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In-vitro Study of Anti-Fusarium Effect of Thymol, Carvacrol, Eugenol and Menthol

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Abstract

This study aims at shedding light on the anti-fusarium action of some Major Compounds (MCs) of Essential oils (EOs). To this end, *Fusarium oxysporum f. sp. dianthi* (Fod) was used as fungal model. A screening of four MCs was conducted in order to identify the most active one with the highest anti-*Fusarium* effect, using both agar and broth dilution methods. The obtained data revealed that thymol, with its minimal inhibitory concentration (MIC) and minimal fungicidal concentration (MFC) values was the most effective, with concentrations between 0.25 mg ml⁻¹ and 1 mg ml⁻¹. Among the four compounds tested, thymol and carvacrol showed anti-*Fusarium* activity by inhibiting the germination and destroying Fod conidia. As a preliminary test, we have also tested the applicability of thymol on soil disinfection, with very promising results. The antifungal activity of the four MCs used in this study involves inhibition of germination and destruction of Fod conidia. Thymol, the most effective MC, has shown great effect on soil disinfection. This work is a preliminary contribution aiming at developing an alternative product, environmental friendly, safe for both the workers and consumers, with high anti-Fusarium activity.

Keywords: Major compounds; Essential oils; *Fusarium oxysporum f. sp. dianthi*; Fungicidal action; Soil disinfection

Introduction

Fusarium wilt in carnations is a vascular disease caused by the fungus *Fusarium oxysporum f. sp. dianthi* (Fod). It is the most common and serious fungal disease that attacks carnations worldwide [1]. The fungus can cause big financial losses to the floriculture industry. To control the *Fusarium* wilt, growers use systemic fungicides like methyl bromide [1], chloropicrin and benomyl [2]. Beside the fact that these chemicals are toxic to humans and the environment [3], they can also cause selection for resistant mutated pathogen [4].

Severe epidemics of *Fusarium* wilt in the 1980s and 1990s have caused a strong reduction in the acreage devoted to the carnation industry in some areas of Southern Europe, such as Italy, France, and Spain [5]. *Fusarium* has obliged the industry players to leave Europe and the United States towards other countries such as Colombia and Morocco [5]. Despite the availability of cultivars that are relatively resistant to some strains of Fod, *Fusarium* wilt remains a threatening disease for this crop wherever it is grown [5].

Several publications from our laboratory have previously reported *in vitro* and *in vivo* antifungal activity of some EOs and their MCs, especially the phenolic compounds such as thymol, carvacrol and eugenol [6-8].

The major aim of the present work was to elucidate the *in vitro* antifungal activity of some MCs of EOs on the growth of Fod in order to develop an alternative to chemical fungicides.

Materials and Methods

Fungal strain

The Fod strain used in this study was isolated in our laboratory from a diseased carnation plant collected from a greenhouse of Prim'Rose Company (Azemmour, Morocco).

The Fod strain identification was done using a conventional method based on the colony's macroscopic characteristics and the microscopic appearance of conidia [9].

Culture media

Potato dextrose agar (PDA) (Biokar, France) was used for fungus identification [9]. Sabouraud dextrose agar (SDA), Sabouraud dextrose broth (SDB) (Biokar, France) and Malt extract broth (MEB) were used for fungus culture and antifungal test [10].

Preparation of conidial suspension

Conidia of fod were harvested from 7-day-old cultures in PDA by pouring a sterile 0.9% aqueous solution of NaCl onto the culture plates and scraping the plate surface with a bent glass rod to facilitate the release of conidia. The number of conidia cells was adjusted to 10^6 ml⁻¹ and was determined by transferring 10 µl of the sample suspension of Fod conidia to a Malassez chamber for microscopic examination and counting. Fod conidia were counted in 10 different fields utilizing standard techniques [11].

Anti-fungal agents

Four MCs were used in this study: Thymol; Carvacrol; Eugenol and Menthol, all were obtained from Sigma-Aldrich (Steinheim, Germany). Each MC was dispersed in a suspension containing 0.2% agar in sterile distillated water in order to maintain viscosity to disperse MCs without adding solvent or detergent [12]. A stock suspension of 100 mg ml⁻¹ for each MC was then prepared.

Determination of anti-Fusarium effect of MCs in agar medium

The MCs were added to the medium at the concentrations of (0;

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0.125; 0.25; 0.5; 1; 2 and 4 mg ml⁻¹) from a stock suspension and poured in sterile Petri dishes (90 × 16 mm). The determination of anti-*Fusarium* effect of each MC in SDA medium was carried out by the inoculation of 10 μ l calibrated drop (10⁴ conidia) in agar plates containing increasing concentrations of MCs. Three Petri dishes were considered for each concentration, each petri dish contained three calibrated drops of Fod and the petri dish were incubated for 5 days at 27°C.

The diameter of mycelial colony of Fod was measured in centimeters for each concentration of all the MCs used and was compared to the control agar plate.

The minimal inhibitory concentration (MIC) was determined as the lowest antifungal agent concentration at which no mycelial growth was visually observed.

The anti-fusarium effect of MCs was expressed by the percentage of growth inhibition of Fod and calculated using the following formula [13]:

$$I(\%) = (D_c - D_i)/D_c \times 100$$

Where D_c and D_i represent respectively the mycelial growth diameters in control and in treated petri dishes.

Determination of anti-Fusarium effect of MCs in liquid medium

The determination of anti-*fusarium* effect of MCs in liquid medium (MEB) was conducted in triplicate using sterile 96 well microplates by adding 20 μ l of conidial suspension of Fod (2 × 10⁴ conidia ml⁻¹) and 70 μ l of increasing concentrations of MCs (0; 0.0625; 0.125; 0.25; 0.5; 1; 2 and 4 mg ml⁻¹) to 160 μ l of MEB medium in order to have a final volume of 250 μ l in each well. After 5 days of incubation at 27°C, the mycelial growth was assessed by measuring optical density (OD) at 600 nm using spectrophotometer (J.P. Selecta).

The anti-*Fusarium* effect of MCs was expressed by the percentage of inhibition of Fod and calculated using the following formula:

 $I(\%) = (OD_{c} - OD_{i})/OD_{c} \times 100$

Where OD_c and OD_i represent respectively the optical density of mycelial growth in control and in treated wells.

The MIC was defined as the lowest MC concentration at which no mycelia growth was visually observed after 5 days of incubation at 27°C.

In order to determine the minimal fungicidal concentration (MFC), 20 μ l from the wells showing no growth were aseptically transferred into sterile tubes containing 980 μ l of sterile MEB medium. The MFC was defined as the lowest antifungal agent concentration at which no mycelial growth was visually observed after 5 days of incubation at 27°C.

The Fod conidial germination assay

To study the effect of MCs on conidial germination, 100 μ l of freshly prepared conidia suspension (10⁶ ml⁻¹) was added to 900 μ l of MCs solutions at different concentrations (0; 0.0625; 0.125; 0.25; 0.5 and 1 mg ml⁻¹), in sterile SDB. The control tube contained 100 μ l of 10⁶ ml⁻¹ freshly prepared conidia suspension in 900 μ l of sterile SDB.

After 20 h at 27°C, 40 μl of Conidia suspensions were deposited between a slide and a coverslip and the conidia was observed at magnification 100x using an optical microscope.

The proportion of germinated conidia was counted as the ratio of germinated conidia among 100 observed conidia.

The Fod conidial destruction assay

Each MC was tested in increasing concentrations i.e., 0; 0.0625; 0.125; 0.25; 0.5 and 1 mg ml $^{\rm -1}$

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100 μl of freshly prepared conidia suspension (10⁶ ml⁻¹) was put in direct contact with 900 μl of MC at various concentrations in sterile 0.9% aqueous solution of NaCl containing 0.2% agar.

The Fusarium conidia destruction assay was evaluated after an incubation at $27^{\rm o}{\rm C}$ for 24 h.

As control tube, we used a 900 μl aqueous solution of sterile 0.9% of NaCl containing 0.2% agar and 100 μl of $10^6\,m l^{-1}$ freshly prepared conidia suspension.

The number of conidia was determined in a Malassez chamber (as previously described) after 1 h, 3 h, 6 h and 24 h of incubation at 27°C and the percentage of conidial destruction was calculated using the following equation [14]:

 $I(\%) = (N_c - Ni)/N_c \times 100$

Where $\rm N_{_c}$ and $\rm N_{_i}$ represent respectively the number of conidia in the control and the treated tubes.

Fusarium density in the soil

18 aliquots of 100 g sterilized soil were placed in sterile 400 ml plastic boxes. Fod was incorporated into sterilized soil (previously sifted through a 2-mm sieve, and then was autoclaved 30 min at 121°C) at an inoculum density of 10⁶ spores per gram of soil.

Experimental treatments were: (1) uninfested and untreated soil, only water added as a check for residual populations of Fod (not included in analysis); (2) infested and untreated soil, only water added; (3) infested and treated soil with thymol. Four concentrations of thymol were used for treatments (0.25; 0.5; 1 and 2 mg g⁻¹ of soil) and each concentration was calculated per gram of soil. Population densities of Fod were determined using the serial dilution technique at day 0 (before soil contamination), 1, 3, 7, 14, and 21 days after soil contamination.

One gram of soil from each batch was placed in a sterile tube containing 9 ml of 0.9% NaCl. From this stock solution, series of dilutions were carried out (10^{-2} , 10^{-3} ... 10^{-6}). 100 µl from each dilution were deposited on the surface of three Petri dishes containing SDA medium. Plates were then incubated at 27°C for 5 days. The number of Fod colonies was defined from countable plates.

Data analysis

The mycelial growth data was subjected to ANOVA statistical analysis. Mean and standard error of data were calculated using IBM SPSS Statistics software, version 20. The separation of means was done using the Least Significant Difference (LSD) test at p<0.05.

Results

Determination of anti-fusarium effect of MCs in agar medium

The effect of MCs on the growth of Fod in agar medium is shown in Tables 1 and 2. MCs exhibited varying levels of antifungal activity against Fod. The MIC values of MCs (Table 1) ranged between 0.25 mg ml⁻¹ and 2 mg ml⁻¹. The most effective inhibitors were thymol and carvacrol with a MIC value of 0.25 mg ml⁻¹ followed by eugenol with a MIC value of 0.5 mg ml⁻¹ while menthol had a MIC value of 2 mg ml⁻¹.

The inhibitory effect of MCs on the growth of Fod is depicted on Table 2. The mycelial growth of Fod was significantly reduced in

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Major compound	MIC (mg mL ⁻¹)
Thymol	0.25
Carvacrol	0.25
Eugenol	0.5
Menthol	2

Table 1: MIC values of the tested MCs on Fod in agar medium.

Majar Compoundo	Mycelial growth inhibition (%)							
Major Compounds	Control	0.125 mg ml ⁻¹	0.25 mg ml ⁻¹ 0.5 mg ml ⁻¹		1 mg ml ⁻¹	2 mg ml⁻¹	4 mg ml ⁻¹	
Thymol	0.00 ± 0.00^{a}	88.9 ± 0.11 ^b	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	
Carvacrol	0.00 ± 0.00^{a}	54.5 ± 0.11°	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	
Eugenol	0.00 ± 0.00^{a}	31.6 ± 0.07 ^d	51.8 ± 0.12°	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	
Menthol	0.00 ± 0.00^{a}	10.8 ± 0.12 ^e	24.3 ± 0.09 ^f	39.8 ± 0.119	86.7 ± 0.11 ^b	100.0 ± 0.00	100.0 ± 0.00	

Data are means (n=9) ± SD, different letters indicate significant differences (p<0.05) by LSD test.

Table 2: Effect of MCs on mycelial growth of Fod after 5 days of incubation at 27°C.

Major compound	MIC (mg ml ⁻¹)	MFC (mg ml ⁻¹)
Thymol	0.5	1
Carvacrol	0.5	1
Eugenol	1	2
Menthol	4	>10

 Table 3: MIC and MFC of the tested MCs on Fod in liquid medium.

Majar Compound	Mycelial growth inhibition (%)						
Major Compound	Control	0.125 mg/ml	0.25 mg/ml	0.5 mg/ml	1 mg/ml	2 mg/ml	4 mg/ml
Thymol	0.00 ± 0.00^{a}	78,7 ± 0.54 ^b	95.9 ± 3.17°	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00
Carvacrol	0.00 ± 0.00^{a}	61.5 ± 3.46 ^b	90.5 ± 0.51 ^h	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00	100.0 ± 0.00
Eugenol	0.00 ± 0.00^{a}	59.8 ± 3.86°	85.5 ± 3.05 ^d	89.7 ± 1.16°	92.2 ± 1.11°	100.0 ± 0.00	100.0 ± 0.00
Menthol	0.00 ± 0.00^{a}	39.3 ± 3.40 ^f	65.9 ± 1.99⁰	78.0 ± 4.30 ^h	86.7 ± 0.11 ^d	97.5 ± 0.26°	100.0 ± 0.00
Data ara Maana (n=	3) + SD different let	tore indicato significan	t differences (p<0.05)	by LSD toot			

Data are Means (n=3) ± SD, different letters indicate significant differences (p<0.05) by LSD test.

Table 4: Inhibitory effect of MCs on the growth of Fod in liquid medium.

Maine anna ann a	Conidia germination (%)							
Major compounds	Control	0.0625 mg ml ⁻¹	0.125 mg ml ^{.1}	0.25 mg ml ⁻¹	0.5 mg ml ⁻¹	1 mg ml ⁻¹		
Thymol	100 ± 0.00ª	66.6 ± 4.44 ^b	19.6 ± 2.44°	0 ± 0.00	0 ± 0.00	0 ± 0.00		
Carvacrol	100 ± 0.00^{a}	100 ± 0.00ª	78.3 ± 3.50 ^b	0 ± 0.00	0 ± 0.00	0 ± 0.00		
Eugenol	100 ± 0.00^{a}	100 ± 0.00ª	100 ± 0.00^{a}	68 ± 2.60 ^b	0 ± 0.00	0 ± 0.00		
Menthol	100 ± 0.00^{a}	100 ± 0.00ª	100 ± 0.00^{a}	100 ± 0.00ª	100 ± 0.00ª	70.3 ± 5.10 ^b		

Values are means (n=3) ± SD, different letters indicate significant differences (p<0.05) by LSD test.

 Table 5: Percentage of germinated conidia after 20 h of incubation at 27°C.

response to very low concentrations of thymol and carvacrol (0.125 mg ml⁻¹), while eugenol only halved the growth potential at a concentration of 0.25 mg ml⁻¹. Menthol was the least active inhibitor tested in the present study, since at the concentration of 0.5 mg ml⁻¹ the reduction of growth was only 39.8%.

Table 2 also shows the partial inhibitory concentrations (PIC) of MCs. Thymol, the most active compound, inhibited 90% of growth with half of its MIC value (0.125 mg ml⁻¹).

Determination of anti-fusarium effect of MCs in liquid medium

The effect of MCs on the growth of Fod in liquid medium is shown in Tables 3 and 4. MCs exhibited varying levels of antifungal activity against Fod. Table 3 shows that the MIC values of MCs in liquid medium ranged between 0.5 mg ml⁻¹ and 4 mg ml⁻¹, with a potent effect of thymol and carvacrol (0.5 mg ml⁻¹) followed by eugenol (1mg ml⁻¹). Menthol showed the least antifungal activity with a MIC value of 4 mg ml⁻¹. The MFC values of MCs ranged between 1 mg ml⁻¹ and more than 10 mg ml⁻¹. Thymol and carvacrol had the lowest MFC value (1 mg ml⁻¹) followed by eugenol (2 mg ml $^{-1}$). Menthol did not show any fungicidal effect even with the highest concentration (10 mg ml $^{-1}$).

The results depicted on Table 4 show that thymol and carvacrol were the most efficient MCs with an inhibition reaching 95.3% and 90.5%, respectively, at low concentration (0.25 mg ml⁻¹) followed by eugenol with a percentage of inhibition of 92.2% at 0.5 mg ml⁻¹. Menthol was the less active MC with a percentage of inhibition of 97.5% at relatively high concentration (2 mg ml⁻¹).

Fod conidial germination assay

The effect of MCs on conidia germination of Fod is shown in Table 5. The results in Table 5 show that the most effective inhibitors on Fod conidial germination were thymol and carvacrol (with an inhibition value of 100% at 0.25 mg ml⁻¹). For eugenol, a complete inhibition of the conidial germination was obtained at 0.5 mg ml⁻¹, while the effect of menthol was the lowest.

The conidia destruction assay

The effect of different concentrations of MCs on Fod conidia

Figure 1: time course of percentage of conida destruction treated by different concentrations of MC. a: Thymol, b: Carvacrol, c: Eugenol, d: Menthol. (Values are means (n=3) \pm SD). Concentrations of MC: (\blacklozenge): 0.0625 mg ml⁻¹, (\blacksquare): 0.125 mg ml⁻¹, (\blacksquare): 0.25 mg ml⁻¹, (\blacksquare): 0.5 mg ml⁻¹, (\blacksquare): 1.25 mg ml⁻¹.

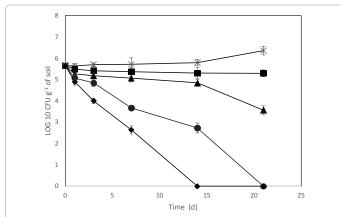


Figure 2: Soil densities of Fod over time as affected by soil treatment with different concentrations of thymol. (Values are means LOG 10 transformation of CFU cm³ of soil. Data represent average of 3 trials of experiment with three replications per treatment per trial). Concentrations of thymol: (×): 0 mg ml⁻¹, (■): 0.25 mg ml⁻¹, (●): 1 mg ml⁻¹, (♦): 2 mg ml⁻¹).

destruction is shown in Figure 1. Thymol had the strongest destruction effect on conidia. Indeed, at the concentration of 1 mg ml⁻¹ and after only 3 h of incubation the conidia were completely destroyed (Figure 1a). At the same concentration, carvacrol needed 6 h to have the same effect obtained with thymol (Figure 1b). On the other hand, eugenol and menthol needed 24 h to destroy respectively 92% and 69% of conidia (Figures 1c and 1d).

Fusarium density in the soil

Treatment of soil with aqueous emulsions of Thymol is shown in Figure 2.

Figure 2 shows significant differences in soil densities of Fod. For

the untreated lot, soil density of Fod increased after 21 days from 5.63 Log_{10} (CFU cm⁻³) to 6.35 Log_{10} (CFU cm⁻³) whereas the treated lot with 0.5 mg ml⁻¹ of thymol has shown a decrease after 21 days from 5.26 Log_{10} (CFU cm⁻³) to 3.56 Log_{10} (CFU cm⁻³). For the treated lot with 2 mg ml⁻¹ of thymol, we noticed a total disappearance of Fod after 14 days.

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Discussion

Choice of anti-fungal agents

Essential oils are a complex mixture of many components, and are known for having an anti-fungal activity Daferera, Barrera-Necha, Ahmad Khan, et al. [15-17]. However, the determination of the active molecules was challenging. Therefore, we have chosen to use pure MC such as thymol, carvacrol, eugenol and menthol.

Determination of MIC in agar medium and MIC and MFC in liquid medium

The Results in this part of the current study show a MIC value of 0.25 mg ml⁻¹ for thymol and carvacrol, and 0.5 mg ml⁻¹ for eugenol. These MIC values are much lower than the values described by Nguefack, Dambolena, Menniti et al. [18-21] with other fusarium species. The difference in values is most likely due to the use of different techniques; more precisely the size of conidia inoculum [22], and the use of detergents or solvents that influence the antifungal activity [23].

We have also determined the MIC and MFC in liquid medium. The results obtained from this liquid medium show that the MIC (99.9% inhibition of growth) of the thymol is 0.5 mg ml⁻¹. Although this value is twice the MIC obtained in agar medium, such difference was expected due to the use of different experimental methods. In fact, other laboratories have observed similar differences [24,25].

We opted for this method that relies on both the MIC (99.9% inhibition) and the concentrations that give partial inhibitions (PICs) because it allows better comparison between the agents used. Additionally, this method allowed us to come up with certain conclusions. In fact, while thymol and carvacrol have the same MIC value in agar medium (0.25 mg ml⁻¹), thymol showed a drastic growth inhibition (89%), whereas carvacrol only showed 54% growth inhibition at the $\frac{1}{2}$ MIC value (0.125 mg ml⁻¹). We therefore conclude that thymol would be more effective than carvacrol.

Within field applications, the aim is not to completely eradicate the mold, but rather to diminish the fungal burden and growth [26].

Calculated MFC values were twice the MIC value, except for menthol, where the maximum concentration used (10 mg m^{-1}) was found to be incapable of exerting a fungicidal action.

This difference between MIC and MFC has been reported by several authors on different Fusarium species [27-29] and other mold species [15,17].

The results of the MIC and MFC obtained in agar and liquid media thus show that thymol and carvacrol possess an anti-*Fusarium oxysporum* dose-dependent activity with values much lower than those described by the other laboratories [30].

Effect of MC on germination of Fod conidia

Since we used an inoculum of conidia in our work, we asked how MCs act on conidia. We therefore decided to evaluate the action of 4 MCs on the germination of conidia.

We worked on a liquid medium as described by Remmal et al.

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[23], we observed the behavior of conidia under microscope in a timedependent manner. Additionally, other laboratories used the same strategy to evaluate germination [21].

The results obtained show an important inhibition of germination at concentrations below the MIC in liquid medium, especially for thymol, followed by carvacrol and eugenol. Results demonstrated this inhibition through a continued reduction in the number of conidia until their eventual disappearance at concentrations above the MIC. These results also show a delayed germination effect at 20 h as indexed by the MC and concentration used. Using thymol as an example, ¼ of MIC caused 80% germinative inhibition and ½ of MIC caused 100% germinative inhibition.

These results are in agreement with results obtained by Dambolena et al. and Remmal et al. [21,31] using other *Fusarium* species, however, the concentrations inhibiting the germination are superior. This difference in concentrations between our lab and theirs is most likely due to methodological reasons cited above. These results led us to ask the following question: Do MCs act inside the fungal cells by modifying the cellular metabolism as previously described by Isman, Chaillot et al. and Divband et al. [32-34] or they act through modifying the structural integrity of the cell as described by Gao et al. [35].

In an attempt to answer this mechanistic question, we perceived three important phenomena through microscopic observation of conidia germination:

1. The decrease in number of conidia corresponded to the concentration of MC.

2. The presence of broken conidia cells, which have lost their intracellular content.

3. The presence of swollen and deformed conidia.

Based on these observations, we believe that the effect of MCs acts through modifying the structural integrity of the cell [35], previously obtained similar observations.

We decided to evaluate the ability of MC to destroy conidia by performing a microscopic count on the Malassez Counting chamber of conidia treated in a non-nutrient medium.

Effect of MC on the destruction of Fod conidia

The results obtained show a dose-dependent and time-dependent decrease in the number of conidia. The results also show that thymol at a concentration equal to MIC in liquid medium (0.5 mg ml⁻¹) causes 100% destruction of the conidia after 6 h, whereas carvacrol at the same concentration causes 100% destruction after 24 h.

As for thymol at the concentration equal to the MFC, a total destruction of the conidia was obtained after 3 h, with 93% destruction of conidia obtained after just 1 h. This rapidity of destructive action would be in favor of thymol for the direct action on the envelope of conidia. This hypothesis is borne out by the work of Bennis and Chami et al. [6,7] which shows a similar action of oregano EO and carvacrol on *Saccharomyces* using a scanning electron microscope (SEM). Moreover, the work of Xing et al. [29] shows the action of cinnamaldehyde on hyphae and conidia of *Fusarium verticillioides* using an SEM and TEM (transmission electron microscope).

On a fundamental level, advanced studies should be able to establish the relationship between the direct action of MC on the envelope of conidia and the changes observed intracellularly on the expression of certain genes [33]. On an applied level, we thought about the possibility of utilizing these results in the agricultural field to fight Fusarium wilt.

Fusarium oxysporum attacks plants through the roots to ascend into the vessels [36]. One of the established methods to fight *Fusarium* is soil decontamination using fungicidal agents before planting [37]. We therefore decided to test soil decontamination using thymol, since it is the MC which proved to be the most effective.

Decontamination of the soil by thymol

We artificially infested soil samples (previously sterilized) with 10⁶ conidia g^1 of soil, treating these samples with different concentrations of thymol and closely monitoring the Fusarium load in each soil sample.

Our results show that 21 days after treatment, the number of conidia increased 10 times in the untreated batch, thereby demonstrating that under our experimental conditions, Fusarium was physiologically capable of surviving and multiplying.

The results also show that at half the MIC (0.25 mg ml⁻¹) the curve remained almost constant whereas a concentration equal to the MIC (0.5 mg ml⁻¹) caused a decrease of 95% from day 14. The concentration equal to MFC caused a very large decrease from day 14 to day 21, where there was no *Fusarium* observed.

These results encouraged us to pre-test the action of thymol on carnations planted in Fod artificially infested soil. The results obtained show a significant reduction of the Fusarium load in the soil compared to the untreated control. This reduction was reflected in the health status of treated plants, which produced much larger roots and stem than plants planted in untreated infested soil (results not shown).

These results are quite promising for the development of a Fusarium treatment using thymol, a natural product safe for farmers, the environment, and consumers. This thymol-based treatment would be an excellent alternative to currently used fungicides such as methyl bromide, chloropicrin, and benomyl which are toxic to the farmers, the consumers and the environment [2].

Conclusion

This work is a fundamental contribution toward showing the antifusarium action of MCs, with the possible development of natural alternatives capable of replacing chemical fungicides currently used in agriculture, specifically floral agriculture.

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Conflict of Interest

No conflict of interest declared.

References

- Borrero C, Trillas I, Avilés M (2009) Carnation fusarium wilt suppression in four composts. European Journal of Plant Pathology 123: 425-433.
- Amini J, Sidovich DF (2010) The effects of fungicides on *Fusarium oxysporium*. sp. lycopersici associated with Fusarium wilt of tomato. Journal of Plant Protection Research 50: 172-178.
- Budnik LT, Kloth S, Velasco-Garrido M, Baur X (2012) Prostate cancer and toxicity from critical use exemptions of methyl bromide: Environmental protection helps protect against human health risks. Environment Health 11: 1-5.
- 4. Hou Y, Luo, Q, Chen C, Zhou M (2013) Application of tetra primer ARMS-PCR

approach for detection of *Fusarium graminearum* genotypes with resistance to carbendazim. Australas. Plant Pathology 42: 73-78.

- Gullino ML, Katan J, Garibaldi A (2012) Fusarium wilts of greenhouse vegetable and ornamental crops. American Phytopathological Society, St. Paul, Minnesota. 191-198 p.
- Bennis S, Chami N, Chami F, Bouchikhi T, Remmal A (2004) Surface Alteration of Saccharomyces Cerevisiae Induced by Thymol and Eugenol. Letter of Applied Microbiology 38: 454-458.
- Chami F, Chami N, Bennis S, Bouchikhi T, Remmal A (2005) Oregano and Clove Essential Oils Induce Surface Alteration of Saccharomyces Cerevisiae. Phytotherapy Research 19: 405-408.
- Bouddine L, Louaste B, Achahbar S, Chami N, Chami F, et al. (2012) Comparative study of the antifungal activity of some essential oils and their major phenolic components against Aspergillus niger using three different methods. Afr J Biotechnol 11: 14083-14087.
- 9. Leslie JF, Summerell BA, Bullock S (2006) The Fusarium laboratory manual. Blackwell Publishing, Ames, Iowa, USA. 212-218 p.
- Fothergill AW (2012) Antifungal susceptibility testing: Clinical laboratory and standards institute (CLSI) methods. In: Hall GS, editor. Interactions of Yeasts, Moulds, and Antifungal Agents: How to Detect Resistance, Hamana Press, New York, USA. 65-74 p.
- Neimann CJ, Baayen RP (1989) Inhibitory effects of Pnenylserine and Salicylic Acid on Phytoalexin Accumulation in Carnation Infected by *Fusarium oxysporum* f. sp. dianthi. Med. Fac. Landbouww. Rijksuniv. Gent 54/2a, pp. 435-438.
- Remmal A, Bouchikhi T, Tantaoui-Elaraki A, Ettayebi M (1993) Inhibition of antibacterial activity of essential oils by Tween 80 and ethanol in liquid medium. J Pharm Belg 48: 352-356.
- Pandey DK, Tripathi NN, Tripathi RD, Dixit SN (1982) Fungitoxic and phytotoxic properties of the essential oil of Hyptis suaveolens. J Plant Dis Protect 89: 344-349.
- Tzortzakis NG, Economakis CD (2007) Antifungal activity of lemongrass (*Cympopogon citratus* L.) essential oil against key postharvest pathogens. Innovative Food Science and Emerging Technology 8:253-258.
- Daferera DJ, Ziogas BN, Polissiou MG (2003) The effectiveness of plant essential oils on the growth of Botrytis cinerea, *Fusarium* sp. and *Clavibacter* michiganensis sub sp. michiganensis `crop proection 22: 39-44.
- Barrera-Necha LL, Garduño CY, García-Barrera J (2009) In vitro antifungal activity of essential oils and their compounds on mycelial growth of *Fusarium oxysporum* f. sp. gladioli (Massey) Snyder and Hensen. Plant Pathology Journal 8:17-21.
- 17. Ahmad Khan MS, Ahmad I (2011) In vitro antifungal, anti-elastase and antikeratinase activity of essential oils of *Cinnamomum-*, *Syzygium-* and *Cymbopogon* species against *Aspergillus fumigates* and *Trichophyton rubrum*. Phytomedicine 19: 48-55.
- Nguefack J, Leth V, Amvam Zollo PH, Mathur SB (2004) Evaluation of five essential oils from aromatic plants of Cameroon for controlling food spoilage and mycotoxin producing fungi. International Journal of Food Microbiology 94: 329-334.
- Dambolena JS, López AG, Cánepa MC, Theumer MG, Zygadlo JA, et al. (2008) Inhibitory effect of cyclic terpenes (Limonene, menthol, menthone and thymol) on *Fusarium verticillioides* MRC 826 growth and fumonisin B1 biosynthesis. Toxicon 51: 37-44.
- 20. Menniti AM, Gregori R, Neri F (2010) Activity of natural compounds on

Fusarium verticillioides and fumonisin production in stored maize kernels. Int J Food Microbiol 136:304-309.

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- Dambolena JS, López AG, Meriles JM, Rubinstein HR, Zygadlo J (2012) Inhibitory effect of 10 natural phenolic compounds on Fusarium verticillioides, a structure, property and activity relationship study. Food Control 28: 163-170.
- 22. Pujol I, Guarro J, Sala J, Riba MD (1997) Effects of incubation temperature, inoculum size, and time of reading on broth microdilution susceptibility test results for amphotericin B against Fusarium. Antimicrobial Agents Chemotherapy 41: 808-811.
- Remmal A, Tantaoui-Elaraki A, Bouchikhi T, Rhayour K, Ettayebi M (1993) Improved method for the determination of antimicrobial activity of essential oils in agar medium. Journal of Essential Oils Research 5: 1179-1184.
- Kalemba D, Kunicka A (2003) Antibacterial and antifungal properties of essential oils. Current Medicinal Chemistry 10: 813-829.
- 25. Abbaszadeh S, Sharifzadeh A, Shokri H, Khosravi AR, Abbaszadeh A (2014) Antifungal efficacy of thymol, carvacrol, eugenol and menthol as alternative agents to control the growth of food-relevant fungi. Journal de Mycologie Medicale 24: 51-56.
- Koul O, Walia S, Dhaliwal GS (2008) Essential oils as green pesticides: potential and constraints. Biopesticides International 4: 63-84.
- 27. Gill TA, Li J, Saenger M, Scofield SR (2016) Thymol based sub micron emulsions exhibit antifungal activity against *Fusarium graminearum* and inhibit fusarium head blight (FHB) in wheat. Journal of Applied Microbiology 121: 1103-1116.
- Horváth G, Jenei JT, Vágvölgyi C, Böszörményi A, Krisch J (2016) Acta Biologica Hungarica. Acta Biol Hung. 67: 205-214.
- Xing F, Hua H, Selvaraj JN, Zhao Y, Zhou L, et al. (2014) Growth inhibition and morphological alterations of Fusarium verticillioides by cinnamon oil and cinnamaldehyde. Food Control 46: 343-350.
- Naeini A, Ziglari T, Shokri H, Khosravi AR (2010) Assessment of growthinhibiting effect of some plant essential oils on different *Fusarium* isolates. Journal Mycologie médicale 20: 174-178.
- 31. Taskeen-Un-Nisa WA, Bhat MY, Pala SA, Mir RA (2011) In vitro inhibitory effect of fungicides and botanicals on mycelial growth and spore germination of *Fusarium oxysporum*. Journal of Biopesticides 4: 53-56.
- 32. Isman MB (2000) Plant essential oils for pest and disease management. Crop Protection 19: 603-608.
- 33. Chaillot J, Tebbji F, Remmal A, Boone C, Brown GW, et al. (2015). The monoterpene carvacrol generates endoplasmic reticulum stress in the pathogenic fungus *Candida albicans*. Antimicrobial Agents Chemotherapy 59: 4584-92.
- 34. Divband K, Shokri H, Khosravi AR (2017) Down-regulatory effect of *Thymus vulgaris* L. on growth and Tri4 gene expression in *Fusarium oxysporum* strains. Microbial Pathogenesis 104: 1-5.
- 35. Gao T, Zhou H, ZhouW, Hu L, Chen J, et al. (2016) The fungicidal activity of thymol against *Fusarium graminearum* via inducing lipid peroxidation and disrupting ergosterol biosynthesis. Molecules 21: 770.
- Fravel D, Olivain C, Alabouvette C (2003) Fusarium oxysporum and its biocontrol. New Phytologist 157: 493-502.
- Bowers JH, Locke LC (2000) Effect of botanical extracts on the population density of *Fusarium oxysporum* in soil and control of Fusarium wilt in the green house. Plant Disease 84: 300-305.