

Open Access

Effect of Citalopram on Reducing Transportation Stress in Rainbow Trout (*Oncorhynchus mykiss*)

Esfandani Keysomi MM^{1*}, Sudagar M¹ and Nasirir Asl²

¹Department of Fishery, University of Agricultural Sciences and Natural Resources, Gorgan, Iran ²Department of Pharmacology, School of Medicine, University of Medical Sciences, Qazvin, Iran

Abstract

Rapid Communication

It is often necessary to transfer live fish between aquaculture facilities to permit on-growing or fishing, handling and the physical disturbances associated with loading, transport, and discharge could be regarded as the potential to cause distress and injury, and then leading to possible long term health impairment. Fish in different ways keep its stability (homeostasis) after stress responds. Stress response is involved in many physiological changes. One of the main reactions to stress is increasing the plasma cortisol level. Purpose of this study was to examine the effect of a selective serotonin reuptake inhibitor (citalopram) on plasma cortisol, and handling stress of rainbow trout. Immature rainbow trout (*Oncorhynchus mykiss*) with an average of 50 ± 7 g weight set in three treatments, control, acute (5 mg/l for 48 hours) and chronic (5 µg/l for 10 days). Plasma cortisol before transport in the control group was 22.11 \pm 5.33 (ng/ml), chronic dose was 15.99 ± 5.85 (ng/ml) and in the acute treatment was 18.81 ± 7.42 (ng/ml). Mean plasma cortisol after transportation in the control, acute and chronic treatment respectively was 286.01 ± 54.26 , 107.12 ± 25.53 and 239.89 ± 57.56 ng/ml. Based on the results, significant difference observed (p<0.05) between mean plasma cortisol before and after transportation (p<0.05). As a conclusion it can be expressed that, chronic treatment was more effective than acute treatment in reducing handling stress in rainbow trout.

Keywords: Citalopram; Stress; Transport stress; Rainbow trout

Introduction

During transport, significant changes in water quality may occur and these may have numerous adverse effects on the vital physiological processes of fishes [1]. When fish are continuously exposed to management practices such as handling, transportation or confinement, which can elicit stress responses in intensive rearing facilities [2]. It was shown that fishes respond in different ways to maintain homeostasis after stress [3]. Many physiological changes are involved in such a stress response including hematology [4] osmoregulation [5], hormone release, and energetic metabolism [6,7].

Increasing plasma levels of cortisol as one of the most accepted primary response to stress has been previously reported [7-9]. Air exposure produced a 50-fold increase in plasma cortisol concentrations in gilthead sea bream (Sparus aurata) [10]. Two days of confinement procedures increased three-fold plasma cortisol in gilthead sea bream, S. aurata [11]. In another study turbot, (Scopththalmus maximus), net confinement procedures provoked a 10-fold increase in plasma cortisol [12]. The stress response is highly comparable in all vertebrates, and the secretion of both the glucocorticoid hormone corticosterone and cortisol is the main component [13]. In fish, the secretion of cortisol is mainly controlled by the hypothalamus-pituitary-interrenal (HPI) axis [14]. Corticosteroids affect behavior through genomic (slow) and non-genomic (fast) mechanisms in the central nervous system [15,16]. In mammals the immediate increase in corticosteroids leads to escalated aggression propagated by a fast feed forward mechanism [17]. Moreover, regulation of glucocorticoid levels is influenced by behavior [18]. Sloman et al. [19] observed that the basal levels of circulating cortisol in rainbow trout (Oncorhynchus mykiss) prior to social interaction were associated with the outcome of fights for social dominance. Schjolden et al. [18] perceived that there is relevance between aggressive behavior and cortisol content. Adding anesthetic and hypnotic drugs to the transport tank water may be significantly useful to reduce the physiological stress associated with handling [20].

Drugs are primarily used to slow down metabolism, thus this will lead to reducing oxygen consumption and ammonia and carbon dioxide production. Furthermore they will mitigate the stress response caused by excitement and handling, and reduce swimming [21,22]. Serotonin is an important neurotransmitter in the regulation of social interactions in many animals. Correlative studies in numerous vertebrate species, including fish, indicated that, aggressive males have lower relative serotonergic activity than less aggressive males [23]. In the last decade, it was established that selective serotonin reuptake inhibitors (SSRIs) such as fluoxetine and sertraline could be effective tools for chronic elevating serotonin activity. In recent studies the mentioned drugs are used to study the role of serotonin in aggressive behavior [24, 25]. The relationship between the brain serotonergic systems, social status, and aggression has been established through a large number of correlative and experimental studies [26-28]. Thus, the aim of this study is to examine the effects of citalopram (a selective serotonin reuptake inhibitor) that caused reduced aggression in many species, on stress and plasma cortisol levels in rainbow trout.

Materials and Methods

Immature rainbow trout (*Oncorhynchus mykiss*) with 50 ± 7 g body weight were obtained from a fish farm in Iran (Golestan, Zarringol

*Corresponding author: Esfandani keysomi, Department of Fishery Sciences Gorgan University of Agricultural Sciences & Natural Resources, Beheshti Ave., Gorgan-49138-15739, Iran, Tel: 98(171) 245965 Ext:464/ +98-911-170-8928; Fax: 98(171) 2245884, E-mail: mm.esfandani@gmail.com

Received December 11, 2012; Accepted January 21, 2013; Published February 02, 2013

Citation: Esfandani Keysomi MM, Sudagar M, Asl N (2013) Effect of Citalopram on Reducing Transportation Stress in Rainbow Trout (*Oncorhynchus mykiss*). J Aquac Res Development 4: 171 doi:10.4172/2155-9546.1000171

Copyright: © 2013 Esfandani Keysomi MM, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Farm). Then they were maintained in small raceways in the freshwater open system (not recirculating system). Experiments were conducted from July to August of 2011.

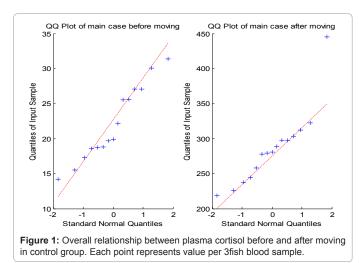
The fish randomly selected and placed in three separate tanks (200 L) and adaptation was carried out for one week. Fish were fed once a day with a commercial diet until 24 h before the experiments. Physical and chemical water parameters were measured at the beginning and during the experiments. Water temperature was 16°C, pH was about 7.2 and water hardness was 145-155 (DH).

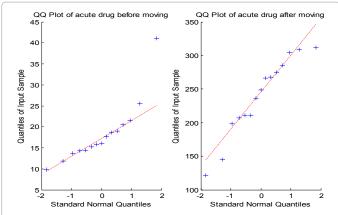
The fish was set in three doses of citalopram including acute (5 mg/l for 48 hours), chronic (5 μ g/l for 10 days) and control treatment. We set 45 fish for each treatment, due to a wide individual difference in fish, each blood sample collected from 3 fishes and then mixed together. For inducing stress to fish, individuals were netted and carried (air exposure 10-15 s (to a transport tanks (40l vol). Fish were transported for 2 hours with a small truck with constant oxygenation. After anesthesia with clove oil blood sample were immediately taken (with 1-ml heparinized syringes) from each fish without any anticoagulant material, The blood collection lasted less than 3 min in order to avoid the cortisol rise induced by the manipulation during sampling [29]. The extracted blood was left to clot at 4°C and centrifuged at 6000 rpm for 5 min at room temperature. The collected serum was stored at 80°C for cortisol assay and then samples were transported to laboratory for plasma cortisol measurements.

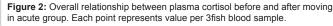
Statistical tests were done by use of a statistical program, SPSS 15.0 for Windows. A one-way ANOVA test was thereafter performed at each sampling time to test for differences among the groups. Data analysis performed with p-value=0.05.

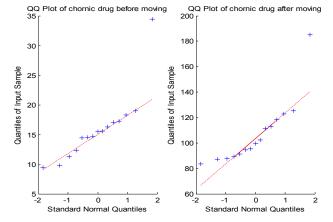
Results

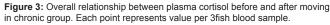
Mean plasma cortisol levels in the control group before transport was 22.11 \pm 5.33 ng/ml and after the transport was 286.01 \pm 54.26 ng/ml (Figure 1). In the acute dose mean plasma cortisol was 18.81 \pm 7.42 ng/ml and after transport was 107.12 \pm 25.53 ng/ml (Figure 2). Mean plasma cortisol in chronic treatment was 15.99 \pm 5.85 ng/ml, before transport was 239.89 \pm 57.56 ng/ml (Figure 3). Based on the results, the difference between mean plasma cortisol in chronic dose before and after the transport was significant (p<0.05). Also, there were significant differences (p<0.05) between the mean plasma cortisol before and after transportation in acute treatment (Figure 4).

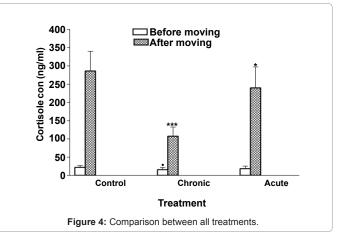












Significant differences between treatments at the same time point are denoted by asterisk (*): *P<0.05, **P<0.01, ***P<0.001.

Discussion

The relationship between brain serotonergic systems, social status, and aggression has been established through a large number of correlative and experimental researches or studies [23]; on the other hand, in other researches (e.g. Schjolden et al. [18]) the relevance between stress and

aggression has been demonstrated. The results showed that artificial modification of the serotonin, which is an important neurotransmitter, can affect the stress behavior in rainbow trout. Because this drug is absorbed through the fish gills and its absorption is depending on temperature and fish metabolism, (e.g. in lower temperatures drug absorption is less), and because in the prolonged duration of exposure in chronic dose, the bioavailability of drug in the fish body will increase. As a conclusion it can be expressed that, chronic treatment was more effective than acute treatment in reducing handling stress in rainbow trout. On the other hand, various drug onset times may be the reason for variant results in the experiment. Notably, this study did not survey other effects of citalopram on various biological factors of (fish); therefore more research should be carried out in order to make perfect this study perfect.

References

- King RH (2009) Fish transport in the aquaculture sector: An overview of the road transport of Atlantic salmon in Tasmania. Veterinary Behavior 4: 163-168.
- Davis CR, Okihiro MS, Hinton DE (2002) Effects of husbandry practices, gender, and normal physiological variation on growth and reproduction of Japanese medaka, *Oryzias latipes*. Aquat Toxicol 60: 185-201.
- Wedemeyer GA, Barton B, McLeay DJ (1990) Stress and Acclimation. In: Schreck CB, Moyle PB (eds.) Methods for Fish Biology. American Fisheries Society, Bethesda, Maryland, pp. 451-489. Chapter 14.
- Dethloff GM, Schlenk D, Khan S, Bailey HC (1999) The effects of copper on blood and biochemical parameters of rainbow trout (*Oncorhynchus mykiss*). Arch Environ Contam Toxicol 36: 415-423.
- McDonald G, Milligan L (1997) Ionic, Osmotic and Acid –Base Regulation in Stress. In: Iwama GK, Pickering AD, Sumpter JP, Schreck, CB (eds.) Fish Stress and Health in Aquaculture. Cambridge University Press, Cambridge, UK, pp. 119-144.
- Carragher JF, Rees CM (1994) Primary and secondary stress responses in golden perch, *Macquaria ambigua*. Comparative Biochemistry and Physiology 107A: 49-56.
- Barton B, Iwama G (1991) Physiological changes in fish from stress in aquaculture with emphasis on the response and effects of corticosteroids. Annual Review of Fish Diseases 1: 3-26.
- Ortuño J, Esteban MA, Meseguer J (2002) Lack of effect of combining different stressors on innate immune responses of seabream (*Sparus aurata* L.). Vet Immunol Immunopathol 84: 17-27.
- Barton BA (2002) Stress in fishes: a diversity of responses with particular reference to changes in circulating corticosteroids. Integr Comp Biol 42: 517-525.
- Arends RJ, Mancera JM, Munoz JL, Wendelaar Bonga SE, Flik G (1999) The stress response of the gilthead sea bream (*Sparus aurata* L.) to air exposure and confinement. J Endocrinol 163: 149-157.
- Tort L, Gomez E, Montero D, Sunyer O (1996) Serum haemolytic and agglutinating activity as indicators of fish immunocompetence: their suitability in stress and dietary studies. Aquaculture International 4: 31-41.
- Waring CP, Stagg RM, Poxton MG (1996) Physiological responses to handling in the turbot. J Fish Biol 48: 161-173.
- Mommsen TP, Vijayan MM, Moon TW (1999) Cortisol in teleosts: dynamics, mechanisms of action, and metabolic regulation. Reviews in Fish Biology and Fisheries 9: 211-268.
- Donaldson EM, Khansari F, Ghazi-Khansari M, Abdollah M (2005). The pituitary-interrenal axis as an indicator of stress in fish. In: Pickering AD (Ed) Stress in fish. London: Academic Press 93: 11-47.

- Haller J, Halasz J, Makara GB, Kruk MR (1998) Acute effects of glucocorticoids: behavioral and pharmacological perspectives. Neurosci Biobehav Rev 23: 337-344.
- DiBattista JD, Anisman H, Whitehead M, Gilmour KM (2005) The effects of cortisol administration on social status and brain monoaminergic activity in rainbow trout (*Oncorhynchus mykiss*). J Exp Biol 208: 2707-2718.
- Kruk MR, Halasz J, Meelis W, Haller J (2004) Fast positive feedback between the adrenocortical stress response and a brain mechanism involved in aggressive behavior. Behav Neurosci 118: 1062-1070.
- Schjolden J, Basic D, Winberg S (2009) Aggression in rainbow trout is inhibited by both MR and GR antagonists. Physiol Behav 98: 625-630.
- Sloman K, Metcalfe NB, Taylor AC, Gilmour KM (2001) Plasma cortisol concentrations before and after social stress in rainbow trout and brown trout. Physiol Biochem Zool 74:383-389.
- Finstad B, Iversen M, Sandodden R (2003) Stress-reducing methods for releases of Atlantic salmon (*Salmo salar*) smolts in Norway. Aquaculture 222: 203-214.
- Wedemeyer GA (1997). Rearing conditions: effects on fish in intensive culture. In: Iwama GK, Pickering AD, Sumpter JP, Schreck CB (eds.) Fish Stress and Health in Aquaculture Society. Experimental. Biology. Seminar Series, vol. 62. University Press, Cambridge pp. 35-71.
- Ross LG, Ross B (1999) Anesthetic and sedative techniques for aquatic animals, (2ndedn.), Blackwell, London, UK, pp: 159.
- Perreault HA, Semsar K, Godwin J (2003) Fluoxetine treatment decreases territorial aggression in a coral reef fish. Physiol Behav 79: 719-724.
- 24. Fuller RW (1996) The influence of fluoxetine on aggressive behavior. Neuropsychopharmacology 14: 77-81.
- 25. Larson ET, Summers CH (2001) Serotonin reverses dominant social status. Behav Brain Res 121: 95-102.
- 26. Blanchard DC, Cholvanich P, Blanchard RJ, Clow DW, Hammer RP, et al. (1991) Serotonin, but not dopamine, metabolites are increased in selected brain-regions of subordinate male rats in a colony environment. Brain Res 568: 61-66.
- Raleigh MJ, McGuire MT, Brammer GL, Pollack DB, Yuwiler A (1991) Serotonergic mechanisms promote dominance acquisition in adult male vervet monkeys. Brain Res 559: 181-190.
- Winberg S, Carter CG, McCarthy JD, He ZY, Nilsson GE, et al. (1993) Feeding rank and brain serotonergic activity in rainbow trout (*Oncorhynchus mykiss*). J Exp Biol 179: 197-211.
- Acerete L, Balasch JC, Espinosa E, Josa A, Tort L (2004) Physiological responses in Eurasian perch (Perca fluviatilis, L.) subjected to stress by transport and handling. Aquaculture 237: 167-178.