Journal of Petroleum & Environmental Biotechnology

**Research Article** 

# Evaluation of Biogas Production by Anaerobic Digestion of Duckweed (*Lemna Minor*) and Cattle Manure

Alemayehu Negassa<sup>1</sup> and Dajene Fikadu<sup>2\*</sup>

<sup>1</sup>EnvironmentalSciences Program, Departmentof Biology, Ambo University, Ethiopia <sup>2</sup>Olankomi Comprehensive Secondary School, West Shoa, Ethiopia

## ABSTRACT

This study entailed evaluation of biogas production by anaerobic digestion of duckweed (Lemna minor) a plant that can be made easily available as much cheaper feedstock and cattle manure. Total solids, volatile solids, and organic carbon content of duckweed and cattle manure and pH of the slurries were determined using standard procedures. Three sets of plastic bottles were interconnected through connecting tube method. Digestion of duckweed to cattle manure in different percentage ratio (100:0, 75:25, 50:50, 25:75, 0:100) was performed respectively each in triplicates at 38°C temperature for 30 days at Ambo University plant Science Department laboratory. The cumulative biogas production in milliliters from 100% duckweed, 75% duckweed and 25% cattle manure; 50% duckweed and 50% cattle manure; 25% duckweed and 75% cattle manure, and 100% cattle manure was found to be 1015.5, 1040, 1159, 1206, and 862, respectively. Statistical analyses indicated significant differences between means of the physico-chemical parameters determined before and after anaerobic digestion (P<0.05) in all samples. The result revealed the attractive potential of duckweed as a feedstock in biogas production which peaked when the plant was co-digested with cattle manure at ratio of 25% to 75%.

Key words: Biogas; Renewable energy; Cow manure; Duckweed; Rumen fluid

# INTRODUCTION

The dependence on fossil fuels as primary energy source has led to global climate change, environmental degradation, and human health problems, and about 80% of the world's energy consumption still originates from combusting fossil fuels [1]. Besides, the rural population in developing countries including Ethiopia heavily depends on traditional fuels, such as Trewood, animal wastes and agricultural residues with quite significant adverse environmental impacts. Fast rate of population growth with increased energy demand and climate change have emerged as the most crucial issues that made researchers, practitioners and different organizations to search for renewable energy sources [2]. Moreover, rising energy prices and concerns about long term sustainability have once again brought renewable energy sources like biogas to the forefronts.

Biogas is produced through digestion of organic matter by anaerobic bacteria with end products consisting mainly of the combustible gas, methane (CH4) and a liquid effluent [3]. Biogas can be readily converted to electrical and thermal energy via a co-generator, typically for onsite consumption [4]. It is a proven eco-friendly technology that contributes to the reduction of the deforestation rate and helps to save the trees to sequester more carbon from the atmosphere and the local effects of trees being cut down that otherwise cause soil erosion, desertification, loss of soil fertility, and landslides [5]. Though evaluation of biogas production from different organic materials such as cattle manure and organic kitchen waste [6], poultry litter [7], and Khat (Catha edulis) waste [8]) has been reported so far no research has been conducted on duckweedin Ethiopia. Duckweed is the smallest flowering aquatic plant of immense biogas potential and can easily grow abundantly in nutrient reach waters including waste water with minimum cost and be made available as much cheaper feedstock. This study attempted to evaluate biogas production by anaerobic digestion of duckweed (Lemna minor) and Cattle manure.

## MATERIALS AND METHODS

Duckweed and cattle manure were used as substrates and rumen fluid was used as inoculum for biogas production in the experiment. About 15 kg of wet duckweed and 5 kg of fresh cattle manure were collected from wastewater ponds and dairy farm

Correspondence to: Dajene Fikadu, Olankomi Comprehensive Secondary School, West Shoa, Ethiopia; Email: falmineetfiqaaduu@gmail.com Received: February 03, 2021, Accepted: February 18, 2021, Published: February 25, 2021

**Citation:** Negassa A, Fikadu AM. (2021) Evaluation of Biogas Production by Anaerobic Digestion of Duckweed (Lemna Minor) and Cattle Manure. J Pet Environ Biotechnol 12: 413.

**Copyright:** © 2021 Negassa A, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

available at Ambo University main Campus, respectively. The collected substrates were dried with sun light and manually sorted to remove foreign materials and kept in a refrigerator at 4°C until used for the experiment [9]. The only biomass preprocessing that was required in this study was grinding into smaller uniform sized particles for samples of substrates.

Fresh rumen was taken from stomach of cow sacrificed for its meat and filtered through 0.5 mm sieve diameter to separate solid content from slurry. The rumen was added to the digesters and kept in anaerobic condition following Aurora et al. [10]. The study was conducted based on five samples of the substrates (Table 1) which were properly mixed before adding to the digesters.

 Table 1: Clinical and echocardiographic features of the study groups.

Samples of substrates	Corresponding digesters
100% Duckweed	А
75% Duckweed and 25% Cattle manure	В
50% Duckweed and 50% Cattle manure	С
25% Duckweed and 75% Cattle manure	D
100% Cattle manure	E

#### Design of the experiment

Duckweed (DW) and cattle manure (CM) were prepared as samples separately and also mixed in three different combinations. The experiment was done in triplicates for each sample. Thus, 15 digesters were charged once during the experimental period of 30 days. Since the focus of this study was mainly about biogas production, the experiment was conducted in a batch mode and all digesters were placed in an oven dry that was adjusted at 38°C.

#### Design of the experiment

Total solids (TS), volatile solids (VS) and organic content of samples of substrates and pH of the slurries were determined following the Standard Methods given in APHA [11].

#### Design of the experiment

Clean crucibles were stored at 105°C for 1 hour and then the mass of the empty crucibles were determined and recorded. Next, the mass of crucibles were measured with 6 g of samples of substrates. Then, the crucibles that contain samples were placed in oven at 105°C for 24 hours. This was mainly aimed to evaporate all the water from the samples to determine total solids (TS). Then, the crucibles containing dried samples of substrates were removed and cooled in desiccators. The mass of total solids was determined by subtracting mass of crucible from mass of crucible containing samples.

The percentage TS was calculated using the following formula.

#### Volatile solids

After total solids (TS) were determined, the oven dried samples of substrates that exist in crucibles were ignited at 550°C in a furnace for 3 hours. Next, crucibles that contain ash were cooled in desiccators and their masses were measured. The mass of volatile solids was determined by subtracting mass of crucible that contain ashes from that contain dried samples. The following formula was employed to calculate the percentage of volatile solids content of the samples of substrates.

#### pН

The pH before and after AD was determined using digital pH meter (HANNA HI 8314). An electrode was inserted into samples of substrates that were diluted using distilled water before inoculation of rumen fluid for measure pH before AD and the pH measurement after AD was done using pH electrode inserted into the fermentation slurry of substrates samples that were digested for about 30 days [12].

#### Organic carbon

Carbon content of the feedstock was obtained from volatile solids data using an empirical equation developed by Badger et al. [13] as

#### Determination of total fluid added to digesters

The anaerobic digestion of substrates samples were conducted in batch mode in 0.6 liter plastic bottles as digesters labeled for all substrate samples and 100.62 g of rumen fluid was added into all digesters of samples of substrates. To get 8% of total solids in the fermentation slurry, the total liquid (distilled water and rumen fluid) that was added to the samples of digesters was independently determined following Tchobanoglous et al. [14] as:

#### Biogas reactor

Sets of five plastic bottles each with 0.6 liter capacity were used as digesters. This was a modification of a compact system digester that digests small volumes of duckweed and cattle manure to produce biogas. The second sets of five plastic bottles with 0.6 liter capacity were used to contain acidified solution, and the third sets of other five plastic bottles were used as conical flasks each with 0.5 liter capacity. All the three set containers were interconnected with a plastic tube in batch mode [12]. The acidified brine solution was prepared by adding sodium chloride to distilled water until a supersaturated solution was formed to prevent the dissolution of biogas in the water following Tamrat Aragaw [6]. The supersaturated solution was formed by dissolving 40 g of sodium chloride into 100 ml of water at 20°C. Then, the five substrates samples were loaded into their digesters and each digester was connected to each of the second plastic bottle filled with acidified brine solution by means of a connecting tube.

Then, three drops of sulphuric acid were added using a dropper to acidify the brine solution. The biogas produced in the digesters by the fermentations of samples substrates passed through the connected tube to the second sets of plastic bottles containing acidified solution. The pressure of the biogas produced in the second bottles caused a displacement of acidified solution through a connected tube into the third sets of plastic bottles. Thus, the biogas produced by fermentation of the slurry was driven from the first sets of bottles to the second sets of bottles so as to displace a volume of the brine solution into the third sets of bottles which is equivalent to the volume of biogas produced. This displaced solution was measured by 250 ml of measuring cylinder in every 2 days in a month which represents the amount of biogas produced

As biogas production commenced in the fermentation chamber, it was delivered to the second chamber which contained the acidified brine solution. Since the biogas is insoluble in the solution, a pressure build-up provided the driving force for displacement of the solution. The displaced solution was measured to represent the amount of biogas produced. Since the experiment was done in triplicated, all samples of substrates triplicated in independent digesters and the main contents of the slurry in each digester were determined.

#### Laboratory analysis

All apparatus were properly cleaned and allowed to dry in the laboratory. Analyses of samples were done by taking some contents of the samples (about 6 grams). Fresh duckweed had high water content and hence only 560 g of the dried form was obtained from 15 kg of fresh duckweed collected. For the anaerobic digestion, 30 g of substrate(s) was/were used. Sample size of substrates was determined based on the dry mass of duckweed and the volume of digesters used for AD. Total solids and volatile solids were determined based on standard procedures. The carbon content of the feedstock was obtained from volatile solids data using an empirical equation as reported by Badger et al. The pH of the fermentation samples of substrates in each digester was measured and recorded using a pH meter. Paired simple t-test was run to compare physio-chemical parameters before and after AD.

### **RESULTS AND DISCUSSIONS**

The total solid content of all samples before AD ranged from 87.33% to 91.45% from 6 grams of the samples while that after AD ranged from 78.66% to 84.66%. The maximum TS before AD was measured in duckweed whereas the minimum was documented in cattle manure. High content of volatile fatty acids (VFAs) in the substrates can produce misleading results in TS and VS since they might volatilize from the substrate when they are first heated and thus, give total solids and volatile solids values that are too low. This in turn can produce incorrect estimates of biogas production, which depend on volatile solids. However, it did not occur in the present research study.

After AD, values of TS significantly decreased in all samples of substrates compared with that before AD. However, the extent of decrement was more pronounced in mixed samples of substrates than these parateones. Removal of TS suggests conversion to biogas and high removal was observed in 25% DW and 75% CM indicating the highest biogas production from the sample. The results revealed that total solids had positive impacts on biogas production by anaerobic digestion of duckweed and cattle manure.

The volatile solids before AD ranged from 78.51% to 80.49% and from 74.75% to 75.85% after AD. High reduction of VSwas measured in 25% DW and 75% CM mix substrates as compared to the rest samples of substrates. Removal of VS also suggests its conversion to biogas.

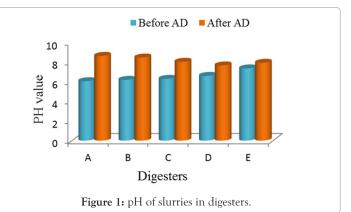
Total solids and volatile solids reduction are a good parameter for evaluating the efficiency of anaerobic digestion and it is a good indicator of biogas production. Similar results were reported by Joung et al. who studied methane production potential of anaerobic co-digestion from swine manure and food waste.

The volatile solids determined for duckweed and cattle manure substrates were 78.31% and 76.53%, respectively. This is in accordance with Fulford et al. who reported the volatile solids in animal and human wastes in the range from 77% to 90%. The ash content of duckweed was found to be much lower than cattle dung which indicated lower amount of non-biodegradable fractions of carbon and hence higher digestibility. This is in accordance with Yadav et al. who reported much greater ash content of cattle manure as compared to that of duckweed. Mean volatile solids before AD

are statistically significantly different from the mean volatile solid safter AD in all samples of substrates (P<0.05).

#### pH and organic carbon

The pH values of slurries after AD increased as compared to that before AD (Figure 1). This indicates that the rumen fluid used for this study had a good buffering capacity as reported earlier. The pH values of all slurries samples before AD ranged from 6.03 to 7.33 while that after AD ranged from 7.63 to 8.60. These results were in accordance with previous reports. The pH value of 100% CM was 7.33 which is optimum for biogas production, whereas that of 100% DW was 6.03 which is less optimal and in agreement. Slurries of mixed samples of substrates resulted in the rise of pH compared to that of DW, but a decrease in pH from CM slurry. The results indicated that co-digestion is a good way of adjusting the pH value to the optimum for duckweed, and in line.



Before AD, the pH was found to increase significantly with increasing of CM proportion in the mix suggesting that CM helps to maintain the pH to meet the optimum required. Increased pH values after AD might be due to the production of ammonia resulting from high organic matter available in duckweed than cattle manure which is supported with Gray et al. [6]. The statistical results indicated that the mean pH before AD is statistically significantly different from the one after AD in all samples (P<0.05).

In all samples reduction of organic carbon was observed after AD which might be because organic carbon can be assimilated, transformed, and decomposed by bacteria in anaerobic digestion process. This observation was in accordance which state that organic carbon can be removed in anaerobic digesters either by being converted to cellular materials for growth and reproduction of bacteria or biogas production. This is also in accordance with Devlin et al. who attributed the decrease in organic carbon to the degradation process during anaerobic digestion. The percent degradation of organic carbon for 25% DW and 75% CM was the highest compared with the rest samples of substrates, suggesting that mixing can enhance degradation and biogas production. Similar result has also been reported by Teame et al. from experiment on co-digestion of cow dung and cactus peel. The statistical results indicated significant difference (P<0.05) between mean organic carbon (OC) values before and after AD in all samples.

#### Evaluation of biogas production

Within initial six days high biogas production was observed from digestion of the digester containing only cattle dung as the feedstock. This may be due to the presence of higher amount of easily degradable organic carbon fractions in the cattle dung which is in line who reported presence of labile carbon sources in cattle dung that could be easily hydrolyzed by acedogenic and acetogenic bacteria. Provenzano et al. attributed the point to the fact that cattle dung contains more simple degradable monomers fermentative bacteria degrade these molecules to volatile fatty acids and to ammonia; methane formation occurs in which the acids are converted to biogas.

The production of biogas in the initial days shows that microorganisms did not need time to acclimatize to the substrate due to active inoculum and in agreement. Also the fact that gas production occurred at the beginning of the experiment suggests the existence of microbes in the added rumen fluid inoculum to act on readily degradable materials of the substrates was concluded by Kamthunzi. The biogas production from duckweed and cattle manure mixtures exceeded that from samples of each substrate due to the balanced (nutrient to microorganism) composition and stable pH which was attained from the inoculation with rumen fluid and mixing ratios used.

The amount of biogas produced from 100% DW exceeded the amount produced from 100% CM and peaked in 25% DW and 75% CM. This might be due to more availability of biodegradable material in DW than CM to serve as a source of energy for microbes in full process of digestion and in agreement. Macias-Corral also states that biogas production is a function of the feedstock's organic content and its biodegradability. The least production resulted in 100% CM probably might be partly due to the partial fermentation that usually takes place in the intestinal tract of the animal as reported by Deublein. Duckweed being a lignocellulosic biomass contains lignin and cellulose which needed more time to break down into more labile carbon moieties and thus gets converted into precursor for methanogenesis as reported by Facchin. The higher production from the mixtures could be due to a proper nutrient balance, increased buffering capacity, and decreased effect of toxic compounds resulting from mixing of the substrates.

As the proportion of DW in the mix ratio increased from 25 % to 75 %, the cumulative biogas yield decreased which is in agreement with Yadav. This observation was also in accordance with the results of an experiment done by Callaghan et al. using Water hyacinth, poultry manure and cow dung, where higher cumulative biogas production was produced in the system with the lower concentration of water hyacinth. This might be due to the high concentration of total nitrogen (ammonia) resulting from anaerobic breakdown of proteins to inhibit anaerobic digestion. Thus, co-digestion of CM and DW was more productive with DW proportion not exceeding 25%.

The present experiment indicated that duckweed has potential for biogas production. In addition to this, the percentage of volatile solid from total solid content of the duckweed substrate was 78.31% whereas that from Cow dung was 76.53% which indicated that a large fraction of duckweed was biodegradable. Based on the total production volume of biogas during a hydraulic retention time of 30 days, the optimum mixing for digestion of duckweed to cattle manure has been found to be at 25%:75% ratio. Biogas produced from duckweed was 15.17% greater than that produced from cattle manure.

# CONCLUSIONS AND RECOMMENDATIONS

The biogass produced from DW exceeded the amount produced from CM which indicated the potential of duckweed as important feedstock in biogas production. The study further revealed that an

# ACKNOWLEDGMENTS

We would like to thanktheHoleta Agricultural Research Center and Ambo University Plant Science Department for allowing us to use laboratory spaces and necessary facilities. We also thank MelkamuFikadu for his continuous technical supports.

## REFERENCES

- 1. Aurora SP. Microbial Digestion in Ruminants. Indian J Agr Res. 1983;2:85-94.
- 2. Badger CM, Bogue MJ, Stewart DJ. Biogas production from crops and organic wastes. J Sci 1979;22:11-20.
- Abubaker B, Ismail N. Anaerobic Digestion of Cow Dung for Biogas Production. ARPN J Eng Appl Sci. 2012;7(2):169-172
- Callaghan FJ, Wase DAJ, Thayanithy K, Forster CF. Co-digestion of waste organic solids: batch studies. Bioresour Technol. 1999;67(2):117-122.
- Devlin DC, Esteves SSR, Dinsdale RM, Guwy AJ. The effect of acid pre-treatment on the anaerobic digestion and dewatering of waste activated sludge. Bioresour Technol 2011;102:4076-4082.
- 6. Facchin V, Cavinato C, Fatone F, Pavan P, Cecchi F, Bolzonella D. Effect of trace element supplementation on the mesophilic anaerobic digestion of food waste in batch trials: the influence of inoculum origin. Biochem Eng J 2013;70:71-77.
- Forster-Carneiro T, Pérez M, Romero L. Influence of total solid and inoculum contents on performance of anaerobic reactors treating food waste. Bioresource Technol 2008;99(15):6994-7002.
- Gerardi MH The microbiology of anaerobic digesters. A John Wiley and Sons 2003;99-103.
- 9. Girma G, Edward J, Peter R. In vitro gas production provides effective method for assessing ruminant feeds. Calif Agric. 2004;58(1):54-58.
- Itodo IN, Lucas EB, Kucha EI. The Effect of Media Materials and its Quality on Biogas Yield. Niger. J Renew Energ. 1992;3(1):45-49.
- Sung JDS, Ki-Cheol E, Shihwu S, SangWon P, Hyunook K. Predicting Methane Production Potential of Anaerobic Co-digestion of Swine Manure and Food Waste. Environ Eng Res. 2008;13(2):93-97.
- 12. Kamthunzi WM. Anaerobic digestion of cattle manure in batch digesters at ambient temperature. Environ Sci Technol 2008;3:8-12.
- 13. Macias-Corral M, Samani Z, Hanson A, Smith G, Funk P, Yu H, et al. Anaerobic digestion of municipal solid waste and agricultural waste and the effect of co-digestion with dairy cow dung. Bioresoure Technol 2008;99:8288-8293.
- Montusiewicz A, Lebiocka M, Pawlowska M. Characterization of the biomethanization process in selected waste mixtures. Arch Environ Prot 2008;34(3):49-61.