

Journal of Food Processing & Technology

**Research Article** 

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# Effect of Different Levels of Sucrose-Glucose Mixture on Overall Quality of Guava Bar

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# Abstract

The effect of sucrose-glucose mixture was studied on the overall quality of guava bar stored at room temperature (25-30<sup>°</sup>C) during three months storage period. Different ratio of sucrose glucose mixture was used. All the treatments were analyzed for physicochemical characteristics and sensory (color, texture, taste and overall acceptability). The results showed that decreased were observed in water activity (from 0.68 to 0.62), moisture (from 18.59 to 14.43), pH (from 3.87 to 3.69) and ascorbic acid (from 3.87 to 3.69) color (from 7.67 to 5.63), texture (from 7.67 to 5.63), texture (from 7.42 to 5.37) and overall acceptability (from 7.53 to 5.48), while reducing sugar (from 14.16 to 14.41), titratable acidity (1.13 to 1.33), and total soluble solids (from 61.85 to 63.70) was increased. The overall results showed that treatment  $GL_2$  followed by  $GL_5$  were found adequate both physicochemical and sensory evaluation.

Keywords: Bar; Guava; Guar gum; Sucrose; Glucose

# Introduction

Guava (*Psidium guajava* L.) belongs to the family Myrtaceae, characterized by 80 genera and 3000 species [1,2]. It has originated from Mexico to Peru [3]. In terms of area and production, guava is the fourth most important fruit after mango, banana and citrus [4]. In Pakistan, guava fruit stands at number three in terms of production, after mango and banana [5]. A guava fruit has a mass of 150-250 grams.

Fruit leather is an intermediate moisture food (IMF), also called as fruit roll, fruit bar or fruit sheet commercially, and developed by dehydration of fruit pastes into leathery sheet [6]. Fruit leathers are probably, originated from the times of Persian Empire. They are recognized with different names in different nations; "Qamar al deen" in Lebanon, Syria and other Arab countries, "Bastegh" or "Pastegh" in Armenia, "Pestil" in Turkey and "Fruit roll" or "Fruit leather" in the United States. In scientific literature, the last name, "Fruit leather" is mostly used. Fruit leather having water activity less than 0. 6 and moisture content of 8-15%. They contain acids and sugar naturally, while humectants are purposely added to minimize water activity and to provide softness even at lower moisture levels [7]. Guar gum is consider as one of the main gums. Guar gum is achieved from a legume crop. It is a complex carbohydrate, broadly grown in Pakistan and India and has very low price. Xanthan and Guar gums are widely used for the thickness of food products all over the food industries [8].

## Materials and Methods

The research was conducted in the laboratory of the Department of Food Science and Technology, The University of Agriculture, Peshawar. Good quality fresh, mature and healthy guava was bought from the Peshawar local market and was transported to the laboratory. The diseased free fruit was selected and washed with water in order to remove dust, dirt and any other foreign material. The fruit was peeled, trimmed, cut and dipped in 1% citric acid to prevent oxidation. Then the fruit was blended in order to get the pulp. After that the treatments (20 °brix) were prepared. Each treatment were poured in stainless steel trays and kept in cabinet drier at 55°C. The treatments were GL<sub>0</sub> = Control, GL<sub>1</sub> = Guava pulp with sucrose + glucose (1:1) and guar gum (0.25%), GL<sub>2</sub> = Guava pulp with sucrose + glucose (3:7) and guar gum

(0.25%), GL<sub>4</sub> = Guava pulp with sucrose + glucose (10:0) and guar gum (0.25%), GL<sub>5</sub> = Guava pulp with sucrose + glucose (0:10) and guar gum (0.25%).

#### Packaging

The prepared bar was wrapped with aluminum foil and then packed in a transparent polythene bags.

#### Chemical analysis

Ascorbic acid: Ascorbic acid determination was done by the standard method as detailed in the AOAC [9] method no 967.21.

**Titratable acidity:** The titratable acidity was measured by the standard method of AOAC [9] method no 942.15.

**pH:** pH was measured by using the standard method of AOAC [9] method no 2005.02.

**Total soluble solids:** The total soluble solids (TSS) were determined by the standard methods of AOAC [9] method no 932.14 and 932.12, using Atago digital refractometer at room temperature.

**Reducing sugar:** Reducing sugars was determined by Lane and Eynon recommended procedure as explained in AOAC [9] method no 920.183.

**Water activity (a**<sub>w</sub>): Water activity was measured by using Novasina RTD 502 apparatus (Novasina, Pfapfikkon, Switzerland).

**Moisture (%):** Moisture of the sample was determined by the standard method of AOAC [9] method no 925.45.

Total microbial count: The sample was analyzed for the total

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Received April 17, 2015; Accepted May 25, 2015; Published June 01, 2015

Citation: Shakoor A, Ayub M, Wahab S, Khan M, Khan A, et al. (2015) Effect of Different Levels of Sucrose-Glucose Mixture on Overall Quality of Guava Bar. J Food Process Technol 6: 469. doi:10.4172/2157-7110.1000469

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microbial count by the total plate count method as describe Dillello.

**Sensory evaluation:** The guava bar was sensory judged for taste, color, overall acceptability and texture by the panels of 10 judges. The evaluation was carried out by using 9 points hedonic scale of Larmond [10].

#### Statistical analysis

All the data concerning treatments and storage interval were statistically analyzed by means of completely Randomized Design (CRD) 2 Factorial and the means were separated by applying least significant difference (LSD) Test at 5% possibility level as defined by Steel and Torrie [11].

## **Results and Discussion**

#### Chemical analysis

Water activity (a\_): Mean values for a\_ reduced from 0.68 to 0.62 for the period of the storage. Highest mean values for treatments were perceived in  $\mathrm{GL}_{_2}$  and  $\mathrm{GL}_{_5}$  (0.66) followed by  $\mathrm{GL}_{_3}$  and  $\mathrm{GL}_{_4}$  (0.65), in compare lowest mean values were documented in  $GL_0$  and  $GL_1$  (0. 64). During the storage highest fall in aw was recorded in GL<sub>0</sub> (10.45%) followed by GL, (8.96), in contrast minimum fall was recorded in GL<sub>3</sub> (4.48) followed by  $GL_{\epsilon}$  (7.35) (Table 1). The association of the added sugars and water through hydrogen bonding reduction was occurred in a... Invert sugar acts as bonding agent and the water passage at the product surface is slow. This action delays the creation of sugar recrystallization on the product surface for the duration of storage [12]. Low water activity value provides a margin of safety for the storage of acid foods at ambient temperatures, because it would not only prevent growth of pathogenic microorganism but also would strongly inhibit growth of non-pathogenic fungi and yeasts as well [13]. In a similar study, Babalola et al. [14] found a decrease in a during study of guava and pawpaw leather (from 0.64- 0.61), Huang and Hung [15] also reported a decrease in  $a_w$  during study of pear fruit leather(from 0.44-0.37) and Irwandi et al. [16] found a decrease in  $a_w$  during study of durian fruit leather from (0.597-0.573) respectively.

**Moisture (%):** The mean values for moisture decrease from 18.59 to 14.43 during storage. Highest mean values for treatments were observed in  $GL_0$  (18.44) followed by  $GL_1$  (16.46), in contrast lowest mean values were recorded in  $GL_2$  (15.59) followed by  $GL_5$  (15.80) and  $GL_4$  (16.33). During storage highest fall in moisture content was recorded in  $GL_2$  (30.53%) followed by  $GL_5$  (28.45%), in compare minimum fall was observed in  $GL_0$  (0.76%) followed by  $GL_1$  (21.50%) (Table 2). There was a strong relationship between moisture content and  $a_w$  the higher the moisture content the higher the  $a_w$ . Similar observations were made on kiwifruit leather by Lodge [17] and jackfruit leather by Che Man and Taufik [18]. In a similar study, Huang and Hung [15] found a decrease in moisture during study of pear fruit leather (12.13-7.97) and Irwandi et al. [16] also observed a decrease in moisture during study of durian fruit leather (15.82-14.36) respectively.

Microbial count: The mean values for microbial count reduced from  $13.33 \times 10^1$  to  $5.17 \times 10^1$  cfu/g for the period of storage. Highest mean values for treatments were perceived in GL<sub>1</sub> (10.29×10<sup>1</sup>) cfu/g followed by  $GL_4$  (9.43×10<sup>1</sup>), in contrast lowest mean values were recorded in GL<sub>5</sub> (7.71×10<sup>1</sup>) cfu/g followed by GL<sub>2</sub> (8×10<sup>1</sup>) cfu/g. During storage highest fall in microbial count was recorded in GL, (76.29×10<sup>1</sup> %) followed by  $GL_5$  (69.23×10<sup>1</sup>%), in compare minimum fall was observed in  $GL_0$  (50×10<sup>2</sup>) and  $GL_1$  (50×10<sup>1</sup>%) followed by  $GL_4$  (57.14×10<sup>1</sup>%) (Table 3). According to Troller [19,20], most of the microorganisms can barely survive a lower than 0.60. Similar result of microbial count was reported by Huang and Hung [15] the results of microbiological examine stated in forgoing studies [19]. Decrease of microorganism might be due to low water activity, low pH and low moisture content minimum water activity required for microbial growth [20], and its pH (3.8) was below the lower limit for bacterial growth (4.0), allowing only moulds and yeasts to grow [21]. Also, different preservation factors,

Treatments		Storage Interval (Days)										
Treatments	0	15	30	45	60	75	90	% Decrease	mean			
GL₀	0.66	0.66	0.65	0.63	0.62	0.61	0.60	10.45	0.63 e			
GL <sub>1</sub>	0.68	0.67	0.66	0.64	0.63	0.62	0.61	8.96	0.64d			
GL <sub>2</sub>	0.69	0.68	0.67	0.66	0.65	0.64	0.63	8.70	0.66 a			
GL <sub>3</sub>	0.67	0.66	0.66	0.65	0.65	0.64	0.64	4.48	0.65bc			
GL₄	0.68	0.67	0.66	0.65	0.64	0.63	0.62	8.82	0.65 c			
GL₅	0.68	0.67	0.67	0.66	0.65	0.64	0.63	7.35	0.66 ab			
Mean	0.68 a	0.67 b	0.66 c	0.65 d	0.64 e	0.63 f	0.62 g					

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.604

LSD at 5% level for intervals = 0.053

 Table 1: Effect of storage period and treatments on water activity (a<sub>w</sub>) of guava bar.

Treatment		Storage Interval										
rreatment	0	15	30	45	60	75	90	%Decrease	Wiedli			
GL <sub>0</sub>	18.53	18.50	18.47	18.44	18.41	18.39	18.39	0.76	18.45a			
GL <sub>1</sub>	18.97	18.45	17.59	16.47	15.21	14.54	13.99	26.26	16.46b			
GL <sub>2</sub>	18.30	17.59	16.76	15.39	14.61	13.81	12.71	30.53	15.59c			
GL <sub>3</sub>	18.90	18.00	17.46	16.71	15.51	14.41	13.84	26.79	16.40bc			
GL <sub>4</sub>	18.45	17.50	16.56	16.01	15.96	15.37	14.48	21.50	16.33bc			
GL₅	18.39	17.05	16.43	15.83	15.17	14.57	13.16	28.45	15.80bc			
Mean	18.59a	17.85ab	17.21bc	16.47cd	15.81de	15.18ef	14.43f					

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.8495

LSD at 5% level for intervals = 0.9176

 Table 2: Effect of storage period and treatments on (%) moisture of guava bar.

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Treat	Storage Interval (Days)									
Treat	0	15	30	45	60	75	90	% Decrease	wean	
$GL_0$	12×10 <sup>2</sup>	11×10 <sup>2</sup>	10×10 <sup>2</sup>	9×10 <sup>2</sup>	8×10 <sup>2</sup>	7×10 <sup>2</sup>	6×10 <sup>2</sup>	50.00×10 <sup>2</sup>	9.00×10 <sup>2</sup> b	
GL <sub>1</sub>	14×10 <sup>1</sup>	12×101	9×101	8×10 <sup>1</sup>	7×10 <sup>1</sup>	6×10 <sup>1</sup>	5×101	50.00×101	10.29×10¹ a	
$GL_2$	13×10 <sup>1</sup>	12×101	10×101	7×10 <sup>1</sup>	6×10 <sup>1</sup>	5×101	3×101	76.92×101	8.00×101cd	
$GL_3$	13×10 <sup>1</sup>	10×101	9×101	7×10 <sup>1</sup>	6×10 <sup>1</sup>	5×101	4×101	64.29×101	8.71×10 <sup>1</sup> bc	
$\operatorname{GL}_4$	14×10 <sup>1</sup>	12×10 <sup>1</sup>	10×10 <sup>1</sup>	9×10 <sup>1</sup>	8×10 <sup>1</sup>	7×10 <sup>1</sup>	6×101	57.14×10 <sup>1</sup>	9.43×101 b	
$GL_{5}$	15×101	13×101	11×101	10×101	9×10 <sup>1</sup>	8×10 <sup>1</sup>	7×101	69.23×101	7.71×10¹d	
Mean	13.33×10¹a	11.67×10¹ a	9.83×10¹a	8.33×10¹a	7.33×10¹a	6.33×10¹a	5.17×10¹a			

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.7239

LSD at 5% level for intervals = 0.7818

Table 3: Effect of storage period and treatments on Microbial load (cfu/g) of guava bar.

Treatments					Storage In	terval (Days)			Mean
Treatments	0	15	30	45	60	75	90	% Increase	
GL <sub>0</sub>	3.26	3.95	3.95	3.95	3.96	3.96	3.97	17.88	3.86f
GL <sub>1</sub>	14.57	14.57	14.57	14.58	14.58	14.58	14.58	0.07	14.58e
GL <sub>2</sub>	17.77	17.79	17.82	17.86	17.89	17.91	17.93	0.89	17.85a
$GL_3$	17.34	17.34	17.35	10.35	17.35	17.36	17.36	0.12	17.35b
GL <sub>4</sub>	15.99	16.02	16.06	16.09	16.13	16.15	16.17	1.11	16.09d
GL₅	16.00	16.42	16.42	16.43	16.43	16.43	16.44	2.68	16.37c
Mean	14.16b	14.35a	14.36a	14.38a	14.39a	14.40a	14.41a		

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.1174

LSD at 5% level for intervals = 0.1268

Table 4: Effect of storage period and treatments on (%) reducing sugar of guava bar.

Treatments		Storage Interval (Days)									
Treatments	0	15	30	45	60	75	90	% Increase	Wear		
GL	14.00	14.40	14.70	15.00	15.20	15.40	15.70	13.93	14.91 f		
GL	77.10	77.40	77.70	78.00	78.40	78.70	79.00	20.00	78.04 e		
GL <sub>2</sub>	78.50	78.90	79.20	79.50	79.80	80.10	80.40	14.53	79.49 a		
GL <sub>3</sub>	77.80	78.10	78.40	78.80	79.10	79.40	79.70	15.61	78.76 c		
GL₄	77.40	77.80	78.10	78.50	78.80	79.20	79.50	14.89	78.47 d		
GL5	78.20	78.50	78.90	79.30	79.60	79.80	80.00	16.67	79.19 b		
Mean	61.85 q	62.20 f	62.50 e	62.83 d	63.13 c	63.40 b	63.70 a				

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.5031

LSD at 5% level for intervals = 0.5434

Table 5: Effect of storage period and treatments on TSS (°brix) of guava bar

such as pH and water activity, usually have not just an additive effect on food stability, but act synergistically to inhibit microbial growth [22].

Reducing sugar (%): The mean values for reducing sugar increased from 14.16 to 14.41 for the period of the storage. Highest mean values for treatments were perceived in GL<sub>2</sub> (17.85) followed by GL<sub>3</sub> (17.35), while the lowest mean values were recorded in  $GL_0$  (3.86) followed by  $\mathrm{GL}_{\scriptscriptstyle 1}$  (14.58). During storage the highest raise in reducing sugar was recorded in GL<sub>0</sub> (17.73%) followed by GL<sub>5</sub> (2.68%), while lowest raise was observed in GL<sub>1</sub> (0.07%) followed by GL<sub>3</sub> (0.12%) as shown in Table 4. Due to the transposition of non-reducing sugars into reducing sugars and the modification of polysaccharides to monosaccharide's the reducing sugar is increase. In a similar result, Sharma et al. [23] reported an increase in reducing sugar (from 43.1-49.8) and Phimpharian et al. [24] also found an increase in reducing sugar from (20.9 to 26.3) respectively. The increase in reducing sugars has also been observed during storage of mango leather by Rao and Roy [25]. Similar results have been record in sapota -papaya bar during 3 months of storage period [26] and in apricot - soy toffees [27].

**Total soluble solids:** The mean values for Total soluble solids increased from 61.85 to 63.70 for the period of storage. Extreme mean values for treatments were perceived in  $GL_2$  (79.49) followed by  $GL_5$  (79.19), but in contrast the deepest mean values were registered in  $GL_0$  (14.91) followed by  $GL_1$  (78.04). During storage the highest raise in TSS was recorded in  $GL_1$  (20%) followed by  $GL_5$  (16.67%), while lowest raise was observed in  $GL_0$  (13.93%) followed by  $GL_2$  (14.53%) (Table 5). The increase in TSS might be due to the renovation of starch and other insoluble carbohydrates into sugars and also due to the loss of moisture content that tends to increase total soluble solid. In a similar result, Phimpharian et al. [24] reported an increase in TSS (from 82.42-86.9).

**Titratable acidity:** Acidity is the quantity of shelf life of the product. Titrable acidity studied to confirm physico-chemical changes during preparation [28] and during storage [29]. The mean values for Titratable acidity increased from 1.13 to 1.33 for the period of storage. Best mean values for treatments were perceived in GL<sub>2</sub> (1.45) followed by GL<sub>5</sub> (1.42), but in contrast the lowest mean values were listed in GL<sub>0</sub>

(1. 30) followed by  $GL_1$  (1. 36). For the period of storage the highest raise in acidity was recorded in  $GL_2$  (15.38%) followed by  $GL_4$  (14.77%), while deepest raise was perceived in  $GL_0$  (19.47%) followed by  $GL_5$  (12.50%) (Table 6). The increase in acidity might be due to development of acidic substances by the degradation of pectic bodies or breakdown and also attributed to hydrolysis of polysaccharides and non-reducing sugars through utilization of acids for converting them to hexose sugar. Rao and Roy [25] found an increase in acidity during storage of mango sheet (from 0.3-0.75). Manu et al. [30] noticed increase in acidity during storage of mango [31] observed an increase in acidity during study of guava leather (from 0.42-0.48) respectively. Acidiy of guava fruit bar increased while pH decreased during storage as per the study result of Gowda et al.

**pH:** The mean values for pH reduced from 3.87 to 3.69 for the period of storage. Highest mean values for treatments were perceived in GL<sub>2</sub> (3.87) followed by GL<sub>5</sub> (3.79) and GL3 (3.78), in contrast lowest mean values were noted in GL<sub>0</sub> (3. 75) followed by GL<sub>1</sub> (3.76) and GL<sub>4</sub> (3.77). During storage highest fall in pH was recorded in GL<sub>3</sub> (5.40%)

followed by  $GL_5$  (5.04%), in compare minimum fall was observed in  $GL_2$  (4.13%) followed by  $GL_4$  (4.29%) (Table 7). The changes in pH values might be due to increase in acidity and also due to the other chemical that occur during storage interval. Phimpharian et al. [24] noticed a reduction in pH values during storage of pineapple leather (from 3.6-3.8). Azeredo et al. [32] observed a decrease in pH values during storage of mango leathers (from 3.8-3.5). Similarly Natalia et al. [33] also observed a decrease in pH values during study of apple leather (from 3.50-3.30) respectively.

Ascorbic acid (Vit. C): Fruits and vegetables are important sources of ascorbic acid. The ascorbic acid content decreased during storage due to oxidation of ascorbic acid to dehyro ascorbic acid. Hence, vitamin C assessment was found out during the storage period. The mean values for ascorbic acid decreased from 92.34 to 74.42 for the period of storage. Supreme mean values for treatments were observed in GL<sub>2</sub> (84.78) followed by GL<sub>5</sub> (83.77), but in difference the deepest mean values were registered in GL<sub>0</sub> (78.60) followed by GL<sub>1</sub> (82.50). During storage the highest raise in ascorbic acid was recorded in GL<sub>0</sub> (21.47%) followed

Treatments		Storage Interval (Days)										
Treatments	Initial	15	30	45	60	75	90	% Increase	Mean			
GL₀	1.22	1.24	1.27	1.30	1.33	1.36	1.39	12.23	1.30 f			
GL <sub>1</sub>	1.24	1.29	1.33	1.36	1.39	1.43	1.46	15.07	1.36 e			
GL <sub>2</sub>	1.36	1.39	1.42	1.45	1.48	1.51	1.54	11.69	1.45 a			
GL <sub>3</sub>	1.32	1.36	1.38	1.41	1.44	1.47	1.50	12.00	1.41 c			
GL <sub>4</sub>	1.27	1.30	1.34	1.38	1.41	1.45	1.49	14.77	1.38 d			
GL₅	1.33	1.35	1.39	1.42	1.46	1.49	1.52	12.50	1.42 b			
Mean	1.13g	1.17f	1.21e	1.24d	1.27 c	1.30 b	1.33 a					

Values having different alphabetical letters are significantly (P<0.05) not same LSD at 5% level for treatments = 0.206

LSD at 5% level for intervals = 0.20LSD at 5% level for intervals = 0.943

Table 6: Effect of storage period and treatments on (%) Titratable acidity of guava bar.

Treetmente					Storage Inte	erval (Days)			Mean
Treatments	0	15	30	45	60	75	90	% Decrease	
GL₀	3.84	3.81	3.78	3.75	3.73	3.70	3.67	4.43	3.75 d
GL <sub>1</sub>	3.86	3.82	3.79	3.77	3.74	3.70	3.67	5.03	3.76 c
GL <sub>2</sub>	3.87	3.85	3.83	3.80	3.77	3.74	3.71	4.13	3.80 a
GL <sub>3</sub>	3.89	3.84	3.82	3.78	3.75	3.72	3.68	5.40	3.78 b
GL₄	3.85	3.83	3.80	3.77	3.75	3.71	3.69	4.29	3.77 c
GL₅	3.89	3.86	3.82	3.79	3.76	3.72	3.69	5.04	3.79 ab
Mean	3.87 a	3.84 b	3.81 c	3.78 d	3.76 e	3.72 f	3.69 g		

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.242

LSD at 5% level for intervals = 0.902

Transformente					Storage Inte	rval (Days)			Moan
Treatments	0	15	30	45	60	75	90	% Decrease	wean
GL <sub>0</sub>	90.40	85.00	79.50	77.00	75.30	72.00	71.00	21.47	78.60 d
GL <sub>1</sub>	92.00	89.00	85.50	82.50	78.50	76.50	73.50	20.11	82.50 c
GL <sub>2</sub>	92.33	90.50	87.50	85.00	81.33	80.00	76.79	16.83	84.78 a
GL <sub>3</sub>	93.50	88.00	85.83	81.83	79.82	77.23	75.83	18.90	83.15bc
GL <sub>4</sub>	92.67	87.83	84.83	81.57	79.57	77.56	74.56	19.54	82.66 c
GL₅	93.17	89.99	85.79	82.82	80.87	78.87	74.87	19.64	83.77 b
Mean	92.34 a	88.39 b	84.82 c	81.79 d	79.23 e	77.03 f	74.42 g		

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.9600

LSD at 5% level for intervals = 1.0369

Table 8: Effect of storage period and treatments on Ascorbic acid (mg/100g) of guava bar.

by GL, (20.11%), while lowest raise was observed in GL, (16.83%) followed by GL, (18.93%) (Table 8). Temperature has a major effect on the rate of loss of ascorbic acid. Losses of ascorbic acid were increased with the increase in temperature [34]. The ascorbic acid content decreased during storage [35]. Loss of ascorbic acid might be due to its oxidation to dehydro ascorbic acid followed by more degradation to 2, 3 - diketogulonic acid and finally to furfural complexes which go in browning reactions Sharma et al. [23]. Sharma et al. [23] noticed loss of ascorbic acid during study of apricot fruit (from 9.5-8.6). Jain and Nema [31] noticed loss of ascorbic acid during study of guava leather (176.27-104.87mg/g) and Ayshaye et al. [36] found a decrease in ascorbic acid during storage of pawpaw (from 83.33-74.70 and guava leather (260.0-237.0) respectively. Loss of ascorbic acid has earlier been reported in mango leather during of 3 months storage by Rao and Roy [25]. Similar results have been reported by Sreemathi et al. [26] in sapota -papaya bar during 3 months of storage.

## Sensory evaluation

**Color:** Initially the mean score of judges for color of guava bar of  $GL_0$  to  $GL_5$  was 6, 8, 8, 8, 8 and 8 which was gradually decrease to 3.5, 6.1, 6.5, 5.9, 5.8 and 6. 0 similarly for the period of storage. The mean values for intervals were significantly (P<0.05) intensified from 7.67 to 5. 63 for the period of storage. Supreme mean values for treatments were perceived in  $GL_2$  (7.2571) 5.8 and 6. 0 similarly for the period of storage. The mean values for treatments were perceived in  $GL_2$  (7.2571) 5.8 and 6. 0 similarly (P<0.05) intensified from 7.67 to 5. 63 for the period of storage. Supreme mean values for treatments were perceived in  $GL_2$  (7.2571) followed by  $GL_1$  (7. 0857), but in contrast the lowest mean values were listed in  $GL_0$  (4.7000) followed by  $GL_4$  (6. 9143) [37-39]. For the period of storage the highest fall in color was recorded in  $GL_0$  (41.67%) followed by  $GL_1$  (27.50%), while lowest fall was observed in  $GL_2$  (18.75%) followed by  $GL_5$  (23.75%) (Table 9). A decrease in color might be due to browning

reaction (millard) that occure during heating process in the drier. In a similar study, Jain and Nema [31] reported loss of color during storage of guava leather (7.10-6.16). Naz [39] also observed a decrease in color during her study (from 6-5) and Babalola et al. [14] (from 6.8-5.2) respectively.

Texture: Originally the mean score of juries for texture of guava bar of GL<sub>0</sub> to GL<sub>5</sub> was 5, 8, 8, 8, 8 and 8 which was progressively reduced to 3.1, 5. 6, 6.5, 5.9, 5. 7 and 6 correspondingly for the period of storage. The mean values for intervals were significantly (P<0.05) intensified from 7.67 to 5. 63 for the period of storage. Maximum mean values for treatments were perceived in GL<sub>2</sub> (7.2286) followed by GL<sub>1</sub> (7. 0286), but in contrast the deepest mean values were listed in GL<sub>0</sub> (3.9714) followed by  $GL_4$  (6.7000). During storage the highest fall in texture was recorded in GL<sub>0</sub> (38.00%) followed by GL<sub>1</sub> (30.00%), while lowest fall was observed in  $GL_2$  (18.75%) followed by  $GL_5$  (25.00%) (Table 10). The texture of fruit leathers is mostly affected by their moisture content and drying temperatures by Che-man et al. [39]. High temperatures and long drying times are related with lower moisture content and rigid texture. Differences in texture of leathers might also be due to variations in genetic makeup of the fruit, rate of water immersion from the surroundings and protein content of the fruit amongst others by Babalola et al. [39]. The texture of fruit leather is also affected by the addition of sugar, which is occasionally completed in order to recover the flavor of the leather by Jain and Nema [31]. Similar result of texture was reported by Naz [37] (from 7-6).

**Taste:** Initially the mean score of juries for taste of guava bar of  $GL_{0}$  to  $GL_{5}$  was 4.5, 8, 8, 8 and 8which was gradually decrease to 2.50, 5.60, 6.30, 5.90, 5.70 and 6.20 similarly for the period of storage. The mean values for intervals were significantly (P<0.05) intensified from 7.42 to 5. 37 for the period of storage. Supreme mean values for treatments were perceived in  $GL_{2}$  and  $GL_{5}$  (7.1143) followed by  $GL_{1}$  (6.9429), but

Treatments				Storage Int	erval (Days)				Mean
Treatments	0	15	30	45	60	75	90	% Decrease	Mean
GL <sub>0</sub>	6	5.6	5.1	4.7	4.2	3.8	3.5	41.67	4.70 d
GL <sub>1</sub>	8	7.6	7.2	6.9	6.6	6.3	5.8	27.50	6.91 c
GL <sub>2</sub>	8	7.8	7.5	7.2	7	6.8	6.5	18.75	7.26 a
GL <sub>3</sub>	8	7.7	7.4	7.1	6.7	6.2	6	25.00	7.01bc
GL <sub>4</sub>	8	7.6	7.3	7	6.8	6.4	5.9	26.25	7.00bc
GL₅	8	7.7	7.4	7.1	6.8	6.5	6.1	23.75	7.09 b
Mean	7.67 a	7.33 b	6.98 c	6.67 d	6.35 e	6.00 f	5.63 g		

Values having different alphabetical letters are significantly (P<0.05) not same LSD at 5% level for treatments = 0.1518

LSD at 5% level for intervals = 0.1640

Table 9: Effect of storage period and treatments on color of guava bar.

Treatments		Storage Interval (Days)										
Treatments	0	15	30	45	60	75	90	% Decrease	wean			
GL₀	5	4.6	4.1	3.9	3.7	3.4	3.1	38.00	3.97 d			
GL <sub>1</sub>	8	7.5	7.1	6.6	6.2	5.9	5.6	30.00	6.70 c			
GL <sub>2</sub>	8	7.7	7.5	7.2	7	6.7	6.5	18.75	7.23 a			
GL <sub>3</sub>	8	7.7	7.3	7	6.7	6.3	5.9	26.25	6.99 b			
GL₄	8	7.6	7.1	6.7	6.3	6	5.7	28.75	6.77 c			
$GL_{5}$	8	7.6	7.3	7.1	6.8	6.4	6	25.00	7.03 b			
Mean	7.40 a	7.02 b	6.62 c	6.28 d	5.98 e	5.66 f	5.36 g					

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.1519

LSD at 5% level for intervals = 0.1641

 Table 10: Effect of storage period and treatments on texture of guava bar.

Citation: Shakoor A, Ayub M, Wahab S, Khan M, Khan A, et al. (2015) Effect of Different Levels of Sucrose-Glucose Mixture on Overall Quality of Guava Bar. J Food Process Technol 6: 469. doi:10.4172/2157-7110.1000469

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Treatments	Storage Interval (Days)										
noutinonto	0	15	30	45	60	75	90	% Decrease	_		
GL <sub>0</sub>	4.50	4.10	3.70	3.50	3.20	2.90	2.50	44.44	3.49 d		
GL <sub>1</sub>	8.00	7.40	7.10	6.70	6.30	5.90	5.60	30.00	6.71 c		
GL <sub>2</sub>	8.00	7.70	7.40	7.10	6.80	6.50	6.30	21.25	7.11 a		
GL <sub>3</sub>	8.00	7.60	7.20	6.90	6.70	6.30	5.90	26.25	6.94 b		
GL₄	8.00	7.50	7.10	6.80	6.40	6.00	5.70	28.75	6.79 c		
GL₅	8.00	7.70	7.40	7.10	6.80	6.60	6.20	22.50	7.11 a		
Mean	7.42 a	7.00 b	6.65 c	6.35 d	6.03 e	5.70 f	5.37 g				

Values having different alphabetical letters are significantly (P<0.05) not same

LSD at 5% level for treatments = 0.1134

LSD at 5% level for intervals = 0.1225

Table 11: Effect of storage period and treatments on taste of guava bar.

Treatments	Storage Interval (Days)								Maan
	0	15	30	45	60	75	90	% Decrease	wear
GL <sub>0</sub>	5.2	4.8	4.3	4	3.7	3.4	3	42.31	4.06 d
GL <sub>1</sub>	8	7.5	7.1	6.7	6.4	6	5.7	28.75	6.77 c
GL <sub>2</sub>	8	7.7	7.5	7.2	6.9	6.7	6.4	20.00	7.20 a
GL <sub>3</sub>	8	7.6	7.3	7	6.8	6.4	6	25.00	7.01 b
GL <sub>4</sub>	8	7.6	7.2	6.8	6.5	6.1	5.8	27.50	6.86 c
GL₅	8	7.7	7.4	7.1	6.7	6.4	6	25.00	7.04 b
Mean	7.53 a	7.15 b	6.80 c	6.47 d	6.17 e	5.83 f	5.48 g		

Values having different alphabetical letters are significantly (P<0.05) not same LSD at 5% level for treatments = 0.1118

LSD at 5% level for intervals = 0.1208

Table 12: Effect of storage period and treatments on overall acceptability of guava bar.

in contrast the deepest mean values were registered in  $GL_0$  (3.4857) followed by  $GL_4$  (6.7143). During storage the highest fall in taste was recorded in  $GL_0$  (44.44%) followed by  $GL_1$  (30.00%), while lowest fall was observed in  $GL_2$  (21.25%) followed by  $GL_5$  (22.50%) (Table 11). Taste and smell perceptions noted when food is taken. The overall flavor impression is the result of taste perceived by the taste buds in the mouth and the aromatic conpounds detected by the epithelium in the olfactory organ in the nose. Jain and Nema [31] recorded a decrease in taste during study of guava leather (from 6.19-6.02), Okilya et al. [38] also found a decrease in taste (from 6.63-4.33) respectively.

**Overall acceptability:** Primarily the mean score of juries for overall acceptability of guava bar of  $GL_0$  to  $GL_5$  was 5.2, 8, 8, 8 and 8which was gradually decrease to 3, 5.7, 6.4, 6, 5.8 and 6 similarly for the period of storage. The mean values for intervals were significantly (P<0.05) intensified from 7.53 to 5. 48 for the period of storage. Supreme mean values for treatments were perceived in  $GL_2$  (7.20) followed by  $GL_5$  (7.04), but in contrast the deepest mean values were listed in  $GL_0$  (4.06) followed by  $GL_1$  (6.77). During storage the highest fall in overall acceptability was recorded in  $GL_2$  (20.00%) followed by  $GL_5$  and  $GL_3$  (25.00%) (Table 12). Overall acceptability generally related to all sensory attributes. It is stated that the suitability of fruits and vegetables is influenced by their aroma by Karmas and Harris [39]. Sharma et al. [23] noticed a decrease in overall acceptability during storage of apricot fruit bar (from7.8-7.2) respectively.

## **Conclusion and Recommendations**

In present study, guava bar was prepared by using different level of sucrose glucose mixture with guar gum. The samples were analyzed for physicochemical, microbiologically and sensory. From this study, physic chemically the samples  $GL_2$  prepared by sucrose: glucose (7:3)

followed by  $GL_5$  prepared by sucrose (10: 0) showed best result, while  $GL_0$  prepared by guava pulp and followed by  $GL_1$  prepared by sucrose: glucose (50: 50) showed lowest result. Sensory and microbiologically  $GL_2$  followed by  $GL_5$  showed good result, while  $GL_0$  followed by  $GL_1$  showed lowest result [40].

## Recommendations

1. Study should be carried out in the effect of different packaging materials with different temperature.

2. Study the effect of different drying methods with different temperature.

3. Further study should be carried out that maintains color clarity.

#### References

- Nakasone HY, Paull RE (1998) Tropical fruits wallirgford, CAB Queensland Agri J 3: 93-98.
- Pervaiz U, Khan A, Javed R, Zeb J (2008) Production constraints of guava in district Kohat. Sarhad. J Agri 24: 549-554.
- Zamir R, Ali N, Shah ST, Muhammad T, Shah SA (2007) In vitro re-generation of guava (*psidiumguajava* L.) from shoot tips of mature trees. Pak J Bot 39: 2395-2398.
- Kadam DM, Kaushik P, Kumar R (2012) Evaluation of guava products quality. Int. J Food Sci Nut Engg 2: 7-11.
- Sindh Board of Investment (2010) Prefeasibility study guava pulping and squash making unit. Govt. Sindh.
- Jaswir T, Yaakobb, Cheman, Yusof S, Selamat J, et al. (1998) Effect of glucose syrup solid, sucrose, sensory acceptability of durian leather hydrogenated palm oil and soy-lecithin on sensory acceptability of durian leather. J Food Proc Pres 22: 13-25.
- 7. Torley PJ, De Boer J, Bhandari BR, Halley PJ (2005) Composition rheology

property relationships of commercial fruit leathers.4  $^{\mbox{\tiny th}}$  Int. symposium on food Rheology and structure.

- Murtaza MA, Uddin GM, Huma N, Shabbir MA, Mahmood S (2004) Quality evaluation of Ice Cream Prepared with Different Stabilizers/Emulsifier Blends. Int J Agri Biol.
- 9. AOAC (2012) Official Methods of Analysis of AOAC. International, 19th edition, Association of Official Analytical Chemists, USA.
- 10. Larmond E (1977) Lab Method of sensory evaluation of food. Canada, Deptt. Agric. Ottawa.
- Steel R, Torrie J, Dickey D (1996) Principles and Procedures of Statistics. A Biometrical Approach, 3<sup>rd</sup> Ed., McGraw Hill book Co., USA.
- Duangmal K, Khachonsakmetee S (2009) Osmotic dehydration of guava: influence of replacing sodium metabisulphite with honey on quality. Int J Food Sci Tech 44: 1887-1894.
- 13. Roos YH (1995) Phase transitions in food. San Diego: Academic Press, Londres, UK.
- Babalola SO, Ashaye OA, Babalola AO, Aina JO (2002) Effect of cold temperature storage on the quality attributes of pawpaw and guava leathers. Afr J Biotech 1: 61-63.
- 15. Huang X, Hsieh FH (2005) Physical properties, sensory attributes, and consumer preference of pear fruit leather. J Food Sci 70: e177-e186.
- Irwandi J, Che Man YB, Yusof S, Jinap S, Sugisawa H (1998) Effects of Type of packaging materials on physicochemical, microbiological and sensory characteristics of durian fruit leather during storage. J Sci Food Agri 76: 427-434.
- 17. Lodge N (1981) Kiwi fruit: Two novel processed products. Food Tech, New Zealand.
- Man YBC, Taufik (1995) Development and stability of Jackfruit leather. Tropical Sci 35: 245-250.
- Irwandi J, Che Man YB (1996) Durian leather: development, properties and storage stability. J Food Quality 19: 479-489.
- Northolt MD (1979) The Influence of water activity on microorganisms in foods. Food Tech 45: 159.
- 21. Alzamora SM (1994) Fundamentos del methodo de conservacion porfactores combinados. Valencia: Universidade Politecnica deValencia.
- 22. Leistner L (1992) Food preservation by combined methods. Food Res. Inte 25: 151-158.
- Sharma SK, Chaudhary SP, Rao VK, Yadav VK, Bisht TS (2013) Standardization of technology for preparation and storage of wild apricot fruit bar. J Food Sci Tech 50: 784-790.
- 24. Phimpharian C, Jangchud A, Jangchud K, Therdthai N, Prinyawiwatkul W

(2011) Physicochemical characteristics and sensory optimisation of pineapple leather snack as affected by glucose syrup and pectin concentrations. Int J Food Sci Tech 46: 972-981.

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- Rao VS, Roy SK (1980) Studies on dehydration of mango pulp. II: storage studies of the mango sheet/leather. Indian Food Packer 34: 72-79.
- 26. Sreemathi M, Sankaranarayanan R, Balasubramanyan S (2008) Sapota papaya bar. Madras. J Agri 95: 170-173.
- Thakur NS, Suman M, Kaushal BBL, Sharma M (2007) Apricot soya toffees a protein enriched product. Indian Food Packer 61: 77-81.
- Sandhu KS, Chander R, Bajwa U, Mahajan BVC (2008) Effect of papaya ripening incorporation of sucrose liquid glucose and citric acid on papaya leather quality formulations. J food Sci Tech 45: 133-138.
- Datey SP, Raut VU (2009) Physico-chemical changes in mango pulp during ambient storage in glass containers. J Food Sci Technol 2: 713-714.
- Manu ML, Oduro, Addo (2013) Effect of dextrin zed sweet potatoes on the physicochemical and sensory quality of infra-red dried mango leather. J Food Proc Tech 4: 5.
- 31. Jain PK, Nema PK (2007) Processing of Pulp of Various Cultivars of Guava (*Psidium guajava* L.) for Leather Production. Agri Engg Int 9: 1-9.
- Azeredo HMC, Brito ES, Moreira GEG, Farias VL, Bruno LM (2006) Effect of drying and storage time on the physicochemical properties of mango leathers. Int J Food Sci Tech 41: 635-638.
- 33. Natalia A, Ruiz Q, Demarchi SM, Giner SA (2011) Research on dehydrated fruit leathers: A Review.
- Johnson M, Hessel M (1982) Stability of Ascorbic acid in ready to drink juices. Varfoda 34: 267-279.
- Vidhya R, Narain A (2010) Development of preserved products using under exploited fruit, wood apple "*Limonia acidissima*". African J Food Sci Tech 1: 51-57.
- Ashaye OA, Babalola, Babalola AO, Aina JO, Fasoyiro SB (2005) Chemical and organoleptic characterization of pawpaw and guava leathers. World J Agric Sci 1: 50-51.
- Cheman YB, Jaswir I, Yusof S, Selamat J, Sugisawa H (1997) Effect of different dryers and drying conditions on acceptability and physicochemical characteristics of durian leather. J Food Proc Pres 21: 425-441.
- Okilya S, Muskia IM, Kaaya AN (2009) Effect of solar drying on the quality and acceptability of jackfruit leather. Electronic J Env Agri Food Chem 9: 101-111.
- Karmas E, Harris RS (1998) Nutritional evaluation of food processing. Van Nostrand Reinhold Publishers, USA.
- 40. Dillello RL (1982) Standard plate count Method in food and diary Microbial, A laboratory training manual for quality control tests.