(January-March, 2015)



GLOBAL JOURNAL OF BIOLOGY, AGRICULTURE & HEALTH SCIENCES (Published By: Global Institute for Research & Education)

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# GASTRIC CANCER RISK IN HELICOBACTER PYLORI INFECTED PATIENTS: A SYSTEMATIC OVERVIEW

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# Abstract

*Helicobacter pylori* is a gastric pathogen that colonizes approximately 50% (over 3 billion) of the world's population, mainly in the developing countries. Infection with *H. pylori* leads to a chronic inflammatory condition called severe chronic atrophic gastritis (SCAG) and significantly increases the risk of developing duodenal and gastric ulcer disease and gastric cancer. Infection with *H. pylori* is the strongest known risk factor for gastric cancer. Gastric cancer ranks fourth in incidence and second in mortality among all cancers worldwide. People with SCAG have an increased risk of gastric adenomas in both the upper and lower parts of the stomach.

Although *H. pylori* affects a large percentage of the population, only a small percentage of carriers develop this malignancy. Recent investigations have begun to identify the factors that lead to these complications. Such clinical diversities are caused by variations of *H. pylori* pathogenicity, host susceptibility, environmental factors, and interactions of these factors. The exact mechanisms underlying how *H. pylori* triggers or causes gastric cancer remain elusive.

Key words: *Helicobacter pylori*; *Helicobacter pylori* infection; Severe chronic atrophic gastritis (SCAG); Gastric cancer.

# **1. Introduction**

The human gastric pathogen *Helicobacter pylori* (Hp) infection plays a crucial role in gastric cancer pathogenesis (You *et al.*, 2000; Peek and Blaser, 2002; IARC Monogr Eval Carcinog Risks Hum., 1994). It is the major cause of chronic gastritis, peptic ulcers, and gastric malignancies, including gastric non-cardia adenocarcinoma and mucosal-associated lymphoid tissue (MALT) lymphoma (Peek and Crabtree, 2006). It accounts for up to two thirds of gastric cancer cases (Parkin, 2006). The estimated incidence in 2008 was 989 600, and the majority of new cases occurred in developing countries (American Cancer Society, 2011). The incidence of gastric cancer has declined dramatically in most countries over the past 70 years. It remains the fourth most common cancer and the second most frequent cause of cancer deaths (Hohenberger and Gretschel, 2003; Parkin, 2004; Parkin *et al.*, 2002), accounting for 10.4% of cancer deaths worldwide (Brenner *et al.*, 2009) with high incidence in definite area (China, Eastern Europe, and Japan) (Bruckner *et al.*, 2003). Reasons for reductions in overall gastric cancer incidence and mortality have not been fully elucidated, but changes in lifestyle/environmental factors and improved health care could be possible factors. These include decreased intake of salted and preserved foods due to use of fridges, increased consumption of fruits and vegetables, reduced chronic Hp infection owing to better hygiene and medication, mass screening measures to detect precancerous lesions in some regions such as Japan, and decreased smoking in developed countries (Bertuccio *et al.*, 2009; Jemal *et al.*, 2010).

Persistent inflammation caused by Hp infection induces hypochlorhydria and chronic atrophic gastritis of the bodyfundus, which are two early precursors of gastric cancer development (Testino *et al.*, 2004) as shown below (Israel and Peek, 2001) (Fig. 1). However, although the prevalence of Hp infection ranges from 40 to 80% in humans, only a small proportion (probably < 3%) of infected patients develops gastric cancer (Peek and Blaser, 2002). Genetic variation in genes encoding cytokines and their receptors, which determine the intensity of the inflammatory response to the bacteria, may contribute to individual differences in severity of outcome of Hp infection and progression of gastric lesions (Gonzalez *et al.*, 2002). It infects the gastric mucosa leading to an acute followed by chronic inflammatory response, accompanied by the production of several proinflammatory cytokines. These cytokines enhance the immune response and inhibit gastric acid secretion. Consequently, an excessive production of gastrin and free radicals ultimately lead to neoplastic transformation of the gastric mucosa (Matthews and Butler, 2005). Individual differences in the intensity of the inflammatory response may contribute to gastric mucosa transformation (Gonzalez *et al.*, 2002).

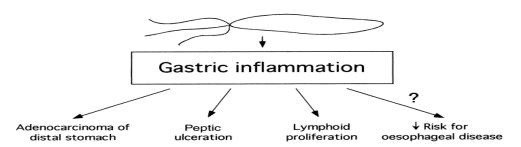


Figure 1: Relationship of Hp-induced gastric inflammation with variable disease outcomes. (Adapted from Israel and Peek, 2001).

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Certain cytokine gene polymorphisms have been associated with the occurrence of gastric cancer, with the most consistent results referring to the increased gastric cancer risk associated to *IL1B-511*, *IL1RN* variable number tandem repeat (*VNTR*), and *TNFA-308*, despite the heterogeneous findings across previous meta-analyses (Camargo *et al.*, 2006; Gorouhi *et al.*, 2008; Kamangar *et al.*, 2006; Loh *et al.*, 2009; Vincenzi *et al.*, 2008; Wang *et al.*, 2007; Zhang *et al.*, 2008). Different risk estimates have been described according to the histologic type of the tumor, with stronger associations for the intestinal type (Camargo *et al.*, 2006; Gorouhi *et al.*, 2008; Kamangar *et al.*, 2006; Wang *et al.*, 2007). These tumors are the most frequent (Lauren and Nevalainen, 1993) and are preceded by a set of sequential precancerous lesions (Correa *et al.*, 1975), from which intestinal metaplasia is much more frequent than dysplasia (Uemura *et al.*, 2001) and more strongly associated with gastric cancer than gastric atrophy. Addressing the potential associations between the cytokine gene polymorphisms and gastric precancerous lesions may contribute to the understanding of some of the previous heterogeneous findings from studies having gastric cancer as the outcome. Here, we review the recent advances in our understanding of the association of *H. pylori* infection and the risk of gastric cancer.

# 2. Diversity of *H. pylori* and Variations in their Genome

### 2.1 H. pylori Populations

There are three types of bacterial population structure: clonal, panmictic and endemic (Achtman, 2004). If intraspecies or inter-species recombination is rare, the genetic diversity of a bacterial species predominantly comes from evolution of the ancestry. This species has a clonal population structure. In a species with high frequency of recombination, introduction of foreign gene fragments into the genome occurs frequently in the evolution history. As foreign genes have a different evolution history, the evolution speed of individual genes is different. In this case, the species possess a panmictic structure. For a bacterial species with a panmictic structure, a temporal clonal structure may occur if it rapidly spread among naive hosts. In this situation, a bacterial species has an endemic structure.

### 2.2 Genomic Variations

Hp shows great inter-strain variation in genetic content (Suerbaum and Achtman, 1999). None of the individual strains is identical as demonstrated by multiple fingerprinting methods (Akopyanz *et al.*, 1992; Han *et al.*, 2003). Sequence divergence is the main cause of this variation. Hp diversity researches on the human population sampling from Asia, Africa, and South America demonstrated that Hp-human coevolution has been for about 58,000 years (Linz *et al.*, 2007; Yamaoka *et al.*, 2002; Falush *et al.*, 2003; Devi *et al.*, 2007). This bacterium is natural competence cell and developed a specified Type IV Secretion System (T4SS), the comB-system, to integrate exogenous DNA into its genome through genetic recombination (Hofreuter *et al.*, 2003; Hofreuter *et al.*, 1998). Human stomach has low bacterial diversity on the level of species but is rich in genetic variants in subpopulations of Hp. The maintenance of high diversification makes this bacterium to cope with particular challenges in individual hosts (Kang and Blaser, 2006).

### 2.3 Cytotoxin-associated Gene A (CagA)

At the phenotypic level, strains can be characterized into two types: those that contain a gene associated with cytotoxin expression, the so called *CagA* (cytotoxin-associated gene A) gene, and those that do not (Telford *et al.*, 1994; Xiang *et al.*, 1995). *CagA* positive Hp, which comprise some 50%-60% of United States isolates, cause more extensive inflammation of the gastric mucosa than *CagA* negative strains (Crabtree *et al.*, 1995; Cover *et al.*, 1995; Crabtree *et al.*, 1991; Peek *et al.*, 1995; Blaser, 1995). *CagA* positive (strains carrying the *cag PAI*) infections are also more likely to progress to atrophic gastritis than *CagA* negative infections (Kuipers *et al.*, 1995). Furthermore, a nested case-control study among Japanese-American men suggested that *CagA* antibodies are more common in infected persons with gastric malignancy than in infected persons without such malignancy (Blaser *et al.*, 1995). Whereas this finding was not statistically significant, the two-fold increase in *CagA* positive Hp among intestinal type cancers appeared unlikely to be due to chance alone.

### 2.4 H. pylori Strains, CagA and Genomic Size

A number of strains of Hp have been sequenced to date (Alm *et al.*, 1999; Tomb *et al.*, 1997; Baltrus *et al.*, 2009; Clancy *et al.*, 2012; Lehours *et al.*, 2011). Of these, the origin and other information of 30 strains are publicly available. These include 14 strains from hpEastAsia (7 from hspEasia and 7 from hspAmerind subpopulations, respectively), 10 from hpEurope, 5 from hpAfrica1 and 1 from hpAfrica2 (Schott *et al.*, 2011; Kawai *et al.*, 2011; Avasthi *et al.*, 2011; Kersulyte *et al.*, 2010). All, except for strain B38 from hpEurope, possess *CagA* and the *Cag* pathogenicity island (*cag PAI*). The genomic size of *CagA*-positive Hp ranges from approximately 1.55 mbp to 1.71 mbp with an average of 1.61 mbp. For *CagA*-negative strains, their genome is generally smaller because of the lack of the *Cag* pathogenicity island of about 40 kbp. The average genomic sizes of *CagA*-positive Hp strains from different populations have been reported (Devi *et al.*, 2010; Thiberge *et al.*, 2010; Farnbacher *et al.*, 2010; Dong *et al.*, 2009; McClain *et al.*, 2009; Oh *et al.*, 2006; Dong *et al.*, 2012). The average genomic size of strains from hpEurope is approximately 1.65 mbp, which is significantly larger than that from hpEastAsia (1.60 mbp, P < 0.05) or hpAfrica1 (1.60 mbp).

The *cag PAI* is a 40-kb DNA fragment which contains 27 to 31 genes flanked by 31-bp direct repeats (Censini *et al.*, 1996). It encodes *CagA*, the major virulence determinant of Hp and components of a type IV secretion system (Covacci *et al.*, 1993; Kutter *et al.*, 2008). The latter translocates *CagA* into host cells (Odenbreit *et al.*, 2000). Once inside the host cells, *CagA* binds to a number of host cell proteins disrupting intracellular signaling systems via tyrosine phosphorylation-dependent or -independent pathways (Murata-Kamiya, 2011). This causes elongation and loss of polarity of host cells, promoting proliferation and inflammation. The presence of the *cag PAI* in Hp is associated with increased risk of severe gastritis, atrophic gastritis, and distal gastric cancer compared with strains that lack the *cag* island (Israel *et al.*, 2001; Miehlke *et al.*, 2000; Plummer *et al.*, 2000).

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A marked difference lies between hpEurope and hpEast- Asia in the prevalence of strains possessing the *cag PAI*. Approximately 60% to 70% of Western Hp strains express *CagA* (Miehlke *et al.*, 2000; Owen *et al.*, 2001), indicating the presence of the *cag PAI*.

Nine Hp genome sequences are available from public databases to date: 26695 (accession number: AE000511) (Tomb *et al.*, 1997), J99 (AE001439.1) (Alm *et al.*, 1999), P12 (EMBL:CP001217, EMBL:CP001218 for plasmid]) (Fischer *et al.*, 2010), HPAG1 (CP000241, CP000242 for plasmid) (Oh *et al.*, 2006), or G27 (CP001173,CP001174 from plasmid) (Baltrus *et al.*, 2009), Shi470 (CP001072) (Devi *et al.*, 2006), B38 (FM991728) (Thiberge *et al.*, 2010), 51(CP000012) and 52 (CP001680). These nine genomes represent the genetic information of isolates from patients with various diseases (from gastritis to cancer) from different geographic regions (Alm *et al.*, 1999). Several articles on the comparisons of Hp genomes have been published (Alm *et al.*, 1999; Tomb *et al.*, 1997; Baltrus *et al.*, 2009; Clancy *et al.*, 2012; Lehours *et al.*, 2011; Schott *et al.*, 2011; Kawai *et al.*, 2011; Avasthi *et al.*, 2011; Kersulyte *et al.*, 2010; Devi *et al.*, 2009; McClain *et al.*, 2009; Oh *et al.*, 2006; Devi *et al.*, 2006; Lara-Ramírez *et al.*, 2011).

# 3. Risk of Gastric Cancer Associated with H. pylori Infection

### 3.1 Prevalence of Hp Infection

Hp is present in the stomach of more than half of the world population. Its infection has an association with the occurrence of gastrointestinal diseases, including gastric inflammation, peptic ulcer, gastric mucosa-associated lymphoid-tissue (MALT) lymphoma and gastric cancer. It is a key strategy to prevent and treat these diseases by eradicating Hp (Hohenberger and Gretschel, 2003). In the MALT-lymphoma patients, approximately 50% of cases were diagnosed with gastrointestinal non-Hodgkin's lymphoma; most are linked to *Hp* infection. In the early stages of low-grade MALT lymphoma, 60-80% can be cured by *Hp* eradication (Wotherspoon *et al.*, 1993; Chen *et al.*, 2005; Stathis *et al.*, 2009). It has been implicated in the pathogenesis of several gastrointestinal, systemic or hematological diseases (Papagiannakis *et al.*, 2013). Gastric cancer occurs in only a minority of infected individuals, however. Such clinical diversities are caused by variations of Hp pathogenicity, host susceptibility, environmental factors, and interactions of these factors. Based on compelling epidemiological evidences, the International Agency for Research on Cancer, World Health Organization (IARC/WHO) concluded in 1994 that Hp had a causal linkage to gastric carcinogenesis and is a definite carcinogen in humans (Sugiyama, 2004).

#### 3.2 Risk Factors of Hp Infection

Hp infection is the most common infectious disease in the world (Pounder and Ng, 1995). The prevalence of Hp ranges from less than 10% to over 80% in children (Jafar *et al.*, 2013) and nearly 50% of the world's population is estimated to be infected (Achtman, 2004). The risk factors for Hp infection include socioeconomic status, household crowding, ethnicity, migration from high prevalence regions, and infection status of family members.

# 4. Diagnostic Tests and Treatment Strategies

Diagnostic tests for Hp are generally divided into two categories: invasive and noninvasive. Invasive tests comprise the histological examination of gastric specimens. Noninvasive tests are based on peripheral samples such as blood, breath, stools, urine, and saliva, in order to detect antibodies, bacterial antigens, or urease activity. The choice of a specific test always depends on local experience and clinical settings, but usually a combination of two methods is often recommended since, for example, the detection of Hp-specific antibodies does not ultimately reflect a current infection (Bauer and Meyer, 2011).

Treatment of infection relies on a combination of antimicrobial agents and antisecretory agents, the elevation of the gastric pH by antisecretory agents being required for the bactericidal effect of the antimicrobial agents. Alternatively, although the mechanism of action is not yet clear, phytomedicines and probiotics have been used to improve eradication of Hp. The effect of antimicrobial agents and antisecretory agents depends not only on their pharmacological activities, but also on their pharmacokinetic properties. Many antimicrobial agents, including amoxicillin, clarithromycin, levofloxacin, metronidazole, tetracycline, rifabutin, and bismuth-containing compounds, have been used for Hp therapy, while the main antisecretory agents used are proton pump inhibitors (PPIs) (Yang *et al.*, 2014).

Although Hp is sensitive to a wide range of antibiotics *in vitro*, they all fail when applied as monotherapy *in vivo*. Therefore, a combined therapeutic strategy is used, usually including two antibiotics (clarithromycin, combined with amoxicillin or metronidazole) and either a bismuth compound or a proton pump inhibitor (PPI). Rarely, quadruple therapies are used in which the bismuth compound and PPI are used in combination with two antibiotics. The use of these drugs has resulted in effective therapies, with eradication rates over 80%. During the past several years, however, resistant bacteria have been detected constantly (Kist, 2007; Wueppenhorst *et al.*, 2009), leading to the search for alternative drugs and treatment strategies.

In the past decade, much effort has been devoted to the development of vaccination strategies. Based on the successful elimination of *Helicobacter felis* after mucosal immunization of mice with urease (Saldinger *et al.*, 1998), the focus of much research has been the induction of a humoral or Th2-driven immune response. To date, however, effective vaccination has only been observed in animal models and no human vaccine trial has been successful (Aebischer *et al.*, 2008). The failure to replicate the success of the vaccine in humans may be due to differences in Hp-specific immune responses or anatomical differences of the stomach. For instance, one of the main surface bacterial virulence factors, *cagPAI*, is usually switched off in mice.

Vaccines and antibiotics are not the only ways to prevent and cure Hp infection or Hp-associated disease. Hppositive individuals infected with helminthes have standard levels of Hp colonization rates and gastritis patterns, but they develop significantly less Hp-associated disease (Du *et al.*, 2006; Elshal *et al.*, 2004). These are intriguing observations (January-March, 2015)

that might result in low-dose administration of immunomodulating agents to Hp-positive patients, which have the same consequences as enteric helminth infections.

Another approach is the application of probiotics. There is convincing evidence that Hp is killed by *Lactobacilli* both *in vitro* and to a limited extent *in vivo* (Lorca *et al.*, 2001; Midolo *et al.*, 1995; Sakamoto *et al.*, 2001). Furthermore, *Lactobacilli* show a positive impact on some Hp therapy-related side effects, and recent studies suggest that *Lactobacilli* supplements could be effective in increasing eradication rates (Zou *et al.*, 2009).

# 5. Environmental, Genetic Factors and Gastric Cancer

Among the environmental and lifestyle factors, tobacco use is reported as the most risky factor (Gonzalez *et al.*, 2003). It causes to develop gastritis ulcers, intestinal metaplasia and finally, gastric cancer. There is strong evidence supporting tobacco use aggravates Hp infection in most cases. Many researchers reported the connection of high salt intake with gastric cancer development. The *in vitro* studies revealed the virulence of Hp bacteria on oncogenicity with increased concentration of salt in the medium. This is due to the increased expression of oncogenic protein (*CagA*). Studies on 36 *cagA*-positive Hp *strains* from Colombian patients established considerable heterogeneity in salt-regulated *CagA* expression (Loh *et al.*, 2012).

Many Hp-associated diseases including peptic ulcer, gastric cancer, and MALT lymphoma only develop decades after infection (Fig. 2), their medical burden is tremendous. Gastric cancer is one of the most common forms of cancer; with approximately 700,000 to 900,000 new cases diagnosed every year, and the second leading cause of cancer-related deaths worldwide (Parkin *et al.*, 2002). Survival rates are very low, ranging from 15% if diagnosed during later stages of the disease to 65% if diagnosed early. Incidence rates vary widely geographically, and, in general, more males than females are affected (50% lower incidence). Although high-risk areas in Japan, China,

Eastern Europe, and certain Latin America countries still remain (WHO, 2009), incidence rates worldwide have been declining for several decades (Bertuccio *et al.*, 2009).

Gastric adenocarcinomas are mainly divided into two histologically distinct forms, diffuse-type gastric adenocarcinoma, and intestinal-type adenocarcinoma (Lauren classification) each exhibiting different epidemiological and pathophysiological features (Lauren, 1965). Diffuse-type gastric adenocarcinoma is found predominantly in younger people, with no gender bias. It consists of individually infiltrating neoplastic cells that do not form glandular structures and are not associated with intestinal metaplasia (Polk and Peek, 2010). The more prevalent form of gastric adenocarcinoma is called intestinal-type adenocarcinoma, which usually occurs in elderly people, predominates inmen, and progresses through a well-defined chain of histological events, typically starting with a transition from normal mucosa to chronic gastritis, followed by atrophic gastritis and intestinal metaplasia which finally ends in dysplasia and adenocarcinoma (Correa, 1992). Hp significantly increases the risk of developing both cancer types. We focus here mainly on the association between Hp and intestinal-type adenocarcinoma. Gastric cancers are also classified by their localization within the stomach: the most important distinction being between cardia (the proximal part of the stomach) and noncardia. Hp is the strongest risk factor for the development of non-cardia (distal) gastric cancer.

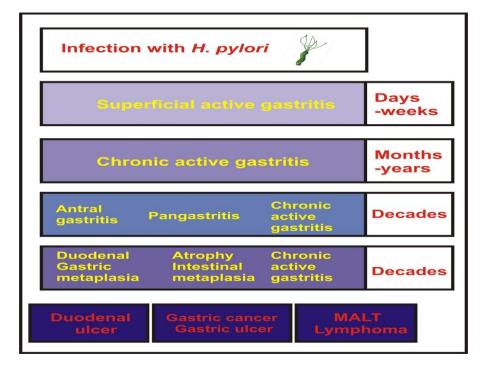


Figure 2: Time line of disease progression in Hp-infected persons. All infected individuals develop a superficial gastritis within the first weeks of infection, followed by a chronic active gastritis which develops after months or years. After decades, patients can develop antral gastritis or pangastritis, depending on the localization of the infection. The antral inflammation can lead to gastric metaplasia, which supports the growth of duodenal ulcer. The latter can lead to atrophy and intestinal metaplasia, two prerequisites for the development of gastric cancer or gastric ulcer. In contrast, constant chronic active gastritis can lead to the growth of MALT lymphomas. (Adapted from Telford *et al.*, 1997).

### 6. Host Genetic Factors

Host genetic factors, as polymorphisms in inflammatory and immune response genes, are mainly related to the recognition of the bacteria by the immune system and the variation in the level of cytokine response (Garza-González *et al.*, 2005). Among host factors, several inflammatory proteins including cytokines, growth factors, and chemokines have been known to control immune response against Hp infection (Achyut *et al.*, 2007; Trejo-de la *et al.*, 2008). Therefore, many studies have focused on the analyses of polymorphisms in genes associated with the inflammatory response in the gastric mucosa and risk for malignancy (Rad *et al.*, 2009; Kumar *et al.*, 2009; Melo Barbosa *et al.*, 2009; Partida-Rodríguez *et al.*, 2010). Extensive epidemiology studies have shown that Hp infection is a major risk factor for gastric cancer and its precursor lesions (Eslick *et al.*, 1999). The risk of developing gastric cancer is estimated to increase 2 to 6 times in patients with Hp infection, as determined by retrospective case-control and prospective epidemiology studies (Covacci *et al.*, 1993). Among individuals infected with Hp, a small percentage develops gastric cancer by a process influenced by bacterial virulence.

# 7. Hp Virulence Factor

The most widely studied Hp virulence factor is the *CagA* antigen, a 96-to 138-kDa protein (Arents *et al.*, 2001). The *CagA* gene, found on a genomic region called the *cag* pathogenicity island (*PAI*), is considered as a marker for enhanced virulence. Moreover, individuals infected with *CagA*-positive Hp strains have a higher risk of developing peptic ulcers and gastric cancer compared to those harboring *CagA*-negative Hp strains (Arents *et al.*, 2001).

The role of Hp infection on gastric cancer is yet to be confirmed but two related mechanisms by which Hp could promote cancer are under investigation. One mechanism involves the enhanced production of free radicals near Hp and an increased rate of host cell mutation. The other proposed mechanism has been called a "perigenetic pathway" and involves enhancement of the transformed host cell phenotype by means of alterations in cell proteins, such as adhesion proteins. Hp has been proposed to induce inflammation and locally high levels of  $TNF-\alpha$  and/or interleukin 6 (*IL-6*). According to the proposed perigenetic mechanism, inflammation-associated signaling molecules, such as  $TNF-\alpha$ , can alter gastric epithelial cell adhesion and lead to the dispersion and migration of mutated epithelial cells without the need for additional mutations in tumor suppressor genes, such as genes that code for cell adhesion proteins (Tsuji *et al.*, 2003; Suganuma *et al.*, 2008).

The entire mechanism underlying tumorigenesis and metastasis of gastric cancer is not yet known. The involvement of signal transducer and activator of transcription 3 (STAT3) in GA is reported by Wang *et al.* (Wang *et al.*, 2013). The STAT3 can be activated by tyrosine phosphorylation in response to growth factors and cytokines ([IL]-6). Inerleukin-6 and its signaling component STAT3, play essential roles in the process of inflammation and abnormal immunity as well as carcinogenesis (Bowman *et al.*, 2000; Hodge *et al.*, 2005; Huang, 2007). Under certain abnormal conditions, STAT3 continues, to trigger oncogene transcription (Kim *et al.*, 2009; Yu and Jove, 2004). This leads to the progression of GA with metastasis.

### 8. Conclusion

Hp infection contributes to the development of diverse gastric and extragastric diseases which are both important public health burdens which could be largely eliminated by Hp eradication. Gastric cancer develops in persons infected with Hp but not in uninfected persons. Those with histologic findings of severe gastric atrophy, corpus-predominant gastritis, or intestinal metaplasia are at increased risk. Persons with Hp infection and nonulcer dyspepsia, gastric ulcers, or gastric hyperplastic polyps are also at risk, but those with duodenal ulcers are not (Uemura *et al.*, 2001). Undoubtedly, future studies must be undertaken to clarify further the role of Hp in the four gastrointestinal disorders discussed in this systematic overview, especially gastric cancer. The infection is necessary but not sufficient for the development of gastric adenocarcinoma. Its eradication would eliminate a major worldwide cause of cancer death, therefore there is much interest in identifying how, if, and when this can be accomplished. There are several mechanisms by which Hp contributes to the development of gastric cancer. Gastric adenocarcinoma is one of many cancers associated with inflammation, which is induced by Hp infection, yet the bacteria also cause genetic and epigenetic changes that lead to genetic instability in gastric epithelial cells. Hp eradication reduces both. However, many factors must be considered in determining whether treating this bacterial infection will prevent cancer or only reduce its risk-these must be considered in designing reliable and effective eradication therapies (Graham, 2015).

Despite of intensive investigation, the mechanisms of Hp-induced gastric cancer remain poorly understood, the stem cell hypothesis hold the promise to elucidate the origin/initiation of gastric cancer. Furthermore, important questions such as (a) variations in disease susceptibility in different populations, (b) gastric cancer progression in relation to Hp virulence genes polymorphism, and (c) the correlation of stem cell with different types of gastric cancer are all waiting for further clarification.

Hp infection triggers inflammation, interactions of bacteria with host cell in local microenvironment also affect gastric stem/progenitor cells and their differentiation, this may potentiate oncogenic transformation. Therefore, focus on Hp-induced molecular pathogenesis and the impact of microenvironment in gastric stem or progenitor cells will be crucial to identify the molecular targets in tumor initiation and the origin of gastric cancer; investigation of which will also provide insights to uncover the carcinogenic mechanisms and options for cancer intervention and prevention (Ding and Zheng, 2012).

Furthermore, Hp infection has been proposed to provide some benefits, such as reducing the risks of obesity or childhood asthma, although there are no convincing data to support the benefits of Hp infections (Graham, 2015).

# References

Achtman, M. (2004). Population structure of pathogenic bacteria revisited. Int J Med Microbiol., 294 pp. 67–73.

Achyut, B. R., Ghoshal, U. C., Moorchung, N., Mittal, B. (2007). Association of Toll-like receptor-4 (Asp299Gly and Thr399Ileu) gene polymorphisms with gastritis and precancerous lesions. Hum Immunol., 68 pp. 901–907.

Aebischer, T., Bumann, D., Epple, H. J., Metzger, W., Schneider, T., Cherepnev, G., Walduck, A. K., Kunkel, D., Moos, V., Loddenkemper, C., Jiadze, I., Panasyuk, M., Stolte, M., Graham, D. Y., Zeitz, M., Meyer, T. F. (2008). "Correlation of T cell response and bacterial clearance in human volunteers challenged with *Helicobacter pylori* revealed by randomised controlled vaccination with Ty21a-based Salmonella vaccines," Gut, 57(8) pp. 1065–1072.

Akopyanz, N., Bukanov, N. O., Westblom, T. U., Kresovich, S., Berg, D. E. (1992). DNA diversity among clinical isolates of *Helicobacter pylori* detected by PCR-based RAPD fingerprinting. Nucleic Acids Res., 20 pp. 5137–5142.

Alm, R. A., Ling, L. S., Moir, D. T., King, B. L., Brown, E. D., Doig, P. C., Smith, D. R., Noonan, B., Guild, B. C., deJonge, B. L., Carmel, G., Tummino, P. J., Caruso, A., Uria-Nickelsen, M., Mills, D. M., Ives, C., Gibson, R., Merberg, D., Mills, S. D., Jiang, Q., Taylor, D. E., Vovis, G. F., Trust, T. J. (1999). Genomic-sequence comparison of two unrelated isolates of the human gastric pathogen *Helicobacter pylori*. Nature, 397 pp. 176–180.

American Cancer Society. Global Cancer Facts and Figures. 2<sup>nd</sup> ed. Atlanta: American Cancer Society, 2011.

Arents, N. L., van Zwet, A. A., Thijs, J. C., Kooistra-Smid, A. M., van Slochteren, K. R., Degener, J. E., Kleibeuker, J. H., van Doorn, L. J. (2001). The importance of *vacA*, *cagA*, and *iceA* genotypes of *Helicobacter pylori* infection in peptic ulcer disease and gastroesophageal reflux disease. Am J Gastroenterol., 96, pp. 2603–2608.

Avasthi, T. S., Devi, S. H., Taylor, T. D., Kumar, N., Baddam, R., Kondo, S., Suzuki, Y., Lamouliatte, H., Mégraud, F., Ahmed, N. (2011). Genomes of two chronological isolates (*Helicobacter pylori* 2017 and 2018) of the West African *Helicobacter pylori* strain 908 obtained from a single patient. J Bacteriol., 193 pp. 3385–3386.

Baltrus, D. A., Amieva, M. R., Covacci, A., Lowe, T. M., Merrell, D. S., Ottemann, K. M., Stein, M., Salama, N. R., Guillemin, K. (2009). The complete genome sequence of *Helicobacter pylori* strain G27. J Bacteriol., 191 pp. 447–448.

Bauer, B., Meyer, T. F. (2011). The Human Gastric Pathogen *Helicobacter pylori* and Its Association with Gastric Cancer and Ulcer Disease. Ulcers, pp. 1–23.

Bertuccio, P., Chatenoud, L., Levi, F. (2009). "Recent patterns in gastric cancer: a global overview," International Journal of Cancer, 125(3) pp. 666–673.

Bertuccio, P., Chatenoud, L., Levi, F., Praud, D., Ferlay, J., Negri, E., Malvezzi, M., La Vecchia, C. (2009). Recent patterns in gastric cancer: a global overview. Int J Cancer, 125, pp. 666–673.

Blaser, M. J. (1995). Intrastrain differences in Helicobacter pylori: a key question in mucosal damage? Ann Med., 27 pp. 559–563.

Blaser, M. J., Perez-Perez, G. I., Kleanthous, H., Cover, T. L., Peek, R. M., Chyou, P. H., Stemmermann, G. N., Nomura, A. (1995). Infection with *Helicobacter pylori* strains possessing *cagA* is associated with an increased risk of developing adenocarcinoma of the stomach. Cancer Res., 55 pp. 2111–2115.

Bowman, T., Garcia, R., Turkson, J., Jove, R. (2000). STATs in oncogenesis. Oncogene, 19 pp. 2474–2488.

Brenner, H., Rothenbacher, D., Arndt, V. (2009). Epidemiology of stomach cancer. Methods Mol Biol., 472, pp. 467-477.

Bruckner, H. W., Kufe, D. W., Pollock, R. E., Weichselbaum, R. R. (2003). Cancer Medicine, Ontario, Canda.

Camargo, M. C., Mera, R., Correa, P., Peek, R. M. Jr., Fontham, E. T., Goodman, K. J., Piazuelo, M. B., Sicinschi, L., Zabaleta, J., Schneider, B. G. (2006). Interleukin-1β and interleukin-1 receptor antagonist gene polymorphisms and gastric cancer: a metaanalysis. Cancer Epidemiol Biomarkers Prev., 15, pp. 1674–1687.

Censini, S., Lange, C., Xiang, Z., Crabtree, J. E., Ghiara, P., Borodovsky, M., Rappuoli, R., Covacci, A. (1996). *cag*, a pathogenicity island of *Helicobacter pylori*, encodes type I-specific and disease associated virulence factors. Proc Natl Acad Sci USA, 93 pp. 14648–14653.

Chen, L. T., Lin, J. T., Tai, J. J., Chen, G. H., Yeh, H. Z., Yang, S. S., Wang, H. P., Kuo, S. H., Sheu, B. S., Jan, C. M., Wang, W. M., Wang, T. E., Wu, C. W., Chen, C. L., Su, I. J., Whang-Peng, J., Cheng, A. L. (2005). Long term results of anti *Helicobacter pylori* therapy in early stage gastric high-grade transformed MALT lymphoma. J Natl Cancer Inst., 97 pp. 1345–1353.

Clancy, C. D., Forde, B. M., Moore, S. A., O'Toole, P. W. (2012). Draft genome sequences of *Helicobacter pylori* strains 17874 and P79. J Bacteriol., 194 pp. 2402.

Correa, P. (1992). "Diet modification and gastric cancer prevention," Journal of the National Cancer Institute. Monographs, 12(12) pp. 75–78.

Correa, P., Haenszel, W., Cuello, C., Tannenbaum, S., Archer, M. (1975). A model for gastric cancer epidemiology. Lancet, 2 pp. 58–60.

Covacci, A., Censini, S., Bugnoli, M., Petracca, R., Burroni, D., Macchia, G., Massone, A., Papini, E., Xiang, Z., Figura, N. (1993). Molecular characterization of the 128-kDa immunodominant antigen of *Helicobacter pylori* associated with cytotoxicity and duodenal ulcer. Proc Natl Acad Sci USA, 90 pp. 5791–5795.

Cover, T. L., Glupczynski, Y., Lage, A. P., Burette, A., Tummuru, M. K., Perez-Perez, G. I., Blaser, M. J. (1995). Serologic detection of infection with *cagA+ Helicobacter pylori* strains. J Clin Microbiol., 33 pp. 1496–1500.

Crabtree, J. E., Covacci, A., Farmery, S. M., Xiang, Z., Tompkins, D. S., Perry, S., Lindley, I. J., Rappuoli, R. (1995). *Helicobacter pylori* induced interleukin-8 expression in gastric epithelial cells is associated with *CagA* positive phenotype. J Clin Pathol., 48 pp. 41–45.

Crabtree, J. E., Taylor, J. D., Wyatt, J. I., Heatley, R. V., Shallcross, T. M., Tompkins, D. S., Rathbone, B. J. (1991). Mucosal IgA recognition of *Helicobacter pylori* 120 kDa protein, peptic ulceration, and gastric pathology. Lancet, 338 pp. 332–335.

Devi, S. H., Taylor, T. D., Avasthi, T. S., Kondo, S., Suzuki, Y., Megraud, F., Ahmed, N. (2010). Genome of *Helicobacter pylori* strain 908. J Bacteriol., 192 pp. 6488–6489.

Devi, S. M., Ahmed, I., Francalacci, P., Hussain, M. A., Akhter, Y., Alvi, A., Sechi, L. A., Mégraud, F., Ahmed, N. (2007). Ancestral European roots of *Helicobacter pylori* in India. BMC Genomics, 8 pp. 184.

Devi, S. M., Ahmed, I., Khan, A. A., Rahman, S. A., Alvi, A., Sechi, L. A., Ahmed, N. 2006). Genomes of *Helicobacter pylori* from native Peruvians suggest admixture of ancestral and modern lineages and reveal a western type *cag*-pathogenicity island. BMC Genomics, 7 pp. 191.

Ding S-Z., Zheng P-Y. (2012). *Helicobacter pylori* infection induced gastric cancer; advance in gastric stem cell research and the remaining challenges. Gut Pathogens, 4:18.

Dong, Q. J., Wang, Q., Xin, Y. N., Li, N., Xuan, S. Y. (2009). Comparative genomics of *Helicobacter pylori*. World J Gastroenterol., 15 pp. 3984–3991.

Dong, Q. J., Zhan, S. H., Wang, L. L., Xin, Y. N., Jiang, M., Xuan, S. Y. (2012). Relatedness of *Helicobacter pylori* populations to gastric carcinogenesis. World J Gastroenterol., 18(45) pp. 6571–6576.

Du, Y., Agnew, A., Ye, X-P., Robinson, P. A., Forman, D., Crabtree, J. E. (2006). "*Helicobacter pylori* and *Schistosoma japonicum* co-infection in a Chinese population: helminth infection alters humoral responses to *H. pylori* and serum pepsinogen I/II ratio," Microbes and Infection, 8(1) pp. 52–60.

Elshal, M. F., Elsayed, I. H., El Kady, I. M. (2004). "Role of concurrent *S. mansoni* infection in *H. pylori*-associated gastritis: a flow cytometric DNA-analysis and oxyradicals correlations," Clinica Chimica Acta, 346(2) pp. 191–198.

Eslick, G. D., Lim, L. L., Byles, J. E., Xia, H. H., Talley, N. J. (1999). Association of *Helicobacter pylori* infection with gastric carcinoma: a meta-analysis. Am J Gastroenterol., 94, pp. 2373–2379.

Falush, D., Wirth, T., Linz, B., Pritchard, J. K., Stephens, M., Kidd, M., Blaser, M. J., Graham, D. Y., Vacher, S., Perez-Perez, G. I., Yamaoka, Y., Mégraud, F., Otto, K., Reichard, U., Katzowitsch, E., Wang, X., Achtman, M., Suerbaum, S. (2003). Traces of human migrations in *Helicobacter pylori* populations. Science, 299 pp. 1582–1585.

Farnbacher, M., Jahns, T., Willrodt, D., Daniel, R., Haas, R., Goesmann, A., Kurtz, S., Rieder, G. (2010). Sequencing, annotation, and comparative genome analysis of the gerbil-adapted *Helicobacter pylori* strain B8. BMC Genomics, 11 pp. 335.

Fischer, W., Windhager, L., Rohrer, S., Zeiller, M., Karnholz, A., Hoffmann, R., Zimmer, R., Haas, R. (2010). Strain specific genes of *Helicobacter pylori*: genome evolution driven by a novel type IV secretion system and genomic island transfer. Nucleic Acids Res., 38(18) pp. 6089–6101.

Garza-González, E., Bosques-Padilla, F. J., El-Omar, E., Hold, G., Tijerina-Menchaca, R., Maldonado-Garza, H. J., Pérez-Pérez, G. I. (2005). Role of the polymorphic *IL-1B*, *IL-1RN* and *TNF-A* genes in distal gastric cancer in Mexico. Int J Cancer, 114 pp. 237–241.

Gonzalez, C. A., Pera, G., Agudo, A., Palli, D., Krogh, V., Vineis, P., Tumino, R., Panico, S., Berglund, G., Simán, H., Nyrén, O., Agren, A., Martinez, C., Dorronsoro, M., Barricarte, A., Tormo, M. J., Quiros, J. R., Allen, N., Bingham, S., Day, N., Miller, A., Nagel, G., Boeing, H., Overvad, K., Tjonneland, A., Bueno-De-Mesquita, H. B., Boshuizen, H. C., Peeters, P., Numans, M., Clavel-Chapelon, F., Helen, I., Agapitos, E., Lund, E., Fahey, M., Saracci, R., Kaaks, R., Riboli, E. (2003). Smoking and the risk of gastric cancer in the European Prospective Investigation Into Cancer and Nutrition (EPIC). Int J Cancer, 107 pp. 629–634.

Gonzalez, C. A., Sala, N., Capella, G. (2002). Genetic susceptibility and gastric cancer risk. Int J Cancer, 100, pp. 249-260.

Gorouhi, F., Islami, F., Bahrami, H., Kamangar, F. (2008). Tumor-necrosis factor-A polymorphisms and gastric cancer risk: a metaanalysis. Br J Cancer, 98, pp. 1443–1451.

Graham D. Y. (2015). *Helicobacter pylori* Update: Gastric Cancer, Reliable Therapy, and Possible Benefits. Gastroenterology. Feb 2. pii: S0016-5085(15)00158-4. doi: 10.1053/j.gastro.2015.01.040.

Han, F. C., Ng, H. C., Ho, B. (2003). Stability of randomly amplified polymorphic DNA fingerprinting in genotyping clinical isolates of *Helicobacter pylori*. World J Gastroenterol., 9 pp. 2021–2024.

Hodge, D. R., Hurt, E. M., Farrar, W. L. (2005). The role of IL-6 and STAT3 in inflammation and cancer. Eur J Cancer, 41 pp. 2502–2512.

Hofreuter, D., Karnholz, A., Haas, R. (2003). Topology and membrane interaction of *Helicobacter pylori* ComB proteins involved in natural transformation competence. Int J Med Microbiol., 293 pp. 153–165.

Hofreuter, D., Odenbreit, S., Henke, G., Haas, R. (1998). Natural competence for DNA transformation in *Helicobacter pylori*: identification and genetic characterization of the comB locus. Mol Microbiol., 28 pp. 1027–1038.

Hohenberger, P., Gretschel, S. (2003). Gastric cancer. Lancet, 362, pp. 305-315.

Huang, S. (2007). Regulation of metastases by signal transducer and activator of transcription 3 signaling pathway: clinical implications. Clin Cancer Res., 13 pp. 1362–1366.

Israel, D. A., Peek, R. M. (2001). Review article: pathogenesis of *Helicobacter pylori*-induced gastric inflammation. Aliment Pharmacol Ther., 15, pp. 1271–1290.

Israel, D. A., Salama, N., Arnold, C. N., Moss, S. F., Ando, T., Wirth, H. P., Tham, K. T., Camorlinga, M., Blaser, M. J., Falkow, S., Peek, R. M. Jr. (2001). *Helicobacter pylori* strain-specific differences in genetic content, identified by microarray, influence host inflammatory responses. J Clin Invest., 107 pp. 611–620.

Jafar, S., Jalil, A., Soheila, N., Sirous, S. (2013). Prevalence of *Helicobacter pylori* Infection in Children, a Population-Based Cross-Sectional Study in West Iran. Iran J Pediatr., 23 pp. 13–18.

Jemal, A., Center, M. M., Desantis, C., Ward, E. M. (2010). Global patterns of cancer incidence and mortality rates and trends. Cancer Epidemiol Biomarkers Prev., 19, pp. 1893–1907.

Kamangar, F., Cheng, C., Abnet, C. C., Rabkin, C. S. (2006). Interleukin-1B polymorphisms and gastric cancer risk-a meta-analysis. Cancer Epidemiol Biomarkers Prev., 15, pp. 1920–1928.

Kang, J., Blaser, M. J. (2006). Bacterial populations as perfect gases: genomic integrity and diversification tensions in *Helicobacter pylori*. Nat Rev Microbiol., 4 pp. 826–836.

Kawai, M., Furuta, Y., Yahara, K., Tsuru, T., Oshima, K., Handa, N., Takahashi, N., Yoshida, M., Azuma, T., Hattori, M., Uchiyama, I., Kobayashi, I. (2011). Evolution in an oncogenic bacterial species with extreme genome plasticity: *Helicobacter pylori* East Asian genomes. BMC Microbiol., 11 pp. 104.

Kersulyte, D., Kalia, A., Gilman, R. H., Mendez, M., Herrera, P., Cabrera, L., Velapatiño, B., Balqui, J., Paredes Puente de la Vega, F., Rodriguez Ulloa, C. A., Cok, J., Hooper, C. C., Dailide, G., Tamma, S., Berg, D. E. (2010). *Helicobacter pylori* from Peruvian amerindians: traces of human migrations in strains from remote Amazon, and genome sequence of an Amerind strain. PLoS One, 5(11) pp. e15076.

Kim, D. Y., Cha, S. T., Ahn, D. H., Kang, H. Y., Kwon, C. I., Ko, K. H., Hwang, S. G., Park, P. W., Rim, K. S., Hong, S. P. (2009). STAT3 expression in gastric cancer indicates a poor prognosis. J Gastroenterol Hepatol., 24 pp. 646–651.

Kist, M. (2007). "*Helicobacter pylori*: primary antimicrobial resistance and first-line treatment strategies," Eurosurveillance, 12(7) pp. E1–2.

Kuipers, E. J., Pérez-Pérez, G. I., Meuwissen, S. G., Blaser, M. J. (1995). *Helicobacter pylori* and atrophic gastritis: importance of the *cagA* status. J Nadl Cancer Inst., 87 pp. 1777–1780.

Kumar, S., Kumar, A., Dixit, V. K. (2009). Evidences showing association of interleukin-1B polymorphisms with increased risk of gastric cancer in an Indian population. Biochem Biophys Res Commun., 387 pp. 456–460.

Kutter, S., Buhrdorf, R., Haas, J., Schneider-Brachert, W., Haas, R., Fischer, W. (2008). Protein subassemblies of the *Helicobacter pylori Cag* type IV secretion system revealed by localization and interaction studies. J Bacteriol., 190 pp. 2161–2171.

Lara-Ramírez, E. E., Segura-Cabrera, A., Guo, X., Yu, G., García-Pérez, C. A., Rodríguez-Pérez, M. A. (2011). New Implications on Genomic Adaptation Derived from the *Helicobacter pylori* Genome Comparison. PLoS ONE, 6(2) pp. e17300.

Lauren, P. (1965). "The two histological main types of gastric carcinoma: diffuse and so-called intestinal-type carcinoma. An attempt at a histo-clinical classification," Acta Pathologica et Microbiologica Scandinavica, 64 pp. 31–49.

Lauren, P. A., Nevalainen, T. J. (1993). Epidemiology of intestinal and diffuse types of gastric carcinoma: a time-trend study in Finland with comparison between studies from high- and low-risk areas. Cancer, 71 pp. 2926–2933.

Lehours, P., Vale, F. F., Bjursell, M. K., Melefors, O., Advani, R., Glavas, S., Guegueniat, J., Gontier, E., Lacomme, S., Alves Matos, A., Menard, A., Mégraud, F., Engstrand, L., Andersson, A. F. (2011). Genome sequencing reveals a phage in *Helicobacter pylori*. MBio, 2 pp. e00239–11.

Linz, B., Balloux, F., Moodley, Y., Manica, A., Liu, H., Roumagnac, P., Falush, D., Stamer, C., Prugnolle, F., van der Merwe, S. W., Yamaoka, Y., Graham, D. Y., Perez-Trallero, E., Wadstrom, T., Suerbaum, S., Achtman, M. (2007). An African origin for the intimate association between humans and *Helicobacter pylori*. Nature, 445 pp. 915–918.

Loh, J. T., Friedman, D. B., Piazuelo, M. B., Bravo, L. E., Wilson, K. T., Peek, R. M. Jr., Correa, P., Cover, T. L. (2012). Analysis of *Helicobacter pylori cagA* promoter elements required for salt-induced upregulation of *CagA* expression. Infect Immun., 80 pp. 3094–3106.

Loh, M., Koh, K. X., Yeo, B. H., Song, C. M., Chia, K. S., Zhu, F., Yeoh, K. G., Hill, J., Iacopetta, B., Soong, R. (2009). Metaanalysis of genetic polymorphisms and gastric cancer risk: variability in associations according to race. Eur J Cancer, 45 pp. 2562– 2568.

Lorca, G. L., Wadström, T., Font de Valdez, G., Ljungh, A. (2001). "*Lactobacillus acidophilus* autolysins inhibit *Helicobacter pylori in vitro*," Current Microbiology, 42(1) pp. 39–44.

Matthews, G. M., Butler, R. N. (2005). Cellular mucosal defense during *Helicobacter pylori* infection: a review of the role of glutathione and the oxidative pentose pathway. Helicobacter, 10, pp. 298–306.

McClain, M. S., Shaffer, C. L., Israel, D. A., Peek, R. M. Jr., Cover, T. L. (2009). Genome sequence analysis of *Helicobacter pylori* strains associated with gastric ulceration and gastric cancer. BMC Genomics, 10 pp. 3.

Melo Barbosa, H. P., Martins, L. C., Dos Santos, S. E., Demachki, S., Assumpção, M. B., Aragão, C. D., de Oliveira Corvelo, T. C. (2009). Interleukin-1 and *TNF-alpha* polymorphisms and *Helicobacter pylori* in a Brazilian Amazon population. World J Gastroenterol., 15 pp. 1465–1471.

Midolo, P. D., Lambert, J. R., Hull, R., Luo, F., Grayson, M. L. (1995). "In vitro inhibition of Helicobacter pylori NCTC 11637 by organic acids and lactic acid bacteria," Journal of Applied Bacteriology, 79(4) pp. 475–479.

Miehlke, S., Kirsch, C., Agha-Amiri, K., Günther, T., Lehn, N., Malfertheiner, P., Stolte, M., Ehninger, G., Bayerdörffer, E. (2000). The *Helicobacter pylori vacA* s1, m1 genotype and *cagA* is associated with gastric carcinoma in Germany. Int J Cancer, 87 pp. 322–327.

Murata-Kamiya, N. (2011). Pathophysiological functions of the CagA oncoprotein during infection by Helicobacter pylori. Microbes Infect., 13 pp. 799–807.

Odenbreit, S., Püls, J., Sedlmaier, B., Gerland, E., Fischer, W., Haas, R. (2000). Translocation of *Helicobacter pylori CagA* into gastric epithelial cells by type IV secretion. Science, 287 pp. 1497–1500.

Oh, J. D., Kling-Bäckhed, H., Giannakis, M., Xu, J., Fulton, R. S., Fulton, L. A., Cordum, H. S., Wang, C., Elliott, G., Edwards, J., Mardis, E. R., Engstrand, L. G., Gordon, J. I. (2006). The complete genome sequence of a chronic atrophic gastritis *Helicobacter pylori* strain: evolution during disease progression. Proc Natl Acad Sci USA, 103 pp. 9999–10004.

Owen, R. J., Peters, T. M., Varea, R., Teare, E. L., Saverymuttu, S. (2001). Molecular epidemiology of *Helicobacter pylori* in England: prevalence of *cag* pathogenicity island markers and IS605 presence in relation to patient age and severity of gastric disease. FEMS Immunol Med Microbiol., 30 pp. 65–71.

Papagiannakis, P., Michalopoulos, C., Papalexi, F., Dalampoura, D., Diamantidis, M. D. (2013). The role of *Helicobacter pylori* infection in hematological disorders. Eur J Intern Med., Pii:S0953–6205(13)00077–0.

Parkin, D. M. (2004). "International variation". Oncogene, 23, pp. 6329-6340.

Parkin, D. M. (2006). The global health burden of infection-associated cancers in the year 2002. Int J Cancer, 118, pp. 3030-44.

Parkin, D. M., Bray, F., Ferlay, J., Pisani, P. (2002). Global cancer statistics, 2002. CA Cancer J Clin., 55, pp. 74-108.

Partida-Rodríguez, O., Torres, J., Flores-Luna, L., Camorlinga, M., Nieves-Ramírez, M., Lazcano, E., Perez-Rodriguez, M. (2010). Polymorphisms in *TNF* and *HSP-70* show a significant association with gastric cancer and duodenal ulcer. Int J Cancer, 126(8) pp. 1861–1868.

Peek, R. M. Jr., Crabtree, J. E. (2006). Helicobacter infection and gastric neoplasia. J Pathol.; 208, pp. 233-248.

Peek, R. M. Jr., Miller, G. G., Tham, K. T., Perez-Perez, G. I., Zhao, X., Atherton, J. C., Blaser, M. J. (1995). Heightened inflammatory response and cytokine expression *in vivo* to *cagA*+ *Helicobacter pylori* strains. Lab Invest., 73 pp. 760–770.

Peek, R. M., Blaser, M. J. (2002). Helicobacter pylori and gastrointestinal tract adenocarcinomas. Nat Rev Cancer, 2, pp. 28–37.

Plummer, M., Vivas, J., Fauchère, J. L., Del Giudice, G., Peña, A. S., Ponzetto, A., Lopez, G., Miki, K., Oliver, W., Muñoz, N. (2000). *Helicobacter pylori* and stomach cancer: a case-control study in Venezuela. Cancer Epidemiol Biomarkers Prev., 9 pp. 961–965.

Polk, D. B., Peek, R. M. (2010). "Helicobacter pylori: gastric cancer and beyond," Nature Reviews Cancer, 10(6) pp. 403–414.

Pounder, R. E., Ng, D. (1995). The prevalence of *Helicobacter pylori* infection in different countries. Aliment Pharmacol Ther., 9 pp. 33–39.

Rad, R., Ballhorn, W., Voland, P., Eisenächer, K., Mages, J., Rad, L., Ferstl, R., Lang, R., Wagner, H., Schmid, R. M., Bauer, S., Prinz, C., Kirschning, C. J., Krug, A. (2009). Extracellular and intracellular pattern recognition receptors cooperate in the recognition of *Helicobacter pylori*. Gastroenterology, 136 pp. 2247–2257.

Sakamoto, I., Igarashi, M., Kimura, K., Takagi, A., Miwa, T., Koga, Y. (2001). "Suppressive effect of *Lactobacillus gasseri* OLL 2716 (LG21) on *Helicobacter pylori* infection in humans," Journal of Antimicrobial Chemotherapy, 47(5) pp. 709–710.

Saldinger, P. F., Porta, N., Launois, P., Louis, J. A., Waanders, G. A., Bouzouréne, H., Michetti, P., Blum, A. L., Corthésy-Theulaz, I. E. (1998). "Immunization of BALB/c mice with Helicobacter urease B induces a T helper 2 response absent in *Helicobacter* infection," Gastroenterology, 115(4) pp. 891–897.

Schistosomes, liver flukes and *Helicobacter pylori* (1994). IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Lyon, 7-14 June 1994. IARC Monogr Eval Carcinog Risks Hum. 61 pp. 1–241.

Schott, T., Kondadi, P. K., Hänninen, M. L., Rossi, M. (2011). Comparative genomics of *Helicobacter pylori* and the human-derived *Helicobacter bizzozeronii* CIII-1 strain reveal the molecular basis of the zoonotic nature of non-pylori gastric *Helicobacter* infections in humans. BMC Genomics, 12 pp. 534.

Stathis, A., Chini, C., Bertoni, F., Proserpio, I., Capella, C., Mazzucchelli, L., Pedrinis, E., Cavalli, F., Pinotti, G., Zucca, E. (2009). Long term outcome following *Helicobacter pylori* eradication in a retrospective study of 105 patients with localized gastric marginal zone B-cell lymphoma of MALT type. Ann Oncol., 20 pp. 1086–1093.

Suerbaum, S., Achtman, M. (1999). Evolution of Helicobacter pylori: the role of recombination. Trends Microbiol., 7 pp. 182-184.

Suganuma, M., Yamaguchi, K., Ono, Y., Matsumoto, H., Hayashi, T., Ogawa, T., Imai, K., Kuzuhara, T., Nishizono, A., Fujiki, H. (2008). *TNF-α*-inducing protein, a carcinogenic factor secreted from *H. pylori*, enters gastric cancer cells. Int J Cancer, 123(1) pp. 117–122.

Sugiyama, T. (2004). Development of gastric cancer associated with *Helicobacter pylori* infection. Cancer Chemother Pharmacol., 54(1) pp. S12–20.

Telford, J. L., Covacci, A., Ghiara, P., Montecucco, C., Rappuoli, R. (1994). Unravelling the pathogenic role of *Helicobacter pylori* in peptic ulcer: potential new therapies and vaccines. Trends Biotechnol., 12 pp. 420–426.

Telford, J. L., Covacci, A., Rappuoli, R., Chiara, P. (1997). "Immunobiology of *Helicobacter pylori* infection," Current Opinion in Immunology, 9(4) pp. 498–503.

Testino, G., De Iaco, F., Cornaggia, M. (2004). Role of atrophic gastritis of the body fundus and achlorhydia in the development of epithelial dysplasia and gastric carcinoma. Acta Gasterol Belg., 67, pp. 327–330.

Thiberge, J-M., Boursaux-Eude, C., Lehours, P., Dillies, M-A., Creno, S., Coppée, J-Y., Rouy, Z., Lajus, A., Ma, L., Burucoa, C., Ruskoné-Foumestraux, A., Courillon-Mallet, A., De Reuse, H., Boneca, I. G., Lamarque, D., Mégraud, F., Delchier, J-C., Médigue, C., Bouchier, C., Labigne, A., Raymond, J. (2010). From array-based hybridization of *Helicobacter pylori* isolates to the complete genome sequence of an isolate associated with MALT lymphoma. BMC Genomics, 11 pp. 368.

Tomb, J. F., White, O., Kerlavage, A. R., Clayton, R. A., Sutton, G. G., Fleischmann, R. D., Ketchum, K. A., Klenk, H. P., Gill, S., Dougherty, B. A., Nelson, K., Quackenbush, J., Zhou, L., Kirkness, E. F., Peterson, S., Loftus, B., Richardson, D., Dodson, R., Khalak, H. G., Glodek, A., McKenney, K., Fitzegerald, L. M., Lee, N., Adams, M. D., Hickey, E. K., Berg, D. E., Gocayne, J. D., Utterback, T. R., Peterson, J. D., Kelley, J. M., Cotton, M. D., Weidman, J. M., Fujii, C., Bowman, C., Watthey, L., Wallin, E., Hayes, W. S., Borodovsky, M., Karp, P. D., Smith, H. O., Fraser, C. M., Venter, J. C. (1997). The complete genome sequence of the gastric pathogen *Helicobacter pylori*. Nature, 388 pp. 539–547.

Trejo-de la, O. A., Torres, J., Pérez-Rodríguez, M., Camorlinga-Ponce, M., Luna, L. F., Abdo-Francis, J. M., Lazcano, E., Maldonado-Bernal, C. (2008). TLR4 single-nucleotide polymorphisms alter mucosal cytokine and chemokine patterns in Mexican patients with *Helicobacter pylori*-associated gastroduodenal diseases. Clin Immunol., 129 pp. 333–340.

Tsuji, S., Kawai, N., Tsujii, M., Kawano, S., Hori, M. (2003). Review article: inflammation-related promotion of gastrointestinal carcinogenesis - a perigenetic pathway. Aliment Pharmacol Ther., 1 pp. 82–89.

Uemura, N., Okamoto, S., Yamamoto, S., Matsumura, N., Yamaguchi, S., Yamakido, M., Taniyama, K., Sasaki, N., Schlemper, R. J. (2001). *Helicobacter pylori* infection and the development of gastric cancer. N Engl J Med., 345 pp. 784–789.

Vincenzi, B., Patti, G., Galluzzo, S., Pantano, F., Venditti, O., Santini, D., Ruzzo, A., Schiavon, G., Caraglia, M., Marra, M., Graziano, F., Tonini, G. (2008). Interleukin 1-511T gene (IL1β) polymorphism is correlated with gastric cancer in the Caucasian population: results from a meta-analysis. Oncol Rep., 20 pp. 1213–1220.

Wang, P., Xia, H. H., Zhang, J. Y., Dai, L. P., Xu, X. Q., Wang, K. J. (2007). Association of interleukin-1 gene polymorphisms with gastric cancer: a metaanalysis. Int J Cancer, 120 pp. 552–562.

Wang, Z., Si, X., Xu, A., Meng, X., Gao, S., Qi, Y., Zhu, L., Li, T., Li, W., Dong, L. (2013). Activation of STAT3 in Human Gastric Cancer Cells via Interleukin (IL)-6-Type Cytokine Signaling Correlates with Clinical Implications. PLoS One, 8(10) pp. e75788. World Health Organization, "Disease and injury country estimates," 2009.

Wotherspoon, A. C., Doglioni, C., Diss, T. C., Pan, L., Moschini, A., de Boni, M., Isaacson, P. G. (1993). Regression of primary low grade B-cell gastric lymphoma of mucosa associated lymphoid tissue type after eradication of *Helicobacter pylori*. Lancet, 342 pp. 572–577.

Wueppenhorst, N., Stueger, H. P., Kist, M., Glocker, E. (2009). "Identification and molecular characterization of triple- and quadruple-resistant *Helicobacter pylori* clinical isolates in Germany," Journal of Antimicrobial Chemotherapy, 63(4) pp. 648–653.

Xiang, Z., Censini, S., Bayeli, P. F., Telford, J. L., Figura, N., Rappuoli, R., Covacci, A. (1995). Analysis of expression of *CagA* and *VacA* virulence factors in 43 strains of *Helicobacter pylori* reveals that clinical isolates can be divided into two major types and that *CagA* is not necessary for expression of the vacuolating cytotoxin. Infect Immun., 63 pp. 94–98.

Yamaoka, Y., Orito, E., Mizokami, M., Gutierrez, O., Saitou, N., Kodama, T., Osato, M. S., Kim, J. G., Ramirez, F. C., Mahachai, V., Graham, D. Y. (2002). *Helicobacter pylori* in North and South America before Columbus. FEBS Lett., 517 pp. 180–184.

Yang, J-C., Lu, C-W., Lin, C-J. (2014). Treatment of *Helicobacter pylori* infection: Current status and future concepts. World J Gastroenterol., 20(18) pp. 5283–5293.

You, W. C., Zhang, L., Gail, M. H., Chang, Y. S., Liu, W. D., Ma, J. L., Li, J. Y., Jin, M. L., Hu, Y. R., Yang, C. S., Blaser, M. J., Correa, P., Blot, W. J., Fraumeni, J. F. Jr., Xu, G. W. (2000). Gastric dysplasia and gastric cancer: *Helicobacter pylori*, serum vitamin C and other risk factors. J Natl Cancer Inst.; 92, pp. 1607–1612.

Yu, H., Jove, R. (2004). The STATs of cancer - new molecular targets come of age. Nat Rev Cancer, 4 pp. 97-105.

Zhang, J., Dou, C., Song, Y., Ji, C., Gu, S., Xie, Y., Mao, Y. (2008). Polymorphisms of tumor necrosis factor- $\alpha$  are associated with increased susceptibility to gastric cancer: a meta-analysis. J Hum Genet., 53 pp. 479–489.

Zou, J., Dong, J., Yu, X. (2009). "Meta-analysis: lactobacillus containing quadruple therapy versus standard triple first-line therapy for *Helicobacter pylori* eradication," Helicobacter, 14(5) pp. 97–107.