

## Low dose Flaxseed Oil Supplementation Alters the Fatty Acids Profile and the Progression of Metabolic Syndrome in Men without Adequate Medical Treatment

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### Abstract

Many studies indicated that increased intakes of  $\omega$ -3 fatty acids could positively impact the progression of metabolic syndrome (MS). This study aimed to characterize the clinical and biochemical changes initiated by a low-dose flaxseed oil supplementation upon the evolution of metabolic syndrome in men without adequate medical treatment.

In a double blind, randomized study, middle-aged men with metabolic syndrome, who were not able to follow the prescribed medical treatment, were assigned to either a group receiving daily 2.4 g flaxseed oil, or the same amount of corn oil, for 90 days, respectively. Analysis of variance, logistic, and bivariate fit analyses were used to describe the statistical significance of parameters changed by either treatment (within and between group comparisons), between the start and end of treatment. While none of the five diagnostic criteria for MS were differently altered between groups and time points, changes in body mass index (BMI) and insulin resistance were significantly correlated with the treatment received. Subjects receiving flaxseed oil registered no increase in BMI, as compared to an increased BMI registered in the corn oil group ( $+1.12 \pm 0.63$ ,  $p < 0.05$ ). Bivariate fit for plasma insulin and derived HOMA index indicated that flaxseed oil maintained the individual correlation of these parameters between the start and end of study, while corn oil supplementation was associated with an increase in insulin resistance with no individual correlation between start and end of treatment ( $1.12 \pm 0.17$ ,  $p < 0.05$  vs.  $2.11 \pm 0.79$ ,  $p > 0.05$  ratios between start and end of study, respectively).

The analysis of total serum fatty acid profiles indicated, among other changes, significance for time-treatment interaction for serum 11-eicosenoic acid ( $p < 0.05$ ). Other correlations on inflammation markers associated with MS are reported. In conclusion, low daily doses of flaxseed oil may improve clinical and metabolic parameters in middle-aged men without adequate treatment for metabolic syndrome.

**Keywords:** Metabolic syndrome; Flaxseed oil; Body weight; Insulin resistance

### Introduction

Metabolic syndrome (MS) is a common metabolic disorder with increasing prevalence all over the world [1]. MS is directly correlated with the development of obesity, and represents a major contributor in the development of cardiovascular diseases and type II diabetes [2]. Moreover, there is a significant overlap between MS and conditions such as polycystic ovarian syndrome, non-alcoholic fatty liver disease, hypogonadism, lipodystrophy, and microvascular disease [1]. The treatment of MS is complex and includes both lifestyle changes (physical activity and nutrition) and drug therapy [3]. However, while the drug therapy is relatively accessible in developed countries, MS patients in underdeveloped and developing countries often do not have adequate access to medical care, or their poor socio-economic status does not allow them to purchase and follow the prescribed medication [4].

Human and animal studies indicated that increased dietary intakes or supplementation with marine or plant derived  $\omega$ -3 fatty acids are inversely correlated with the presence of MS [5]. While the benefits of  $\omega$ -3 fatty acids have been amply supported by numerous studies, their mechanism of action is not completely understood, nor has a definitive answer been given regarding the dietary intakes necessary to attain such protective effects [6]. Polyunsaturated n-3 fatty acids may improve defects in insulin signaling, and prevent alterations in glucose homeostasis and the further development of type II diabetes [7-8]. Diets rich in  $\alpha$ -linolenic acid (ALA) also appear to be beneficial in secondary cardiovascular prevention [9]. However, there is conflicting evidence regarding the specific role of ALA, eicosapentaenoic (EPA),

and docosahexaenoic (DHA) acids on decreasing insulin resistance, and regarding their influence on inflammatory cytokine modulation [10].

The aim of this study was to determine the consequences that low-dose supplementation with flaxseed oil may have upon clinical and biochemical outcomes in subjects with MS who did not receive proper medical treatment.

### Materials and Methods

The study was performed according to the Declaration of Helsinki and approved by the Institutional Review Board at the University of North Carolina at Chapel Hill, and by the Ethics Committee at Transilvania University. Written consent was obtained from all participants before study initiation.

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## Study population

The study was a double-blind, controlled, randomized intervention trial in 20 male volunteer subjects, between 50-65 years of age, with diagnosed MS, recruited from Romania (Brasov area) between August 2008 and September 2009. MS was assessed according to the National Cholesterol Education Program's Adult Treatment Panel III report criteria (ATP III), for the presence of three or more of the following criteria: 1) waist circumference  $\geq 40$  inches (102 cm), 2) triglycerides (TG)  $\geq 150$  mg/dl, 3) HDL-cholesterol  $\leq 40$  mg/dl, 4) blood pressure (BP, systolic/diastolic)  $\geq 130/\geq 85$  mmHg or antihypertensive treatment, and 5) fasting glucose (FG)  $\geq 110$  mg/dl or antidiabetic medication (insulin or oral agents) [11].

Exclusion criteria consisted of: 1) any coagulation syndrome and/or diagnosed blood disorder related to the number, morphological features, or physiology of red blood cells (RBC), white blood cells (WBC), or platelets, 2) family history of hereditary blood disorder, even in the absence of a diagnosed blood disorder, any carrier of a diagnosed genotype known to primarily cause a blood disorder, 3) history or presence of an embolic event (any organ), stroke, transient ischemic attack (TIA), ischemic heart disease, myocardial infarction, unstable angina, or varicose veins, 4) current use of a treatment or dietary supplements (for the past three months) containing any omega-3 fatty acid or any lipid modifying agent (e.g. statins, fibrates, ezetrol), and 5) any treatment including the administration of aspirin, ticlopidine/clopidogrel, non-steroidal anti-inflammatory drugs, and other hypocoagulant medication before and during the study.

## Study design

Upon enrollment, the Brasov team assigned a number (from 1 to 20) to each enrolled subject. Following enrollment, the Chapel Hill team randomly assigned each of the subjects to either a group receiving corn oil (C group, n=10), or to a group receiving flaxseed oil (F group, n=10). Due to the small number of subjects, stratification prior to randomization was done only for BMI and age. The characteristics of subjects assigned to each group are presented in Table 1. Identical containers containing capsules with either 1.2 g of either corn oil or flaxseed oil were numbered by the Chapel Hill team according to each subject's assigned number, and shipped to Brasov. The lipid content of corn oil and flaxseed oil capsules is indicated in Table 2. The local team was blinded against both the content of capsules and group assignment. Each subject was instructed to consume two capsules a day for 90 days, and to not eat any food or take dietary supplements containing significant amounts of omega-3 fatty acids (fish, seeds, nuts, etc.). Compliance was calculated based on the formula: number capsules taken/number capsules needed to be taken x 100. Compliance over 80% was targeted and achieved in both groups.

## Measurements

Clinical and biochemical investigations were assessed on days 0 (D0, one day before the beginning of supplementation) and 90 (D90, end of study) for each subject. Safety monitoring visits were performed monthly during this period and consisted of clinical examination, blood pressure measurements, distribution of capsules for the next month, and evaluation of compliance. At D0 and D90, clinical evaluation included anthropometric and blood pressure measurements. Anthropometric measurements were performed under a fasting state with subjects wearing light clothing and no shoes. The waist circumference was measured at the high point of the iliac crest at minimal respiration to the nearest 0.1 cm. BMI was calculated as kg/m<sup>2</sup>.

For each study day, blood samples were collected by venipuncture

in the morning following an overnight fast, using both 8 mL CPT tubes and 8 mL tubes with heparin. Mononucleate cells were obtained using the CPT tubes, stored in RNA stabilization reagent (RNAlater, Qiagen, Germany) and shipped to Chapel Hill for further assessment. Plasma was obtained using heparinized tubes and used for subsequent determinations, or shipped to Chapel Hill for fatty acids measurements. In addition, serum was obtained from tubes without anticoagulant. Biochemical measurements (Table 3) included red blood cells (RBC), white blood cells (WBC) and platelet count, fasting glucose, and lipid measurements (total cholesterol (TC), HDL-cholesterol (HDL-C), LDL-cholesterol (LDL-C), triglycerides (TG)), hepatic (Aspartate transaminase, AST; Alanine transaminase, ALT; total plasma proteins) and renal function (creatinine, urea), using reagents and kits from Roche Diagnostics (Manheim, Germany). Coagulation factors (international normalized ratio, INR; activated partial thromboplastin time, APTT, Instrumentation Laboratory SpA, Milano, Italy) were assessed during the same day. Serum for biochemical measurement of insulin, inflammatory markers (high sensitivity C-reactive protein CRP, IL6, IL1 $\beta$ , TNF- $\alpha$ ), was frozen and stored at -20°C and eventually analyzed using ELISA kits (insulin EIA-1825, CRP HS EIA-3954, TNF- $\alpha$  EIA-4774, IL6 EIA-1869, IL10 EIA 0773 from DRG Instruments GmbH, Germany, IL1 $\beta$  EL10028 from Bio Supply, UK). Apolipoprotein A1 (ApoA1) and apolipoprotein B (ApoB) were assessed based on immunoturbidimetric tests (DiaSys Diagnostic Systems GmbH, Holzheim, Germany).

PARAMETER	C group	F group	P value
Age	54.80 $\pm$ 1.98	57.10 $\pm$ 1.26	NS
BMI	30.92 $\pm$ 1.95	29.64 $\pm$ 0.84	NS
Waist	117.00 $\pm$ 4.49	111.45 $\pm$ 1.48	NS
SBP(mm Hg)	145.50 $\pm$ 5.13	149.00 $\pm$ 6.22	NS
DBP(mm Hg)	90.00 $\pm$ 3.07	90.50 $\pm$ 2.83	NS
Glucose (mg/dl)	126.00 $\pm$ 11.00	127.00 $\pm$ 19.00	NS
HDL-cholesterol (mg/dl)	56.78 $\pm$ 2.49	44.78 $\pm$ 3.37	<0.05
Triglycerides	166.50 $\pm$ 25.56	220.40 $\pm$ 20.57	NS

Clinical and biochemical measurements were performed at the beginning of the study, after group randomization (group means  $\pm$  standard error). C group, the group assigned to corn oil supplementation; F group, the group assigned to flaxseed oil supplementation; BMI, body mass index; SBP, systolic blood pressure; DBP, diastolic blood pressure. Statistical significance was determined using Student's t-test for p<0.05. NS, no significance.

**Table 1:** Baseline Measurements.

FA species		FA concentration $\mu$ mol/mL	
		Corn oil	Flaxseed oil
14:0	myristic	ND	1.26
16:0	palmitic	368.51	200.44
16:1n7	palmitoleic	3.78	3.26
18:0	stearic	55.47	119.01
18:1n9	oleic	844.36	528.58
18:2n6	linoleic	1653.74	554.79
18:3n3	linolenic	13.51	1638.40
20:0	eicosanoic	25.24	ND
20:1n9	11-eicosenoic	9.31	5.79
20:2n6	11,14-eicosadienoic	ND	1.27
20:3n3	11,14,17-eicosatrienoic	ND	1.53
22:0	behenic	3.62	3.93
22:1n9	erucic	ND	1.28
22:5n3	7,10,13,16,19-docosapentaenoic	1.66	1.06
24:0	lignoceric	5.42	3.69

ND, not detectable

**Table 2:** Fatty acid composition of supplements.

<p><b>Clinical</b></p> <p>BMI</p> <p><b>Blood cells</b></p> <p>Haemoglobin (Hb)</p> <p>Hematocrit (Ht)</p> <p>Leucocytes count (LE)</p> <p>Neutrophyles count (NE)</p> <p>Lymphocytes count (LY)</p> <p>Monocytes count (MO)</p> <p>Eosinophyles count (EO)</p> <p><b>Glucose metabolism</b></p> <p>Glucose (Glu)</p> <p>Glycated haemoglobin (HbA1)</p> <p>Insulin</p> <p>HOMA (derived)</p> <p><b>Lipid metabolism</b></p> <p>Total cholesterol (Col)</p> <p>Triglycerides (TG)</p> <p>HDL cholesterol-(HDL)</p> <p>LDL-cholesterol (LDL)</p> <p>Apolipoprotein A1 (ApoA1)</p> <p>Apolipoprotein B (ApoB)</p> <p>Lipoprotein A (LpA)</p> <p>HDL/Col (derived)</p>	<p><b>Blood coagulation</b></p> <p>Platelets (PLT)</p> <p>International normalized ratio (INR)</p> <p>Fibrinogen (r-Fibe)</p> <p>Activated partial thromboplastin time (APTT)</p> <p>Von Willebrand factor</p> <p><b>Inflammation markers</b></p> <p>C-reactive protein (CRP)</p> <p>Oxidized LDL (oxLDL)</p> <p>Interleukin 6 receptor (IL6R)</p> <p>Interleukin 10 (IL10)</p> <p>Interleukin 17 (IL17)</p> <p>Interleukin 1 beta (IL1beta)</p> <p>Tumor necrosis factor alpha (TNF<math>\alpha</math>)</p> <p>CD40</p> <p>ICAM1</p> <p><b>Kidney function</b></p> <p>Urea</p> <p>Creatinine</p> <p>Sodium (Na)</p> <p>Potassium (K)</p> <p>Chloride (Cl)</p> <p>Uric acid</p>	<p><b>Liver function</b></p> <p>Aspartate transaminase (AST)</p> <p>Alanine transaminase (ALT)</p> <p>Total proteins</p> <p><b>Fatty acids</b></p> <p>14:0 (myristic)</p> <p>14:1 (myristoleic)</p> <p>16:0 (palmitic)</p> <p>16:1n7 (palmitoleic)</p> <p>18:0 (stearic)</p> <p>18:1n9 (oleic)</p> <p>18:2n6 (linoleic)</p> <p>18:3n3 (linolenic)</p> <p>20:0 (eicosanoic)</p> <p>20:1n9 (11-eicosenoic)</p> <p>20:2n6 (11,14-eicosadienoic)</p> <p>20:3n6 (dihomo-gamma-linolenic)</p> <p>20:4n6 (arachidonic)</p> <p>20:3n3 (11,14,17-eicosatrienoic)</p> <p>22:0 (behenic)</p> <p>22:1n9 (erucic)</p> <p>20:5n3 (eicosapentaenoic)</p> <p>24:0 (lignoceric)</p> <p>24:1 (nervonic)</p> <p>22:5n3 (7,10,13,16,19-docosapentaenoic)</p> <p>22:6n3 (docosahexaenoic)</p>
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**Table 3:** Parameters measured at the beginning and end of study.

Platelet count and coagulation factors were assessed at inclusion and at the end of the study for safety reasons, due to the possible antithrombotic activity, reduced platelets aggregability and substantial increase of bleeding time after 3 months of flaxseed administration, as previously reported [12,13]. However, no such adverse events were reported in our study.

Insulin resistance was calculated using the HOMA-IR index, which derives an estimate of whole-body insulin sensitivity from fasting glucose and insulin concentrations:  $\text{glucose (mg/dL)} \times \text{insulin } (\mu\text{U}/\text{mL})/405$  [14].

Total lipids from capsules and plasma (200 $\mu$ l) were extracted using the method of Bligh and Dyer at the UNC Nutritional Biochemistry and Molecular Biology Core. The lower (chloroform) phase was transferred to a clean tube and evaporated to dryness under nitrogen. The residual lipids were saponified and the fatty acids trans-methylated by sequential 1 ml addition of 4.25 % NaOH in  $\text{CHCl}_3$ ; MeOH (2:1, v/v) and 1N HCl in saline [16]. The samples were mixed vigorously then centrifuged at 1500 rcf for 5 minutes. The lower phase containing the fatty acid methyl esters was carefully transferred to a clean, dry tube and evaporated to dryness under nitrogen. Fatty acid methyl esters were then resuspended in 50  $\mu$ l undecane, and analyzed using capillary gas chromatography (GC) on a Perkin Elmer AutoSystem XL Gas Chromatograph (Shelton, CT) split injection, with helium as the carrier gas. The methyl esters were separated on a capillary column coated with 70% cyanopropyl polysilphenylene – siloxane (10 m x 0.1 mm ID- BPX70 0.2  $\mu$ m; SGE, Austin, TX) injector 240  $^{\circ}$ C and detector 280  $^{\circ}$ C. Data was analyzed with the Perkin Elmer Totalchrom Chromatography Software, version 6.2. Heptadecanoic acid (17:0) was added to the samples as an internal standard to correct for recovery and quantitation. Individual fatty acids were identified by comparing their retention with authentic standards (Nu Chek Prep, Elysian, MN).

### Data analysis

Data was analyzed using the JMP 8 analysis software (SAS Institute, Cary, NC). Time-treatment interaction was assessed by two factor Anova. Student's t-test was used to assess significance between the two groups, for each variable and time point (D0 or D90). Paired t-tests

were used for assessing significance of change in each group between start and end of study (D0 vs. D90). Because variances were, in most cases, not equal between groups, additional testing was performed. Bivariate analysis was performed for each parameter and group, to determine the dependency relationship between D0 and D90 values. Logistic fit testing was performed on D90/D0 ratios for each parameter and between groups, in order to assess the predictive value of changes for subject classification in either of the treatment groups. For all tests,  $p < 0.05$  was considered to declare significance of change.

### Results

In this study 64 parameters were analyzed for potential changes between the group receiving corn oil (C) and the group receiving flaxseed oil (F), for two time points (D0 and D90, Table 3). Preliminary analysis of data revealed that, for many parameters, variances were not equal between groups and, therefore, in such cases, the assumptions for t-testing were not met (data not shown). The unequal variances were present because of the small sample size ( $n=10$  per group) and, for some biochemical parameters, due to the high variability between subjects. In order to determine whether changes between D90 and D0 have predictive value for group assignment, logistic fit was performed using the D90/D0 ratios, calculated as D90/D0 values for each subject. Bivariate analysis was used to determine the correlation degree between D0 and D90 individual values for each treatment group. Table 4 indicates the parameters for which significance (maintenance of correlation between D0 and D90 values) was present in either of the groups, but lost in the other.

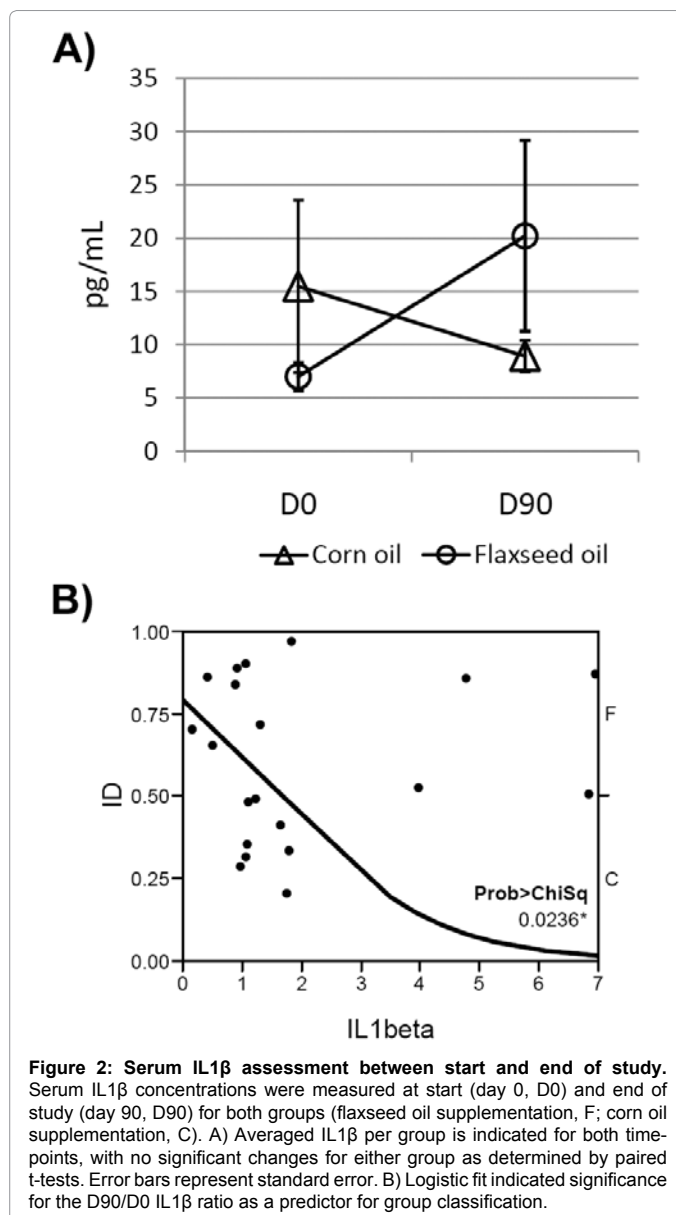
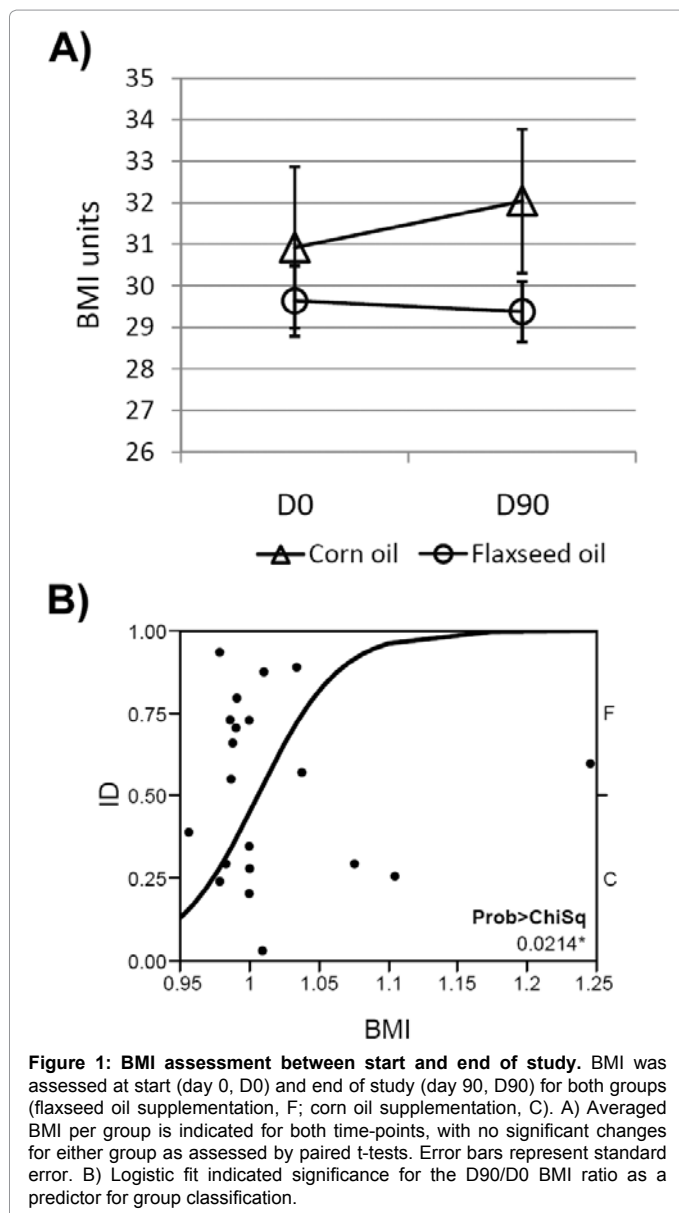
### MS-related parameters

Analysis using paired-t tests revealed no changes between groups and time points (data not shown). Analysis of two other related parameters, BMI and insulin concentrations, revealed significant changes when either logistic fit or bivariate analysis was applied. While BMI averages between groups and time points did not indicate significant changes (Figure 1A) the relative BMI ratios (D90/D0) were smaller in the F group ( $0.99 \pm 0.01$  SE) than in the C group ( $1.04 \pm 0.03$  SE), with significant predictive value (Figure 1B) for group assignment. Serum insulin concentration and the derived HOMA index both

increased in the C group (D90/D0 ratios  $2.11 \pm 0.79$  SE and  $2.00 \pm 0.66$  SE, respectively), while for the F group both parameters maintained their individual correlation between time points (average D90/D0 ratio  $1.16 \pm 0.20$ ,  $p < 0.05$ , Table 4). The bivariate model for total cholesterol (Col) and triglycerides (TG) associated with MS indicated that, although no statistical significance was achieved for averages between groups, correlation between individual D90 and D0 values were maintained only for the C group, while the F group registered loss of correlation (Table 4). Lower ApoA1 D90/D0 ratios were also predictive for the F group, as indicated in Figure 3.

### Markers of inflammation

Among the markers of inflammation indicated in Table 3, flaxseed oil supplementation induced an increase in IL1 $\beta$ , while opposite changes were induced by corn oil supplementation (Figure 2A). The D90/D0 ratios ( $2.90 \pm 0.80$  SE in F group versus  $1.13 \pm 0.17$  SE in C group) also had predictive value for group assignment (Figure 2B). Within the F group, TNF $\alpha$  individual values were correlated between



D0 and D90 ( $p < 0.018$ ) while for the C group the correlation was not present between the same time-points.

### Plasma fatty acids composition

Among the fatty acid species in plasma,  $\alpha$ -linolenic acid (ALA, 18:3n3) had lower D90/D0 ratios assigned to the F group as compared to the C group values ( $0.63 \pm 0.11$  SE vs  $1.25 \pm 0.32$  SE, respectively, Figure 4). Time-treatment interaction analysis indicated significance for 11-eicosenoic plasma levels (Figure 5A), as determined by the ANOVA-two factor test ( $p < 0.05$ ). These changes were also significant for group assignment, as indicated by logistical fit analysis of the D90/D0 ratios, with higher ratios assigned to the C group ( $1.63 \pm 0.20$  SE) when compared to the F group ( $1.00 \pm 0.11$  SE) (Figure 5B). Six other fatty acid species were differently correlated between the two groups, indicating that the low dose of either flaxseed or corn oil could induce specific changes in their plasma profiles (Table 4).

### Discussion

In our study, several parameters were modified by flaxseed oil

Variable	Corn oil		Flaxseed oil	
	D90/D0 ratio	D90/D0 bivariate model (Prob > F)	D90/D0 ratio	D90/D0 bivariate model (Prob > F)
AST	0.90 ± 0.10	NS	<b>0.84 ± 0.11</b>	<b>0.0096</b>
ALT	1.26 ± 0.35	NS	<b>0.97 ± 0.14</b>	<b>0.0017</b>
Col	<b>1.03 ± 0.04</b>	<b>0.0146</b>	0.98 ± 0.04	NS
TG	<b>1.05 ± 0.10</b>	<b>0.0057</b>	0.93 ± 0.09	NS
Insulin	2.11 ± 0.79	NS	<b>1.12 ± 0.17</b>	<b>0.0359</b>
HOMA	2.00 ± 0.66	NS	<b>1.16 ± 0.20</b>	<b>0.0243</b>
TNFα	1.20 ± 0.70	NS	<b>0.75 ± 0.19</b>	<b>0.0188</b>
<b>Fatty acid species</b>				
14:0 (myristic)	0.94 ± 0.11	NS	<b>0.99 ± 0.09</b>	<b>0.0102</b>
18:0 (stearic)	1.02 ± 0.05	NS	<b>0.93 ± 0.04</b>	<b>0.0489</b>
18:2n6 (linoleic)	1.05 ± 0.07	NS	<b>0.99 ± 0.04</b>	<b>0.0032</b>
20:0 (arachidic)	<b>1.21 ± 0.21</b>	<b>0.0185</b>	1.88 ± 0.43	NS
20:2n6 (eicosadienoic)	1.23 ± 0.09	NS	<b>1.12 ± 0.06</b>	<b>0.0034</b>
20:3n6 (dihomo-gamma-linolenic)	1.14 ± 0.14	NS	<b>1.01 ± 0.08</b>	<b>0.0258</b>

Linear regression was performed within each group for correlation between time points (D0 and D90). Bivariate analysis was considered significant for  $p < 0.05$

**Table 4:** Parameters with significant correlation between time points.

supplementation, as compared with the group receiving corn oil. BMI did not change over the 90 days treatment period in the F group, while the C group registered an increase in BMI. Logistic fit of the BMI D90/D0 ratios indicated that such changes have predictive value for group assignment. Similar predictive values were obtained for changes in IL1beta, ApoA1, ALA, and eicosenoic acid (Figures 1-5).

Bivariate analysis indicated that the flaxseed oil supplementation reduced insulin resistance as assessed by insulin levels and the derived HOMA index. In the F group, the values from D0 did not predict the D90 values, in contrast with the C group, where the correlation was maintained. TNFα also presented a similar pattern (Table 4).

Surprisingly, lower ALA D90/D0 ratios were assigned to the F group, which could indicate an increase of the ALA desaturation/elongation rates. However, changes in the levels of other omega-3 fatty acids were not identified. One could speculate that the amount of ALA received (1.2 g/day) would be insufficient to provide enough substrate for such detectable increases. In addition, the desaturation/elongation of ALA has been reported to be lower in men than in women (reviewed in [17]).

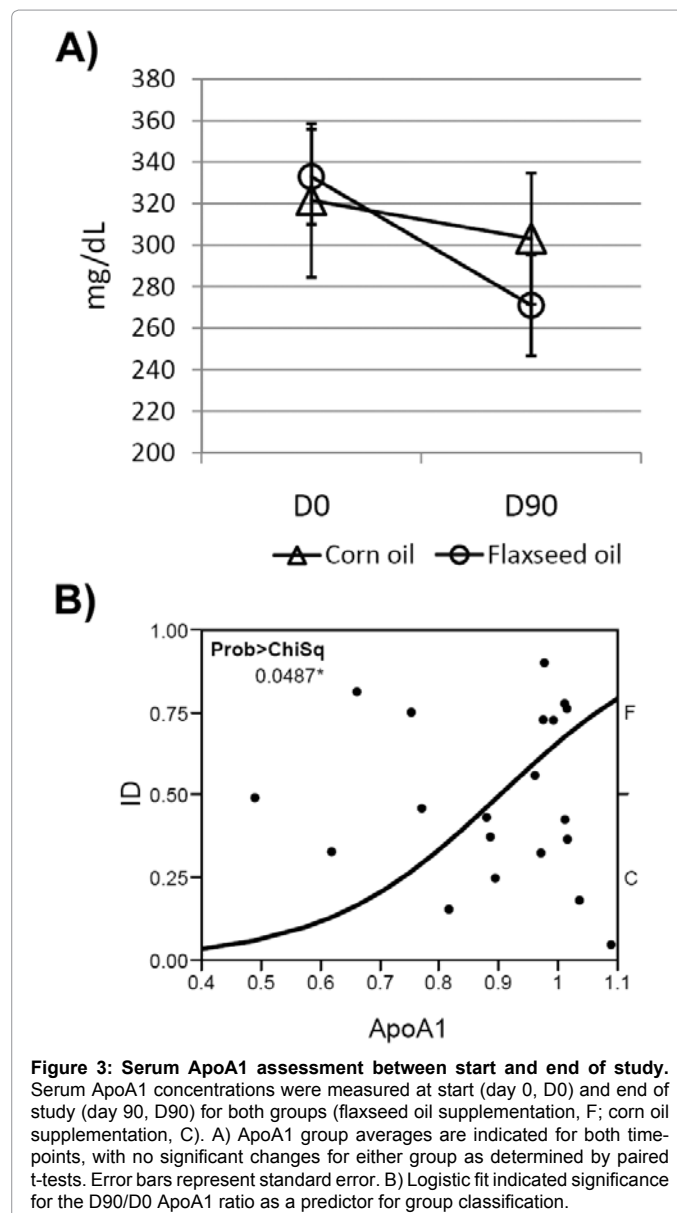
As indicated in Table 2, the major difference in fatty acid composition between corn oil and flaxseed oil supplements, consists of linoleic (LA) and alpha-linolenic (ALA) acids, respectively. Both species had similar molar concentrations (more than 1.6 mmol/mL). Therefore, it is not clear whether the observed changes could be assigned to only one of the treatments. However, we may speculate that the flaxseed oil supplementation was favorable against the BMI increase and the increased insulin resistance observed with corn oil supplementation.

While most studies using omega-3 fatty acids have focused on the effects of EPA and DHA, and upon the role that ALA has in increasing their synthesis, a limited number of studies suggested that ALA may also act as a cell-signaling molecule. In animal and cell culture models ALA has a distinct role in intra-cellular signalling [18-19] and its role in the regulation of gene expression is mediated either through peroxisome proliferator activated receptor (PPAR) -dependent,

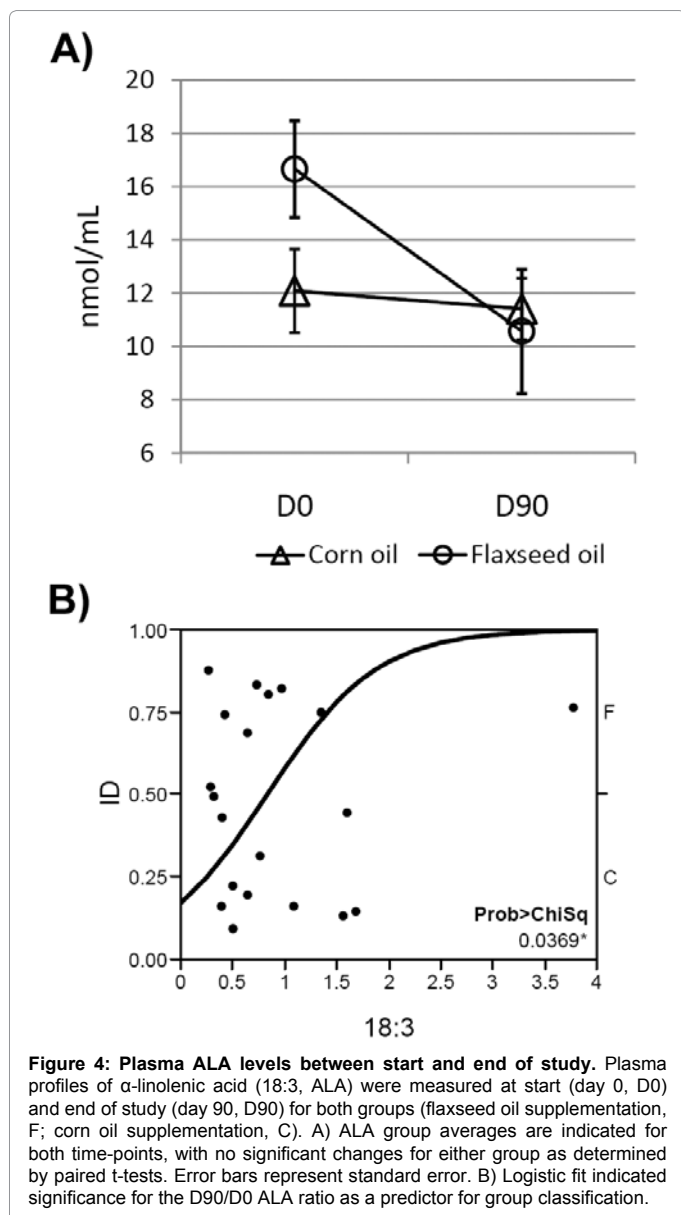
PPAR-independent mechanisms (reviewed in [20]), or by altering the phosphorylation of mitogen-activated protein kinases (MAPKs) involved in cell proliferation, differentiation and apoptosis [19,21]. ALA inhibits both TNF-α gene expression and NF-kB-dependent transcriptional activity [19]. In our study, TNF-α values were correlated between D0 and D90 in the F group, while corn oil supplementation was associated with a loss of correlation between the same time points (Table 4).

Because it is considered that ALA-derived DHA in humans accounts for less than 1% (reviewed in [22]), most of the research on ω-3 fatty acids has been focused on dietary interventions using DHA, which is a major component of plasma membranes [22]. Similarly, in animal models, the conversion rate of ALA to DHA in rat brain and liver is less than 1% [23,24]. Moreover, n-3 PUFA deficient diets up-regulate the ALA to DHA conversion [25]. Therefore, there is considerable debate as to whether ALA deficiency reflects into subsequent DHA reduction.

The present study was designed to assess the effect of a low-dose



**Figure 3: Serum ApoA1 assessment between start and end of study.** Serum ApoA1 concentrations were measured at start (day 0, D0) and end of study (day 90, D90) for both groups (flaxseed oil supplementation, F; corn oil supplementation, C). A) ApoA1 group averages are indicated for both time-points, with no significant changes for either group as determined by paired t-tests. Error bars represent standard error. B) Logistic fit indicated significance for the D90/D0 ApoA1 ratio as a predictor for group classification.



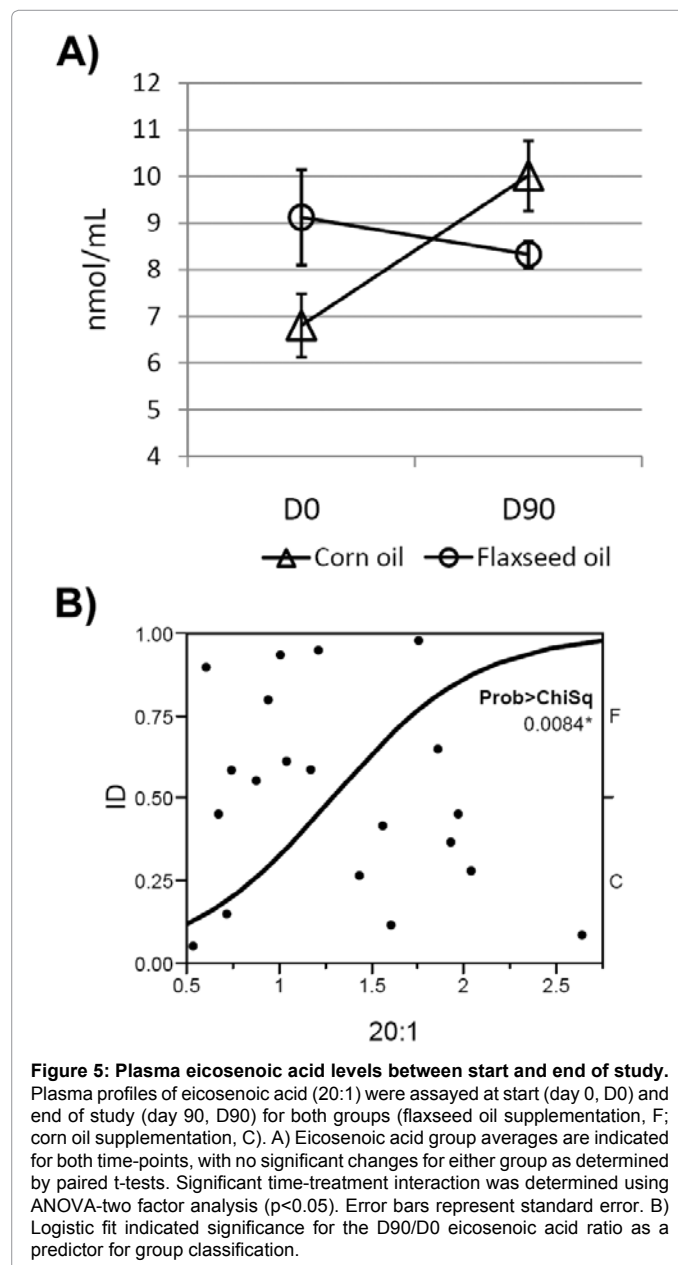
**Figure 4: Plasma ALA levels between start and end of study.** Plasma profiles of  $\alpha$ -linolenic acid (18:3, ALA) were measured at start (day 0, D0) and end of study (day 90, D90) for both groups (flaxseed oil supplementation, F; corn oil supplementation, C). A) ALA group averages are indicated for both time-points, with no significant changes for either group as determined by paired t-tests. Error bars represent standard error. B) Logistic fit indicated significance for the D90/D0 ALA ratio as a predictor for group classification.

flaxseed oil supplementation in men with metabolic syndrome and it has several particularities. As indicated in Table 2, the flaxseed oil is very rich in ALA (at least 50%), which is the precursor for EPA and DHA. Our subjects received 2.4 g flaxseed oil per day, which contained a dose of approximately 1.2 g ALA. Due to the relative low socio-economic status of this developing country, the Romanian population has, by large, been only weakly exposed to dietary supplements as compared to consumers from developed countries [26]. In addition, the two main types of cooking oils that contain ALA (canola and soybean oil) are not generally used by the majority of Romanians. Therefore, the selection of the study site was designed to include subjects with a lower probability of significant omega-3 fatty acids intakes.

Another particularity of this study is that the subjects did not have access to adequate treatment against MS, largely due to the lack of financial means to procure the prescribed medicine, although they had unrestricted access to medical supervision. With the exception of antihypertensive treatment, these subjects unfortunately were not able to adequately follow the prescribed medication, nor were in the

position to follow the dietary recommendations that would include healthier but more expensive foods. Therefore, this study was designed to provide information regarding a less expensive alternative for omega-3 fatty acids consumption.

Previous studies using flaxseed oil supplementation reported discordant results. According to a meta analysis published in 2006, dietary supplementation with ALA was shown to only decrease fasting glucose and fibrinogen concentrations, with no other influence on cardiovascular risk factors (lipid parameters, inflammation markers, systolic or diastolic blood pressure, or BMI) [27]. In relative contrast to our study, ALA administration was ineffective upon glycemic control and insulin resistance in diabetic patients receiving different amounts of ALA [28-30]. Paschos et al indicated that, in subjects with dyslipidemia but no metabolic syndrome, flaxseed oil (8.1 g/d) supplementation decreased plasma adiponectin, increased plasma TNF- $\alpha$ , and produced a small but significant decrease in serum HDL-



**Figure 5: Plasma eicosenoic acid levels between start and end of study.** Plasma profiles of eicosenoic acid (20:1) were assayed at start (day 0, D0) and end of study (day 90, D90) for both groups (flaxseed oil supplementation, F; corn oil supplementation, C). A) Eicosenoic acid group averages are indicated for both time-points, with no significant changes for either group as determined by paired t-tests. Significant time-treatment interaction was determined using ANOVA-two factor analysis ( $p < 0.05$ ). Error bars represent standard error. B) Logistic fit indicated significance for the D90/D0 eicosenoic acid ratio as a predictor for group classification.

cholesterol. Other studies in healthy individuals revealed different effects of omega-3 fatty acids administration on lipid parameters, from none to decreased LDL-C and LpA, increased HDL in men, and improved insulin resistance [31-34]. A recent study using flaxseed supplementation reported beneficial effects upon subjects with MS (decreased HOMA), but it was not clear whether the reported results were due to the presence of lignans or ALA, or both [35].

In our study, with the exception of a correlated increase of D90/D0 IL1 $\beta$  ratios within the F group, we found no changes of inflammation markers (averaged values). However, regression analysis revealed that TNF- $\alpha$  was correlated between start and end of study only in the F group. Similarly, insulin sensitivity was correlated in the F group. Our results are in agreement with published studies that have indicated that TNF- $\alpha$  plays an important role in mediating insulin resistance [36]. Increased intakes of dietary ALA demonstrated anti-inflammatory effects by inhibiting IL-6, IL-1, and TNF- $\alpha$  production in cultured peripheral blood mononuclear cells (PBMCs) [37].

An unexpected finding was the significant logistic fit of higher IL1 $\beta$  D90/D0 ratios in the F group. In many studies, MS was associated with increased pro-inflammatory markers, including IL1 $\beta$  (reviewed in [38]), and our results are not in agreement with these findings. However, the roles of IL1 $\beta$  in mediating MS effects, including insulin resistance, are not yet well understood. IL1 $\beta$  genetic polymorphisms were associated with increased risk of MS in subjects with low PUFA levels, suggesting that IL1 $\beta$  loss of function could also increase the risk for MS [39].

The logistic fit analysis indicated that decreased serum ApoA1 D90/D0 ratios are associated with flaxseed oil supplementation (Figure 2). While the ApoB/ApoA1 ratio is the best indicator for MS risk, ApoA1 levels alone are not considered as an adequate indicator for MS assessment [40,41].

The present study has important limitations. The most important factor was the high and unequal variance in data distribution, due to a small sample size in both groups, and to high physiological variations between individuals. As indicated in Table 1, group randomization failed in regard to HDL-cholesterol distribution. Therefore, parametric assumptions for t-testing were frequently not met among the measured variables. While logistic fit or bivariate analyses rendered significance for the discussed variables, it is not clear whether the reported changes have biological significance. We surmise that flaxseed oil supplementation, in opposition to corn oil, was associated primarily with no increase in BMI and insulin resistance, while other reported changes are difficult to interpret in the context of our study design.

Our study aimed to determine whether a low-dose flaxseed oil supplementation regimen could be effective in improving physiological and biochemical parameters associated with metabolic syndrome. Flaxseed oil supplementation associated with no increases in BMI and insulin resistance over a period of 90 days, as opposed to corn oil supplementation. Other alterations were reported for markers of inflammation (TNF- $\alpha$  and IL1 $\beta$ ), apolipoprotein A1, and plasma fatty acid composition. We hypothesize that these outcomes could be the result of  $\alpha$ -linolenic acid supplementation, in accordance with previous similar studies indicating its role in improving insulin sensitivity. However, two notable aspects differentiate our study from previous reports: 1) the enrolled subjects were patients receiving no adequate treatment against metabolic syndrome, and 2) the reported changes were a result of a low dose of flaxseed oil as compared to previous studies.

While the present study suggested that low-dose flaxseed oil supplementation could be beneficial for middle-aged men with untreated metabolic syndrome, more and better controlled studies are necessary to clarify these initial findings.

### Competing Interests

The authors declare that they have no competing interests.

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