

Effect of *Capsicum annum* L. and *Capsicum frutescens* L. Varieties Extracts on *in vitro* Growth of Fungal Strains

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ABSTRACT

This study aims to test different extracts (water, 70% ethanolic-water and acetal) of *Capsicum* sp. on the growth of *Alternaria* sp., *Penicillium* sp., *Fusarium* sp. and *Aspergillus flavus*. The nature of the antifungal activity, the antifungal parameters (MIC, MFC, IC50) and the percentage of survival of the strains were determined with the dilution method.

On *Alternaria* sp., *Penicillium* sp. and *Fusarium* sp. strains, values of MFC/MIC ratios of water and 70% ethanolic extracts of *Capsicum annum* antillais were equal to two. Concerning *Capsicum frutescens* (soudanais, attié, doux and oiseau) varieties, these values varied from 1 to 2 on the fungal strains.

These ratios for most of the strains were lower than 4. These extracts exerted a fungistatic action on chosen strains. The curves obtained had a declining appearance with more or less steep slopes depending on the extracts. These curves were used to obtain the inhibitory concentrations for 50% survival (IC50) of the moulds. The highest activity was achieved at the lowest IC50 values which varied from 0.2 mg/mL to 0.3 mg/mL. Therefore, *Capsicum* extracts proved to be active on *in vitro* growth of the fungal strains studied in a dose-dependent manner.

Keywords: *Capsicum*; Antifungal activity; Dose-dependent; Antifungal parameters

INTRODUCTION

Medicinal plants have been used for centuries for various purposes by civilisations [1]. Plants are used as food, energy, clothing, and housing constructions. Sometimes, they are used to cure diseases [2]. According to these some authors, “soft” medicines, particularly herbal medicine, are gaining considerable interest in many parts of Africa, Asia and Europe. Worldwide, there are serious and life-threatening diseases caused by microorganisms such as bacteria, viruses, fungi and parasites [3]. For a very long time, humans have been relatively disarmed by the diseases caused by fungi. Over the last twenty years, a large increase in these diseases in which affect the wide range of hosts has been observed. These diseases are caused by surprisingly high number of fungal species [4]. They cause serious damage to humans as well as to plant species [5]. Food industries are also facing problems such as appearance changing, organoleptic and chemical qualities spoilage, non-stable altered substances produced,

that food can appear during conservation [6]. The need for new prevention strategies against fungal infections and diseases is a major interest, not only for food safety, but also for consumer health, and moreover for the protection of countries’ economy. Thus, at the therapeutic level, there are several antifungals. However, some of these molecules sold on the market and therapeutically used have lost their effectiveness due to antifungal resistance and are, out of reach for African populations with low incomes due to their high cost [7]. Therefore, these populations are increasingly using medicinal plants their health issues. According to the annual report of the World Health Organization (WHO), about 80% of the African population use plants for treatment of ailments [8,9]. However, the overuse of these plants exposes them to various accidents because; traditional medicines may contain other molecules having toxic effects. In Africa, pepper is always cited among the therapeutic arsenal [10]. Especially, *Capsicum* fruits are used in traditional medicine for their antimicrobial and antifungal properties as condiment [11,12]. As a

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result, the use of *capsicum* extracts in the fight against mould from food causes the choice of our subject.

MATERIALS AND METHODS

Plant materials

Plant materials consisted of fruits of *Capsicum annuum* variety *antillais*, *Capsicum frutescens* variety *doux*, *Capsicum frutescens* variety *attié*, *Capsicum frutescens* variety *soudanais*, *Capsicum frutescens* variety *oiseau* and *Capsicum frutescens* variety *pendulum*. Six varieties from fruits coming from *Capsicum annuum*, *Capsicum frutescens* and *Capsicum chinense* were chosen. These varieties are fresh, mature and firm fruits of *Capsicum annuum* var. *antillais* and *Capsicum annuum* var. *pendulum* were used. For the other varieties (*Capsicum frutescens* var. *doux*, *Capsicum frutescens* var. *attié*, *Capsicum frutescens* var. *soudanais*, *Capsicum frutescens* var. *oiseau* and *Capsicum chinense* var. *pendulum*) based on the way they are traditionally used by local people, air dried materials were used from food were used for the tests. These different strains were sub cultured and stored in nutrient agar at 4°C. The varieties of peppers are obtained directly from five main district of Abidjan markets: The large market of Abobo, the large market of Yopougon, the large market of Adjamé, the large market of Treichville and the large market Port Bouët. Fruit varieties of *Capsicum annuum* variety *antillais* and *Capsicum chinense* variety *pendulum* are obtained fresh while those varieties of *Capsicum frutescens* (*soudanais*, *attié doux* and *oiseau*) are desiccated (dried form). These varieties have identified all summers University Floristic National Centre Félix Houphouët-Boigny (Côte d'Ivoire).

One hundred and twenty samples of peppers were collected and kept in sterile sampling bags. The fruits of *Capsicum* were sorted then washed under tap water and dried in an oven at 55°C for eight to ten days for fresh fruits and five days dried fruits. The dried fruits were reduced to fine powder using an electronic mixer at 3000 rpm then sieved (1 mm) to obtain the final powder (Figure 1).

Microorganism

Five fungal strains, namely *Aspergillus niger*, *Fusarium sp.*, *Alternaria sp.*, *Aspergillus flavus* and *Penicillium sp.* They are foodborne pathogens and they also cause epidermal diseases, respiratory allergies, vomiting, and sinusitis [13]. In fact, all these strains were provided by the Institute Pasteur in Côte d'Ivoire (IPCI).



Figure 1: Chilli powder packaged in Stomacher sachets.

Extract preparation

The extracts (Water, 70% water-ethanol and acetate) were prepared according to successive extraction methods developed and described by Guedé Guina et al. [14] from the chili pepper powder previously obtained.

Thus, 50 grams were extracted by homogenization in one litre (1 L) of distilled water. The mixture was filtered through Whatman paper No.2 (Whatman International, Maidstone England) and then oven evaporated at 60°C until completely dry. The powders obtained constitute the crude aqueous extract coded Etaq.

25 grams of the crude aqueous extract were dissolved in 500 mL of ethanol (70%). Solution per homogenization during 24 hours with room temperature (25-30°C). The mixture was filtered through Whatman paper No. 2 and the filtrate was concentrated in a rotavapor at 50°C. The powders obtained constitute the ethanol 70% coded Eeth 70%.

10 grams of ethanolic 70% extract are dissolved in 500 mL of a solution composed of a mixture of ethyl acetate and distilled water (v/v). The mixture was mixed for 24 hours using a magnetic stirrer. The homogenate was filtered through Whatman paper No. 2 and the filtrate was concentrated in a rotavapor at 50°C. The dried extracted product constituted the acetatic 1 coded Eace 1.

Performance calculation

The yield of extract expressed on dry weight basis of pepper powder, was calculated from the following formula: $Rd (g/100 g) = (M_1 \times 100) / M_0$ where Rd: Yield; M1: weight of the extract residue obtained after solvent removal and M0: weight of the chilli powder taken.

Antifungal tests

Antifungal tests of *Capsicum* extracts were carried out with subcultures of each fungal species were made on Sabouraud agar. Agar well diffusion was used to determine antimicrobial activity [15,11] and antifungal parameters (MIC, MFC, IC50) were determined by tube dilution method [16].

Minimum inhibitory concentrations determination

Antifungal activities of pepper varieties were determined in sterile haemolysis tubes. The concentration ranges of the *Capsicum* extracts of each strain initially prepared according to the double dilution method in liquid medium, namely 6.72; 3.36; 1.68; 0.84; 0.42; 0.21 and 0.10 mg/mL were divided into seven sterile haemolysis tubes 1 mL per tube. An inoculum whose turbidity was adjusted to 10^8 CFU/mL in Sabouraud broth was also prepared for each fungal strain. In each of the seven haemolysis tubes containing 1 mL of each *Capsicum* extract, 1 mL of the previously prepared fungal inoculum was added. Two control tubes were prepared: A growth control tube containing 1 mL of sterile distilled water and 1 mL of fungal inoculum, followed by a sterility control tube containing 2 mL of Sabouraud broth. The contents of each haemolysis tube thus prepared was homogenized by a vortex mixer and then incubated at 25°C for 72 hours.

The experiments were repeated three times to ensure the reliability of the results. After distribution of the broth inoculated into the seven experimental tubes, the remaining broth was diluted to 10^{-4} . The dilutions obtained were subsequently seeded onto plates of Petri containing Sabouraud agar, by 5 cm streaks (box A) and incubated at 25°C for 72 hours [17].

Minimum fungicide concentration (MFC)

The MFC was determined in an agar medium after reading the MIC. To do this, the different cultures in each experimental tube were seeded by 5 cm streaks on Sabouraud agar using a calibrated loop. On each seeded agar plate (box B), there is the inoculum of the control tube of growth control, the inoculum where the turbidity was not visible, that of the tube which made it possible to determine the MIC. The different seedlings were incubated at 25°C for 72 hours. The determination of MFC was possible by comparing the number of colonies on the streaks of the dish containing the remainder of the diluted broth up to 10⁻⁴ (box A), with that of box B [18].

Number of antimicrobial effect of plant extract

The MFC/MIC ratio was calculated for each extract to determine the nature of antimicrobial effects of plant extracts. When, the MFC/MIC ratio <4, the phytochemical has a fungicidal effect. However, if the MFC/MIC ratio ≥ 4 the plant extract has a fungistatic effect [19].

Percentage survival (Cytotoxic activity)

The fungal flora tested was counted by direct counting using a colony counter. Their growth in the experimental dishes was evaluated as a percentage of survival calculated with respect to 100% survival in the control boxes. The method of calculating the percentage of survival of the seeds in the different experimental boxes is translated by the following formula.

$$S = (n/N) \times 100.$$

S=survival of fungal germs (%)

n=number of colonies in the experimental petri dishes

N=number of colonies in the controls petri dishes

RESULTS

Yield of different extracts

Table 1 surmises the extraction yield for different solvents and

different *C. annuum* and *C. frutescens* varieties. The extraction yield is lower for acetate extract of *Capsicum frutescens doux* (3.2%) and higher for water extract (24.8%) of *C. chinense pendulum*.

Antifungal parameters

Tables 2 and 3 show antifungal activities of various extract of *Capsicum* varieties used in Ivorian traditional pharmacopeia. The most interesting activity on *Alternaria sp.* was obtained with the lowest value of MIC (0.84 mg/mL) was observed with *Capsicum chinense pendulum* extracts on *Alternaria sp.*

The lowest concentration was observed with the acetal extract of *Capsicum chinense var. pendulum* (3.36 mg/mL) and *Capsicum frutescens* (6.72 mg/mL) on *Aspergillus flavus*.

For the other varieties and extracts, the MIC could not be determined because there was growth in all experimental tubes. The 70% ethanol extract was the most effective extract.

The lowest minimum fungicide concentration was obtained with the 70% ethanol extract of the fruits of *Capsicum chinense var. pendulum* (0.84 mg/mL) followed by the acetate and the aqueous then ethanol extracts of *Capsicum annuum var. antillais* with 3.36 mg/mL each.

The MFC/MIC ratios of the aqueous and 70% ethanol extracts

<i>Capsicum</i> varieties	Extraction solvent (%)		
	Aq	EtOH 70%	Ace
<i>C. annuum antillais</i>	24.6	10.2	4.5
<i>C. frutescens soudanais</i>	23.7	9.8	3.6
<i>C. frutescens attié</i>	23.6	9.5	3.4
<i>C. frutescens doux</i>	23.1	9.1	3.2
<i>C. frutescens oiseau</i>	23.4	9.2	3.3
<i>C. chinense pendulum</i>	24.8	10.4	4.7

C.: *Capsicum*; Aq: water extract; EtOH 70%: 70% ethanol extract; Ace: acetone extract

Table 1: Extraction yield for different extracts and different varieties of *Capsicum*.

<i>Capsicum</i> varieties	Extracts	<i>Alternaria sp.</i>	<i>Penicillium sp.</i>	<i>Fusarium sp.</i>	<i>Aspergillus flavus</i>
<i>C. annuum var. antillais</i>	water	2	2	2	-
	Ethanol 70%	2	2	2	-
	Acetate	4	-	-	-
<i>C. frutescens var. soudanais</i>	water	2	-	-	-
	Ethanol 70%	2	-	2	-
	Acetate	-	-	-	-
<i>C. frutescens var. attié</i>	water	1	-	-	-
	Ethanol 70%	-	-	-	-
	Acetate	-	-	-	-
<i>C. frutescens var. doux</i>	water	2	-	-	-
	Ethanol 70%	-	2	-	-
	Acetate	-	-	-	-
<i>C. frutescens var. oiseau</i>	water	2	-	-	-
	Ethanol 70%	2	-	1	-
	Acetate	-	-	-	1
<i>C. chinense var. pendulum</i>	water	1	4	-	-
	Ethanol 70%	1	2	2	-
	Acetate	2	-	2	2

C.: *Capsicum*; (-): not evaluate; A: *Aspergillus flavus*

Table 2: *In vitro* antifungal effect of *Capsicum sp.* extract on the growth of different strains.

<i>Capsicum</i> varieties	Extracts	<i>Alternaria</i> sp.	<i>Penicillium</i> sp.	<i>Fusarium</i> sp.	<i>Aspergillus flavus</i>
<i>C. annuum</i> var. antillais	water	1.68	3.36	3.36	-
	Ethanol 70%	1.68	1.68	3.36	-
	Acetate	1.68	-	-	-
<i>C. frutescens</i> var. soudanais	water	3.36	-	-	-
	Ethanol 70%	1.68	-	3.36	-
	Acetate	-	-	-	-
<i>C. frutescens</i> var. attié	water	6.72	-	-	-
	Ethanol 70%	-	-	-	-
	Acetate	-	-	-	-
<i>C. frutescens</i> var. doux	water	3.36	-	-	-
	Ethanol 70%	-	3.36	-	-
	Acetate	-	-	-	-
<i>C. frutescens</i> var. oiseau	water	3.36	-	-	-
	Ethanol 70%	3.36	-	6.72	-
	Acetate	-	-	-	6.72
<i>C. chinense</i> var. pendulum	water	1.68	3.36	-	-
	Ethanol 70%	0.84	1.68	3.36	-
	Acetate	1.68	-	3.36	3.36

Table 3: Minimal Inhibitory Concentrations (MICs) of *Capsicum* extracts on four fungal strains.

<i>Capsicum</i> varieties	Extracts	<i>Alternaria</i> sp.	<i>Penicillium</i> sp.	<i>Fusarium</i> sp.	<i>Aspergillus flavus</i>
<i>C. annuum</i> var. antillais	water	3,36	6,72	6,72	-
	Ethanol 70%	3,36	3,36	6,72	-
	Acetate	6,72	-	-	-
<i>C. frutescens</i> var. soudanais	water	6,72	-	-	-
	Ethanol 70%	3,36	-	6,72	-
	Acetate	-	-	-	-
<i>C. frutescens</i> var. attié	water	6,72	-	-	-
	Ethanol 70%	-	-	-	-
	Acetate	-	-	-	-
<i>C. frutescens</i> var. doux	water	6,72	-	-	-
	Ethanol 70%	-	6,72	-	-
	Acetate	-	-	-	-
<i>C. frutescens</i> var. oiseau	water	6,72	-	-	-
	Ethanol 70%	6,72	-	6,72	-
	Acetate	-	-	-	6,72
<i>C. chinense</i> var. pendulum	water	1,68	6,72	-	-
	Ethanol 70%	0,84	3,36	6,72	-
	Acetate	3,36	-	6,72	6,72

Table 4: Minimum Fungicidal Concentrations (MFCs) of *Capsicum* extracts on fungal strains.

of *Capsicum annuum* var. *antillais* were equal to 2 on *Alternaria* sp., *Penicillium* sp. and *Fusarium* sp. strains. This ratio is 4 for the acetate extract on *Alternaria* sp.

Capsicum frutescens species (*sudanese*, *attié*, *doux* and *oiseau*) provided MFC/MIC ratios, varying from 1 to 2 on the fungal strains.

On *Capsicum chinense* var. *pendulum*, the values of these ratios varied from 1 to 4 with the extracts and the fungal strains (Table 2).

The smallest minimal inhibitory concentration of *Capsicum annuum* var. *antillais* (MIC) extract was 1.68 mg/mL on *Alternaria* sp. The smallest MIC was observed for *Capsicum chinense* var. *pendulum* extracts on *Alternaria* sp. (0.84 mg/mL).

On *Aspergillus flavus*, the lowest MIC was observed with the ethyl acetate extracts of *Capsicum chinense* var. *pendulum* (3.36 mg/mL) and *Capsicum frutescens* (6.72 mg/mL). The MIC for the other

varieties and extracts could not be determined, as there was growth in all experimental tubes (Table 3).

The results of the minimum fungicidal concentrations (MFCs) range from 0.84 to 6.72 mg/mL revealed that the 70 % ethanol extract *Capsicum chinense* var. *pendulum* was the lowest concentration (0.84 mg/mL), followed by the acetate, water and ethanolic extracts of *Capsicum annuum* var. *antillais* (3.36 mg/mL each). On *Aspergillus flavus* strain, the fungicidal minimal concentration was obtained only in the presence of *Capsicum frutescens* var. *oiseau* and *Capsicum chinense* var. *pendulum* acetate extracts (6.72 mg/mL each) (Table 4).

Survival efficiency

Representative survival curves of *Alternaria* sp., *Penicillium* sp. and *Fusarium* sp. in presence of *Capsicum annuum* var. *antillais* and *C. chinense* var. *pendulum* are presented in Figures 2 and 3.

With *Capsicum annum* var. *antillais* extracts, the lowest IC₅₀ value was obtained for 70% ethanolic extract on *Alternaria* sp. and 0.3 mg/mL on the *Penicillium* sp. The lowest activity of the water extract was obtained at the highest IC₅₀ values. These values were 0.3 mg/mL on *Alternaria* sp., 0.5 mg/mL on *Penicillium* sp. and 1 mg/mL on *Fusarium* sp. On *Alternaria* sp. acetone extract and 70% ethanolic extract showed the same IC₅₀ (0.2 mg/mL). Thus, *Alternaria* sp. is the most sensitive strain.

Indeed, the curves have steeper slopes in the presence of different extracts of *Capsicum*. The least sensitive strain is *Penicillium* sp. (Figure 2).

On *Capsicum chinense* var. *pendulum* extracts, the lowest IC₅₀ (0.1 mg/mL) was obtained with the ethanol extract while the highest IC₅₀ was obtained with on *Alternaria* sp.

Water and ethanol extracts showed the same IC₅₀ (0.2 mg/

mL). The IC₅₀ of the 70% ethanol extract on *Penicillium* sp. and *Aspergillus flavus* were respectively 0.2 and 0.3 mg/mL.

While the IC₅₀ of the acetate extract was the lowest on *Fusarium* sp. and *Aspergillus flavus*, 0.4 mg/mL and 0.5 mg/mL respectively. Whatever the extract tested, *Alternaria* sp. was the most sensitive. While *Aspergillus flavus* and *Aspergillus niger* were the most resistant.

DISCUSSION

In order to study the effect of *Capsicum* extracts on the growth of fungal strains, extractions using different solvents were performed with the fruits of six *Capsicum* varieties. We found that water extract lead to the best extraction ratio. Thus water may be the best solvent to extract the maximum of component from *Capsicum*, compared to ethanol and ethyl acetate solvents. However, according to Bouharb et al. [20-22], the yield of extraction should not always be considered as effectiveness criteria. Indeed, the best extraction

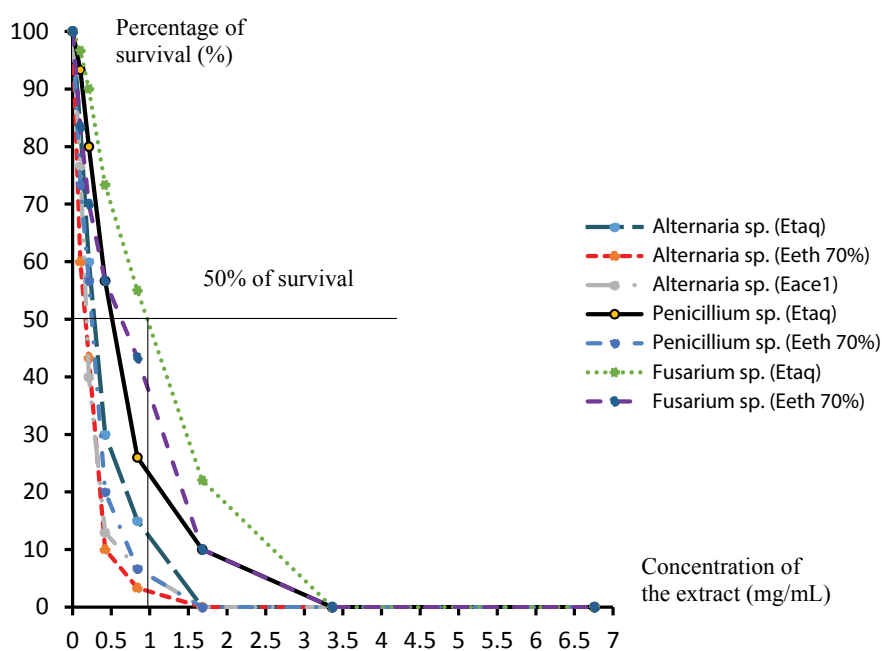


Figure 2: Survival curve of *Alternaria* sp., *Penicillium* sp., *Aspergillus flavus* and *Fusarium* sp. strains at different concentrations of *Capsicum annum* var. *antillais* extracts.

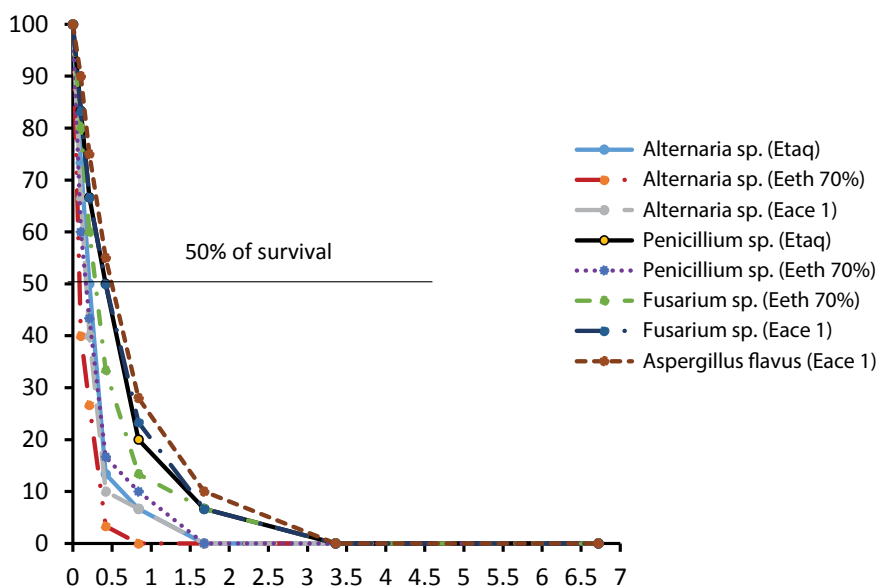


Figure 3: Survival curve of *Alternaria* sp., *Penicillium* sp., *Aspergillus flavus* and *Fusarium* sp. strains at different concentrations of *Capsicum chinense* var. *pendulum* extracts.

yields were obtained with the water extracts, while 70% ethanol extracts exhibited the best antifungal activity.

However, it could be difficult to compare the yield extraction results of this study with those of other authors, because it was not only related on the extraction methods and its conditions. The plants and the organ collected may also influence the yield ratio. Indeed, according to Adjou and Soumanou [23], the extraction yield could influence by the harvesting site, the nature of the soil, the stage of development of the plant and the organ used. The different varieties of *Capsicum* used exhibited antifungal activity on moulds from food. The culture wells method used showed a high diffusion of the extracts in the agar. This study gives consistent results with those obtained by Kouassi and Koffi-Nevry [11] who showed that extracts of *Capsicum* varieties diffuse better with the well method because the extracts are in direct contact with the agar.

None of the extracts had any effect on the growth of *Aspergillus flavus*. *A. flavus* is the less sensitive strain. These results corroborate those of Tournas and Katsoudas [24] and those of Ouattara et al. [25] which showed that *Aspergillus sp.* are particularly resistant to therapeutic agents. *Aspergillus flavus* is one of the most resistant microorganisms to antifungals [24]. Koffi-Nevry et al. [26] reported that *Aspergillus sp.* was the least sensitive strain in presence of Polyhexamethylene-Guanidine Hydrochloride Disinfectant (PHMGH) on *Mucor sp.*, *Botrytis sp.*, *Penicillium sp.*, *Geotrichum sp.*, *Aspergillus sp.*, and *Colletotrichum sp.* isolated from the papaya Delahousse [27] showed that some plants having good activity also have an inhibitory effect on *Penicillium sp.*, *Alternaria sp.* and *Aspergillus sp.* strains.

Capsicum extracts activities vary depending on the nature of the extract and the mould strains. This activity could be due to the phytochemicals present in pepper fruit.

Activities from 70% Ethanolic extracts of caribbean and *pendulum* varieties exhibited antifungal activity higher than those of water extracts. These results could be explained by the fact that the 70% ethanolic extract could be rich in the specific compounds (phenolic and terpene compounds) capable of destroying fungi. These results may be due to the fact that the 70% ethanolic extract could have more phytochemicals such as phenols and terpenes capable of destroying fungal strains. Moreover, *Capsicum annum* var. *antillais* and *Capsicum chinense* var. *pendulum* are rich in these compounds. Ethanol extracts appeared to be more active than aqueous extracts [28,21]. The lowest values of antifungal parameters were obtained with *Alternaria sp.* strains in the presence of ethanol extracts of *Capsicum chinense* var. *pendulum*. The highest values were obtained with the aqueous extracts of *Capsicum frutescens* *attié* on *Alternaria sp.* and with acetate extracts of *Capsicum frutescens* *bird* on *Aspergillus flavus*. According to Daroui-Mokaddem [28], inhibition diameters are related to the desired MIC values, so *capsicum* extracts with the highest inhibition diameters are those which would possess the lowest MIC. The sensitivity curves to the extracts revealed differences of sensitivity between the fungal strains. In fact, the IC₅₀s of the water, 70% ethanolic and acetate extracts of the *pendulum* variety on *Alternaria sp.* were lower than the IC₅₀s of these extracts on *Penicillium sp.* and *Fusarium sp.* Our study demonstrates that the ethanol extracts of *Capsicum annum* var. *antillais* and *Capsicum chinense* var. *pendulum* are the most active extracts. *Alternaria sp.* was the most sensitive strain regardless of the extract used.

CONCLUSION

Both the 70% ethanol and the water extract were the most active extracts on the fungal strains. *Alternaria sp.*, *Penicillium sp.* and *Fusarium sp.* were sensitive to the extracts used. However, the most sensitive strain is *Alternaria sp.* and the lowest one is *Aspergillus flavus*. In the absence of an extract (0 mg/ml), the germs survival is 100%. Nevertheless, any concentrations of extracts added to the culture medium, lead to progressively decrease the growth of fungal strains. The *capsicum* extracts are therefore active on the *in vitro* growth of the fungal strains studied in a dose-dependent manner. This study provides an undeniable scientific argument for the traditional use of *capsicum* varieties in the treatment of infections due to microorganisms.

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