (**GLM**) and tubular degenerations (**TD**). The tubular lumens were moderately filled with fluid and casts (**MTC**)

Figure 4: A sectional representation of 200 mg/kg/day CP*E* pretreated, CCl_4 -intoxicated rat kidney at x400 magnification (Haematoxylin and Eosin) showing mesengial proliferation (**MP**) with thinning out of the Bowman's capsule (**BM**). There is mild tubular cast deposition (**MTC**) interposed with normal proximal convoluted tubule (**PC**) and distal convoluted tubule (**DC**)

Figure 5: A sectional representation of 400 mg/kg/day CP*E* pretreated, CCl₄-intoxicated rat kidney at x400 magnification (Haematoxylin and Eosin) showing normal glomeruli (**GL**) encapsulated by normal Bowman's capsule (**BM**). There is no obvious tubular cast deposition

Figure 6: A sectional representation of 400 mg/kg/day CPE3 hr pretreated, CCl₄--intoxicated rat kidney at x400 magnification (Haematoxylin and Eosin) showing moderate tubular degeneration (**TD**) with normal glomeruli (**GL**) and Bowman's capsule (**BM**).

Figure 7: A sectional representation of 400 mg/kg/day CP*E* 0 hr pretreated, CCl₄–intoxicated rat kidney at x400 magnification (Haematoxylin and Eosin) showing normal glomeruli (**GL**) encapsulated in normal Bowman's capsule (**BM**). There is mild tubular degeneration (**TD**) and tubular cast deposition (**MTC**) interposed with normal proximal convoluted tubule (**PC**) and distal convoluted tubule (**DC**)

Figure 8: represents the kidney section (x400 magnification, Haematoxylin and Eosin stain) of 400 mg/kg/day CPE 1 hr post- CCl_4 treatment showing some degree of mesengial proliferation (**MP**) with mild hydropic tubular degeneration (**TD**) and mild tubular cast (**MTC**) interposed with normal proximal convoluted tubules (**PC**) and distal convoluted tubules (**DC**)

Figure 9: represents the kidney section (x 400 magnification, Haematoxylin and Eosin stain) of 400 mg/kg/day CPE 3 hr post-CCl₄ treatment showing moderate hydropic glomerular degeneration (**GLM**), tubular degeneration (**TD**) and moderate tubular cast (**MTC**) and tubular obliteration (**OTL**) interposed with normal proximal convoluted tubules (**PC**) and distal convoluted tubules (**DC**)

Figure 10: A section of rat kidney (x400 magnification, Haematoxylin and eosin stain) treated with 400 mg/kg/day CP*E* 6 hr post-CCl₄ treatment showing severe glomerular degeneration (**GLM**) and severe hydropic tubular degeneration (**TD**) and severe tubular cast (**MTC**) and obliteration of the tubular lumen (**OTL**)

Discussion

Various environmental toxicants and clinically useful drugs, like acetaminophen and gentamicin, can cause severe organ toxicities through the metabolic activation to highly reactive free radicals including the superoxides and oxygen reactive species (Abraham and Wilfred, 1999). One of the most extensively studied of the environmental toxicants is carbon tetrachloride (CCl₄). CCl₄ is known to undergo reductive metabolism by CYP2E1 into a highly reactive trichloromethyl radical (.CCl₃) and phospene that initiates lipid peroxidation, disrupts membrane integrity and causes cell death (Pohl et al., 1984; Fadhel and Amran, 2002; Basu, 2003). Evidence suggests that various enzymatic and non-enzymatic systems have been developed by the cell to cope with the oxidative stress that is associated with reactive oxygen species (ROS) and other free radicals generated (Tom et al., 1984). Literature shows that oxidative stress has been implicated in the aetiopathogenesis of CCl₄ organ injury and carcinogenesis (Slater, 1984; Stal and Olston, 2000). However, when the oxidative stress is overwhelming, the various inherent defence mechanisms antioxidant (such as the defence mechanisms, intracellular concentration of glutathione, superoxide dismutase (SOD) and catalase (CAT) activities become significantly impaired and insufficient (Szymonik-Lesiuk et al., 2003).

As a measure of renal function status, serum urea and creatinine are often regarded as reliable markers (Adelman et al., 1981). Thus, elevations in the serum concentrations of these markers are indicative of renal injury (Adelman et al., 1981). In the present study, the doseand time-dependent nephroprotective effect of the aqueous extract of the unripe seeds of Carica papaya were evaluated in CCl₄ induced acute nephrotoxic rats by assessing the effect of the extract pretreatments on the serum uric acid, urea and creatinine in the CCl₄ injected renal injured rats. In addition, the associated histological lesions were studied. In the present study, it was observed that treatment with CCl₄ induced a significant elevation in the levels of serum uric acid, urea and creatinine within 72 hours of These biochemical exposure to it. alterations were corroborated by the

histological findings of glomerular and tubulo-interstitial necrosis in the untreated model control group. However, daily pretreatment with CPE for 7 days conferred nephroprotection on the CCl₄ renal injured rats in a dose-dependent fashion and 400 mg/kg dose offered maximum protection within 3 hours of exposure to it. Again, the histological findings of almost normal renal histological architecture corroborate the protection conferred by the extract within the stipulated time interval, especially at the maximum oral dose of 400 mg/kg/day of the extract. Although, the possible mechanism(s) of its protection against CCl₄ induced nephrotoxicity was not studied in the current study, it is possible that the protective effect of the extract is mediated through antioxidant and/or free radical scavenging activities. Literature shown medicinal plants has with nephroprotective properties to mediate their protection via antioxidant and/or free radical scavenging activities due to the high concentration of flavonoids and alkaloids they contain (Miller and Rice-Evans, 1997; Adeneye and Benebo, 2008). Equally, saponins have been reported to protect liver and kidneys against carbon tetrachloride intoxication (Jeong et al., 1996). In addition, CPE has been reported to contain flavonoids, alkaloids, saponins and other active phytocomponents (Adeneve and Olagunju, In Press). Summing these facts, it is plausible for the alkaloid, flavonoid and saponin components of CPE to be responsible for the observed biological effects. These could constitute areas of the future research. Again, nephroprotection offered by the extract could be due to the presence of any of the phyto-principles contained in it.

In conclusion, it is proposed that the nephroprotective activities of the aqueous seed extract of the unripe, mature fruits of *Carica papaya* Linn. in carbon tetrachloride-induced nephrotoxicity may involve its antioxidant and/or oxidative free radical scavenging activities. Also, the results of this study have confirmed the rationale for the folkloric use of the aqueous seed extract of *Carica papaya* Linn. in the treatment of poison-related renal disorders.

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