

Linking Histone Methylation to Active DNA Demethylation

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The role of epigenetic modifications in regulating gene expression and disease progression is continuously evolving with the vast increase in our knowledge about these modifications and their physiological function. The histone code hypothesis initially proposed by Strahl and Allis [1] indicated that different combinations of covalent histone modifications influence chromatin structure and the output of the transcription machinery. The hypothesis was further extended to link the distinct histone combinations to physiological processes like apoptosis and the description of the apoptotic histone code [2]. Histone acetylation by histone acetyltransferase (HAT) and deacetylation by histone deacetylases (HDAC) are considered the most studied histone modifications and had been the focus of intense investigation in the last two decades. Those efforts were acclaimed by the FDA approval of the HDAC inhibitors vorinostat and romidepsin for the treatment of Cutaneous T Cell Lymphoma (CTCL). The link between histone acetylation and other epigenetic modifications like DNA methylation was unclear until Bird and others [3,4] described the recruitment of HDAC repressor complex by methyl-CpG-binding proteins to methylated cytosines on DNA. The concept of hierarchical organization of histone acetylation and DNA methylation was proposed later on and speculated that DNA methylation is dominant over histone acetylation [5,6]. Emerging clinical trials using the combination of HDAC inhibitors and DNA methyltransferase (DNMT) inhibitors embraced that concept and adopted a sequential approach of administration starting with DNMT inhibitor followed by HDAC inhibitor [7,8].

Histone lysine methylation is another histone modification that can regulate the DNA methylation machinery. Histone lysine methylation is a dynamic process regulated by histone lysine methyltransferases (KMTs) and demethylases (KDMs) and plays an important role in tumor progression. Epigenetic readers like the inhibitor of growth protein 1 (ING1) recognize histone trimethylation on a specific lysine residue (H3K4) [9]. The ING family (ING1-5) has been an emerging putative tumor suppressor gene that binds to histones in a methylation-sensitive manner through a conserved Plant Homeodomain (PHD). Recently, the role of ING1 as an adaptor for the DNA demethylation machinery was identified [10]. ING1 binding to trimethylated H3K4 recruits the growth arrest and DNA Damage protein 45a (Gadd45a). Gadd45a induces gene-specific DNA demethylation through several steps that include deamination of methylated cytosine by Activation-Induced Deaminase (AID) enzyme followed by base excision repair using the thymine DNA glycosylase MBD4 [11].

Based on such findings, it is tempting to hypothesize that the combination of lysine-specific demethylase inhibitor with DNMT inhibitor would potentiate the DNA demethylation activity of the latter and enhance the re-expression of epigenetically silenced genes. Lysine-Specific Demethylase1 (LSD1) suppresses gene expression by demethylating mono- and dimethyl-H3K4 histone marks that are associated with active gene expression. KDM5C/Jarid1C is another transcriptional repressor lysine-specific demethylase that acts on trimethylated and dimethylated H3K4. Inhibitors of LSD1 and KDM5C could potentiate the induction of silenced genes by DNMT inhibitors. Indeed, LSD1 inhibitors as single agents were successful in re-expressing epigenetically silenced genes like *CDH1* in leukemia

cells and augmented apoptosis induction when combined with HDAC inhibitors in glioblastoma cells [12,13]. Although such combination suffers the common disadvantage of epigenetic agents; induction of global changes in histone and DNA methylation with consequent nonspecific gene expression changes, it may constitute a successful combination therapy for different types of cancer.

References

- Strahl BD, Allis CD (2000) The language of covalent histone modifications. *Nature* 403: 41-45.
- Füllgrabe J, Hajji N, Joseph B (2010) Cracking the death code: apoptosis-related histone modifications. *Cell Death Differ* 17: 1238-1243.
- Bird AP, Wolffe AP (1999) Methylation-induced repression--belts, braces, and chromatin. *Cell* 99: 451-454.
- Knoepfler PS, Eisenman RN (1999) Sin meets NuRD and other tails of repression. *Cell* 99: 447-450.
- Cameron EE, Bachman KE, Myöhänen S, Herman JG, Baylin SB (1999) Synergy of demethylation and histone deacetylase inhibition in the re-expression of genes silenced in cancer. *Nat Genet* 21: 103-107.
- Jones PA, Baylin SB (2002) The fundamental role of epigenetic events in cancer. *Nat Rev Genet* 3: 415-428.
- Gore SD, Baylin S, Sugar E, Carraway H, Miller CB, et al. (2006) Combined DNA methyltransferase and histone deacetylase inhibition in the treatment of myeloid neoplasms. *Cancer Res* 66: 6361-6369.
- Fandy TE, Herman JG, Kerns P, Jiemjit A, Sugar EA, et al. (2009) Early epigenetic changes and DNA damage do not predict clinical response in an overlapping schedule of 5-azacytidine and entinostat in patients with myeloid malignancies. *Blood* 114: 2764-2773.
- Peña PV, Hom RA, Hung T, Lin H, Kuo AJ, et al. (2008) Histone H3K4me3 binding is required for the DNA repair and apoptotic activities of ING1 tumor suppressor. *J Mol Biol* 380: 303-312.
- Schäfer A, Karaulanov E, Stapf U, Döderlein G, Niehrs C (2013) Ing1 functions in DNA demethylation by directing Gadd45a to H3K4me3. *Genes Dev* 27: 261-273.
- Rai K, Huggins IJ, James SR, Karpf AR, Jones DA, et al. (2008) DNA demethylation in zebrafish involves the coupling of a deaminase, a glycosylase, and gadd45. *Cell* 135: 1201-1212.
- Murray-Stewart T, Woster PM, Casero RA Jr (2013) The re-expression of the epigenetically silenced e-cadherin gene by a polyamine analogue lysine-specific demethylase-1 (LSD1) inhibitor in human acute myeloid leukemia cell lines. *Amino Acids*.
- Singh MM, Manton CA, Bhat KP, Tsai WW, Aldape K, et al. (2011) Inhibition of LSD1 sensitizes glioblastoma cells to histone deacetylase inhibitors. *NeuroOncol* 13: 894-903.

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