

Current Therapeutic Drugs Treating COVID-19

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ABSTRACT

Corona virus disease 2019 (COVID-19) is a kind of viral pneumonia with an unusual outbreak in Wuhan, China, in December 2019, which is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Since the outbreak of corona virus disease 2019 (COVID-19), many researchers in the world have immediately carried out clinical research scheme of the COVID-19. Vaccines are being rapidly developed but will likely come too late to have an impact on the first wave of a potential pandemic. It highlights antiviral strategies involving small molecules and biologics targeting complex molecular interactions involved in coronavirus infection and replication. The drug-repurposing effort documented herein focuses primarily on agents known to be effective against other RNA viruses including SARS-CoV and MERS-CoV. The patent analysis of coronavirus related biologics includes therapeutic antibodies, Anti-inflammatory agents, and nucleic acid-based therapies targeting virus gene expression as well as various types of vaccines. Many patents disclose methodologies of these biologics with the potential for treating and preventing coronavirus infections, which may be applicable to COVID-19. But, there is still a lack of a review on Therapeutic Agents and Vaccines for COVID-19. Therefore, this review summarizes studies which are ongoing development of therapeutic agents and vaccines.

Keywords: COVID-19; Therapeutic drugs; Vaccines

INTRODUCTION

Novel corona virus induced pneumonia, which was named as corona virus disease 2019 (COVID-19) by the WHO on the 11th of February 2020, has rapidly increased in epidemic scale since it first appeared in Wuhan, China, in December 2019 [1]. At present, the cases of COVID-19 have been found in many countries around the world [2]. Corona Virus Disease 2019 (COVID-19), being an emerging infectious disease, is a serious threat to human health [3-5]. According to the latest data, up to March 23, 2020 the number of confirmed cases in world reached 351,083, of which 15,337 were dead, and 100,569 were cured.

To date, no clinical intervention trial has been completed and reported. Due to the urgent need for treatment prevention and control of the disease, it is necessary to develop effective intervention methods for COVID-19 to facilitate disease control. Since the outbreak of the COVID-19, many researchers in China have carried out clinical research trials, aiming to develop strategies for the treatment, prevention and diagnosis of COVID-19 [6].

Corona Viruses (CoVs) are relatively large viruses containing a single-stranded positive-sense RNA genome encapsulated

within a membrane envelope. The viral membrane is studded with glycoprotein spikes that give corona viruses their crown like appearance. While corona viruses infect both humans and animals, certain types of animals such as bats that host the largest variety of corona viruses appear to be immune to corona virus-induced illness [7]. There are four classes of corona viruses designated as alpha, beta, gamma, and delta. The beta corona virus class includes Severe Acute Respiratory Syndrome (SARS) virus (SARS-CoV), Middle East Respiratory Syndrome (MERS) virus (MERS-CoV), and the COVID-19 causative agent SARS-CoV-2. Similar to SARS-CoV and MERS-CoV, SARS-CoV-2 attacks the lower respiratory system to cause viral pneumonia, but it may also affect the gastrointestinal system, heart, kidney, liver, and central nervous system leading to multiple organ failure [8,9]. Current information indicates that SARS-CoV-2 is more transmissible or contagious than SARS-CoV [10]. Patients with COVID-19 show manifestations of respiratory tract infection, such as fever, cough, pneumonia, and in severe cases, death [11,12]. The information included in this review provides a strong intellectual groundwork for the ongoing development of therapeutic agents and vaccines.

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Received date: February 11, 2021; **Accepted date:** February 25, 2021; **Published date:** March 04, 2021

Citation: Kifle ZD (2021) Current Therapeutic Drugs Treating COVID-19. J Clin Exp Pharmacol. 11:281.

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Existing drugs with potential therapeutic applications for covid-19

Since SARS-CoV-2 is a newly discovered pathogen, no specific drugs have been identified or are currently available. An economic and efficient therapeutic strategy is to repurpose existing drugs. On the basis of genomic sequence information coupled with protein structure modeling, the scientific community has been able to rapidly respond with a suggested list of existing drugs with therapeutic potential for COVID-19 [13].

The main measure in clinical management is focused on alleviating clinical symptoms and supportive care [14]. Therapeutic options that could be evaluated and used for COVID-19 include molecules binding to the virus, molecules, or inhibitors that target specific enzymes involved in viral replication and transcription, small-molecule inhibitors targeting helicase, essential proteases, or other proteins of the virus, host cell protease inhibitors, host cell endocytosis inhibitors, siRNA, anti-sense RNA and ribozyme, neutralizing antibodies, mAbs targeting host receptor or interfere with S1 RBD, antiviral peptide targeting S2, and natural products [15,16]. Baricitinib was proposed because of its anti-inflammatory

effect and possible ability to reduce viral entry [17]. A fixed dose of the anti-HIV combination, lopinavir–ritonavir, is currently in clinical trials with Arbidol or Ribavirin [18]. Remdesivir, developed by Gilead Sciences Inc., was previously tested in humans with Ebola virus disease and has shown promise in animal models for MERS and SARS. The drug is currently being studied in phase III clinical trials in both China and the USA. Favipiravir, a purine nucleoside leading to inaccurate viral RNA synthesis, was originally developed by Toyama Chemical of Japan, and has recently been approved for a clinical trial as a drug to treat COVID-19 [19,20]. Chloroquine, an antimalarial drug, has proven effective in treating coronavirus in China [21]. In addition to the above-mentioned, many other antiviral drugs are also listed.

Additionally, clinicians combined Chinese and Western medicine treatment including lopinavir/ritonavir (Kaletra®), arbidol, and Shufeng Jiedu Capsule (SFJDC, a traditional Chinese medicine) and gained significant improvement in pneumonia associated symptoms in Shanghai Public Health Clinical Center, China [22]. The other antiviral drugs include nitazoxanide, favipiravir, nafamostat, and so on (Table 1).

Table 1: Neutralizing monoclonal antibodies targeting SARS-CoV and their mechanism of action.

Monoclonal antibody	Antibody mechanism of action	Authors
80R	<ul style="list-style-type: none"> • Binding to the conformational epitope (amino acid residues 426-492) on S1 fragment of SARS-CoV. • Blocking the interaction of S1 subunit protein with cellular receptor ACE2 using 6 Complementary Determining Region (CDR) <i>in vitro</i> and <i>in vivo</i> (Mouse). 	Zhang L, et al. Sui J, et al. [91,92].
CR3014	<ul style="list-style-type: none"> • Binding to the amino acid residues 318-510 and amino acid residue 565 with high affinity on S1 fragment of SARS-CoV. • Blocking the interaction of S1 subunit protein with cellular receptor ACE2 <i>in vitro</i> and <i>in vivo</i> (Ferret). 	Brink ENVD, et al. Shanmugaraj B, et al. Ter Meulen J, et al.[93-95].
CR3022	<ul style="list-style-type: none"> • Binding to the amino acid residues 318-510 on S1 fragment of SARS-CoV. • Blocking the interaction of S1 subunit protein (RBD) with cellular receptor ACE2 <i>in vitro</i>. 	Ter Meulen J, et al. [95].
F26G18	<ul style="list-style-type: none"> • Binding to the linear epitope (amino acid residues 460-476) on S1 fragment of SARS-CoV. • Blocking the interaction of S1 subunit protein (RBD) with cellular receptor ACE2 <i>in vitro</i>. 	Berry JD, et al. [96].
F26G19	<ul style="list-style-type: none"> • Binding to the conformational epitope (amino acid residues 359-362, 391-392, 424-427, and 486-492) on S1 fragment of SARS-CoV. • Blocking the interaction of S1 subunit protein (RBD) with cellular receptor ACE2 <i>in vitro</i>. 	Berry JD, et al. [96].
m396	<ul style="list-style-type: none"> • Binding to the conformational epitope (amino acid residues 482-491) on S1 fragment of SARS-CoV. • Blocking the interaction of S subunit protein using CDR loops H1, H2, H3, and L3 with cellular receptor ACE2 <i>in vitro</i>. 	Zhu Z, et al. [97].
1A9	<ul style="list-style-type: none"> • Binding to the Heptad repeat (HR) loops including heptad repeat 1 (HR1) and heptad repeat 1 (HR2) domain on S2 fragment of SARS-CoV. • Blocking the interaction of S2 subunit protein (amino acid residues 1111-1130) with cellular receptor <i>in vitro</i>. 	Shanmugaraj B, et al. Lip KM, et al. [94,98].
201	<ul style="list-style-type: none"> • Binding to the amino acid residues 490-510 on S1 fragment of SARS-CoV. • Blocking the interaction of S1 subunit protein with cellular receptor ACE2 <i>in vitro</i> and <i>in vivo</i> (Mouse Syrian Hamster). 	Greenough TC, et al. [99].
68	<ul style="list-style-type: none"> • Binding to the amino acid residues 130-150 of SARS-CoV <i>in vitro</i> and <i>in vivo</i> (Mouse) 	Greenough TC, et al. [99].
4D4	<ul style="list-style-type: none"> • Binding to the amino acid residues 12-261 of SARS-CoV and N-terminal of RBD • Inhibiting the post-interaction in the viral penetration <i>in vitro</i>. 	Code 71 Coughlin MM, et al. Elshabrawy HA, et al. [57,100].
S230	<ul style="list-style-type: none"> • Binding to epitopes partially overlapping with receptor binding motifs on B domain of SARS-CoV. • Blocking the interaction of S1 subunit protein with cellular receptor ACE2 <i>in vitro</i>. 	Walls AC, et al. [101].

Research and development of vaccines

With the emergence of 2019-nCoV, there are many potential vaccine candidates in the pipeline globally, in which a wide range of technology (such as messenger RNA, DNA-based, nano particle, synthetic and modified virus-like particle) was applied. The cellular receptors of SARS-CoV and MERS-CoV have been identified, and the virion spike (S) glycoprotein, was also well studied. S glycoprotein includes two subunits, S1 and S2, resulting from cleavage of the one precursor into two parts. S1 determines the virus host range and cellular tropism with the key functional domain Receptor Binding Domain (RBD), while S2 contains two tandem domains, heptad repeats 1 (HR1) and Heptad Repeats 2 (HR2), to mediate virus-cell membrane fusion. It is believed that the fusion process is similar to that of HIV-1; for example, when S1 binds to the receptor on the cell membrane, the fusion peptide at the N terminus of S2 inserts into the cell membrane, then three HR1s attach to each other in parallel as a trimer, followed by binding of three HR2s separately onto the outside of the trimer to form a 6-helix bundle, thus bringing virus and cell membranes close to each other to trigger fusion. As the major vaccine target, the S protein has been evaluated in different types of vaccines against infection by CoVs [23-27].

Apart from the inactive whole virus particle, live attenuated virus with gene deletion, four more vaccines which mainly contain S protein were studied. These include a virus-like particle which incorporated S protein into hepatitis virus or influenza virus protein; virus vectors, such as Modified Vaccinia Virus Ankara (MVA) or Adenovirus carrying S protein; S protein subunit vaccine, like RBD-based protein; and DNA vaccine which encodes the full length or part of the S protein gene [28-36,27]. Most of them have been tested in mouse models and showed the ability to elicit neutralizing antibodies. The first SARS-CoV DNA vaccine was tested in humans only 19 months after the

virus sequence was published, while the DNA vaccine GLS-5300, the first MERS-CoV vaccine, went to clinical trials in 2016 [37]. In addition to these conventional vaccines, Liu et al. analyzed the T cell epitopes of SARS-CoV and MERS-CoV, revealed the potential cross-reactivity of the coronaviruses, and assessed the possibility of developing universal vaccines against coronavirus infections [38].

Most CoVs share a similar viral structure, similar infection pathway, and a similar structure of the S proteins, suggesting that similar research strategies should also be applicable for the 2019-nCoV. For example, the study of MERS-CoV vaccines was accelerated by virtue of strategies that had been established for SARS-CoV [39,40]. Therefore, to predict whether vaccines developed for SARS-CoV will also be effective against 2019-nCoV infection, the full-length S protein sequences from the 2019-nCoV, a SARS-CoV, and two genetically similar bat CoV strains were selected for alignment. The results indicated more than 50% homology of the viruses. However, the most variable residues are located in S1, a critical vaccine target, implying that neutralizing antibodies that were so effective against SARS-CoV infection may fail to recognize the 2019-nCoV, and that multiple amino acid differences at the receptor binding motif may modify virus tropism, a possible reason for cross-species transmission. However, several bottlenecks typically delay the approval of vaccines to prevent CoVs infection. Firstly, lack of proper animal models for evaluating vaccine efficacy. Second, there are limitations from the S protein itself, such as mutations in the neutralization antibody epitopes in S protein that can cause virus escape, or non-neutralization antibody epitopes in vaccines that may elicit antibody-mediated disease enhancement (ADE) [41,42]. Third, DNA vaccines may recombine with other viruses. Fourth, pre-existing immunity may eliminate the vaccine by removing the general human virus vectors (Table 2) [43].

Table 2: Neutralizing monoclonal antibodies targeting MERS-CoV and their mechanism of action.

Monoclonal antibody	Antibody mechanism of action	Authors
MERS-4	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 5$-$\beta 6$, $\beta 6$-$\beta 7$ and $\beta 7$-$\beta 8$ loops on the receptor-binding subdomain in RBD of MERS-CoV with no overlap DPP4 binding surface. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i> by inducing $\beta 5$-$\beta 6$ shallow groove on the RBD. 	Yi Y, et al. Ying T, et al. [102,103].
MERS-27	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 6$-$\beta 7$ loop and $\beta 7$ strand on RBD of MERS-CoV and overlap with the DPP4 binding surface. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i>. 	Yi Y, et al. Yu X, et al. [102,104].
4C2	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 6$-$\beta 7$ loop and $\beta 7$ strand on RBD of MERS-CoV and overlap with the DPP4 binding surface. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i> and <i>in vivo</i> (Mouse). 	Zhang S, et al. Li Y, et al. [105,106].
m336	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 5$-$\beta 8$ strands, $\beta 5$-$\beta 6$ loop and $\beta 6$-$\beta 7$ loop in RBD of MERS-CoV and overlap with the DPP4 binding surface. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 by mimicking the interaction between RBD and DPP4 in the similar binding angle <i>in vitro</i> and <i>in vivo</i> (Mouse and rabbit). 	Doremalen NV, et al. Ying T, et al. [107,108].
G4	<ul style="list-style-type: none"> • Binding to the glycosylated surface on the S2 subunit protein <i>in vitro</i>. 	Wang L, et al. Pallesen J, et al. [109,110].

D12	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 6$-$\beta 7$ loop and $\beta 7$ strand on RBD of MERS-CoV and overlap with the DPP4 binding surface. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i>. 	Wang L, et al. Pallesen J, et al. [41,110].
JC57-14	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 6$-$\beta 7$ loop and $\beta 7$ strand on RBD of MERS-CoV and overlap with the DPP4 binding surface. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i>. 	Wang L, et al. [41].
MERS-GD27	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 5$-$\beta 8$ strands, $\beta 5$-$\beta 6$ loop and $\beta 6$-$\beta 7$ loop in RBD of MERS-CoV. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 by mimicking the interaction between RBD and DPP4 in the same binding angle <i>in vitro</i> and <i>in vivo</i> (Mice). 	Niu P, et al. [111].
MERS-GD33	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 5$-$\beta 8$ strands, $\beta 5$-$\beta 6$ loop and $\beta 6$-$\beta 7$ loop in RBD of MERS-CoV. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 mimicking the interaction between RABD and DPP4 in the same binding angle <i>in vitro</i>. 	Niu P, et al. [112].
LCA60	<ul style="list-style-type: none"> • Binding to the C-terminal segment of the $\beta 8$ strand, $\beta 6$-$\beta 9$ loop, and $\beta 6$-$\beta 8$ loop on RBD of MERS-CoV. • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i>. 	Walls AC, et al. [101].
MCA1	<ul style="list-style-type: none"> • Binding to RBD with 6 complementarity-determining regions • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i> and <i>in vivo</i> (Mouse). 	Wit ED, et al. [113].
CDC2-C2	<ul style="list-style-type: none"> • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i> and <i>in vivo</i> (Mouse). 	Wang L, et al. [41].
7D10	<ul style="list-style-type: none"> • Binding to N-terminal domain of S protein of MERS-CoV • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i> and <i>in vivo</i> (Mouse). 	Niu P, et al. [111].
G2	<ul style="list-style-type: none"> • Binding to N-terminal domain of S protein of MERS-CoV • Blocking the interaction of S1 subunit protein with cellular receptor DPP4 <i>in vitro</i>. 	Zhou H, et al. [114].

Finally, there is the problem of return on investment which may be slow and, hence, inhibit investments and slow down the clinical study. Jiang and colleagues have demonstrated that RBD in the SARS-CoV-S protein is the major target of neutralizing antibodies in SARS patients and is able to induce highly potent neutralizing antibody responses and long-term protective immunity in animal models. It contains 6 different conformational neutralizing epitopes, to which a series of mouse monoclonal antibodies (mAbs) with different neutralizing activity were generated. Interestingly, these mAbs exhibited cross-neutralizing activities against divergent SARS-CoV strains isolated from SARS patients at different stages of SARS epidemics and those from palm civets [44,45]. This group has also shown that these SARS-CoV-RBD specific neutralizing mAbs can cross-neutralize bat SL-CoVs, such as bat SL-CoV-W1V1, indicating that these antibodies may also cross-neutralize 2019-nCoV. Most importantly, RBD-based vaccine could induce neutralizing antibody responses and protection against SARS-CoV infection in the immunized animals, while it did not elicit ADE or other harmful immune responses, unlike the virus inactivated vaccines or full-length S protein-based vaccines as discussed above. Therefore, this RBD-based SARS vaccine is expected to be safer and more effective than the vaccines targeting other sites in S protein. Jiang and Du's

groups have collaborated with Hotez's group at Baylor College of Medicine in Houston and Tseng's group at the University of Texas Medical Branch at Galveston, Texas, USA in development of an effective and safe vaccine at the late stage of preclinical study [46,47].

CoV fusion or entry inhibitors

Based on the previous experience in developing the HIV-1 fusion inhibitor SJ-2176, Jiang et al. [48]. discovered the first anti-SARS-CoV peptide (SC-1) from the HR2 domain of SARS-CoV S protein S2 subunit. SC-1 could bind onto the HR1 domain to form a six-helical bundle (6-HB), blocking S protein-mediated membrane fusion and inhibiting SARS-CoV infection [49]. When MERS-CoV was circulating in human populations in 2012, following similar mechanistic design, Jiang's research group developed another peptide, designated HR2P, which was derived from the virus HR2 region as well and effectively inhibited MERS-CoV infection [50]. The further modified version of HR2P, HR2PM2, presented even better anti-MERS-CoV activity and pharmaceutical properties. Development of broad-spectrum pan-CoV fusion inhibitors would be an ideal way to cope with epidemics or pandemics caused by emerging HCoVs. The conservative amino acid sequence of the HR1 region across

different CoVs has the potential to be a target domain for development of an inhibitor. Continuing to work on the HR1 and HR2 domains, Jiang's group discovered that the peptide OC43-HR2P, derived from the HR2 domain of HCoV-OC43, broadly inhibited fusion by multiple HCoVs. By optimization of this peptide, a pan-CoV fusion inhibitor, EK1, was generated. It could form a stable six-helix bundle (6-HB) structure with HR1s and showed significantly improved fusion-inhibitory activity and pharmaceutical properties [51]. The alignment of S protein exhibited 100% identity at the HR2 domains between the 2019-nCoV and SARS-CoV; however, they found 7 amino acid changes in the fusion core of the HR1, located in the EK1 binding motif. Fortunately, the substitutions were conservative replacements which would not dramatically disrupt the interactions between EK1 and HR1, meaning that EK1 would still have the potential to be an effective inhibitor for 2019-nCoV infection. The table shows selected patents associated with the aforementioned potential drugs, together with patents disclosing small molecules for treatment of SARS or MERS (Table 3).

CoV s-rbd-specific neutralizing antibodies

So far, most neutralizing antibodies recognize the RBD in the S protein S2 of CoVs. Compared with the high mutation rate in the S1 protein, S2 is much more conservative, thereby decreasing the off target risk caused by amino acid replacement, and also bypassing the special epitopes that may cause ADE [52,53]. This means that the cocktail of monoclonal antibodies binding to different epitopes of RBD would be more desirable for therapeutic purposes [54]. For treatment, the monoclonal antibodies are from a human source or are humanized antibodies, isolated or generated with various approaches. For example, wild-type mice were immunized with soluble recombinant RBD containing the S protein. Then mouse antibodies were humanized and isolated, or transgenic mice were directly immunized, to express human versions of the antibodies [55,56]. However, direct cloning of single B cells from human survivors, used in combination with the phage-display antibody library, could provide authentic human antibodies. Until now, it should be noted that many

Table 3: Selected patents associated with potential drugs (Repurposing) for COVID-19 or small molecules for treatment of SARS or MERS.

Patient Number	Title	Organization
WO2009114512	Preparation of azetidine and cyclobutane derivatives as JAK inhibitors	Incyte Corporation, USA
WO2014028756	Deuterated baricitinib	Concert Pharmaceuticals, Inc., USA
JP5971830	Preparation of polycyclic pyridone derivatives as cap-dependent endonuclease (CEN) inhibitors and prodrugs thereof	Shionogi and Co., Ltd., Japan
US20160122374	Preparation of nucleosides and methods for treating Filoviridae virus infections	Gilead Sciences, Inc., USA
US20170071964	Preparation of amino acid-containing nucleotides and methods for treating arenaviridae and coronaviridae virus infections	Gilead Sciences, Inc., USA
WO2007075145	Preparation of benzopyranone derivatives as anti-coronaviral agents Singapore Polytechnic,	Singapore; Shanghai Institute of Materia Medica Chinese Academy of Sciences, China
WO2005021518	Preparation of 3,4-dihydro-2H-1,4-benzoxazine-2-carboxylic acid derivatives as cysLT2 receptor antagonists for treatment of respiratory diseases	Ono Pharmaceutical Co., Ltd., Japan
WO2007120160	Preparation of N-heterocyclic acetamides useful for viral inhibition	Novartis AG, USA
WO2009119167	Aniline derivative having anti-RNA viral activity	KinoPharma, Inc., Japan
WO2013049382	Broad-spectrum antivirals against 3c or 3c-like proteases of picornavirus-like supercluster: picornaviruses, caliciviruses and coronaviruses	Kansas State University Research Foundation; The Ohio State University; Wichita State University-all in USA
WO2018042343	Preparation of peptides that inhibit 3C and 3CL proteases and methods of use thereof	GlaxoSmithKline, UK
WO2007067515	Five-membered iminocyclitol derivatives as selective and potent glycosidase inhibitors: new structures for antivirals and osteoarthritis therapeutics	Academia Sinica, Taiwan

neutralizing antibodies have been successfully discovered for treatment of SARS-CoV and MERS-CoV infection.

These antibodies have all been described favorably in the literature [57]. A similar approach is known as single chain fragment variable (scFv) library screening, whereby the use of RBD as a bait protein allows some neutralizing antibodies to be screened out from non-immune humans [58,59]. Antibodies effective at inhibiting SARS-CoV infection should also have the potential for treatment of 2019-nCoV as well, as long as the binding motif in RBD shares the same sequences. The new neutralizing monoclonal antibodies would also be isolated from the patients using the established techniques. Hence the therapies for SARS-CoV can be extrapolated to use for SARS-CoV-2. The specific neutralizing monoclonal antibodies either against Receptor Binding Domain (RBD) in spike protein or specific antibody that binds to ACE2 could effectively block the virus entry.

CoV replication inhibitors

Similar to developing vaccines, drugs effective against other RNA viruses were also repurposed for CoVs. Two major types of drugs being nucleoside analogues and immunomodulators. So far, the most common therapies tried in patients with CoVs are ribavirin, lopinavir/ritonavir, IFN, or their combinations [60]. Despite the antiviral activity observed with *in vitro* studies, the clinical effect was not consistent, in that ribavirin does not prolong the survival of SARS-CoV patients, while lopinavir/ritonavir plus ribavirin seemed to improve clinical outcomes for SARS patients, but the improvement was not confirmed in MERS-CoV patients [61-63]. IFNs showed effective at inducing antiviral activity against both SARS-CoV and MERS-CoV, but without significant improvement in the outcomes for the patients [64,65]. In addition to the drug regimens used in patients, numerous drugs developed for the treatment of infection with CoVs were thoroughly discussed in the literature [66]. However, replication of an RNA virus usually generates progeny viruses with a highly diverse genome. Recombination also easily takes place between viral genomes, and these gene level changes may result in drug resistance if the mutations affect the drug target domain [67]. Development of drugs is also hampered by various evaluation methods and animal models used for testing drug activity among different labs worldwide, which could postpone selection of the best drug for clinical trials.

Angiotensin receptor blockers

Angiotensin Receptor Blockers (ARBs) have been reported to associate with viral infection, including HCoV [68,69]. Irbesartan, a typical ARB, was approved by the FDA for treatment of hypertension and diabetic nephropathy. Here, network proximity analysis shows a significant association between irbesartan's targets and HCoV-associated host proteins in the human interactome. Irbesartan targets SLC10A1, encoding the sodium/bile acid cotransporter (NTCP) protein that has been identified as a functional pre-S1-specific receptor for the Hepatitis B Virus (HBV) and the Hepatitis Delta Virus (HDV). Irbesartan

can inhibit NTCP, thus inhibiting viral entry [70,71]. SLC10A1 interacts with C11orf74, a potential transcriptional repressor that interacts with nsp-10 of SARS-CoV [72]. There are several other ARBs (such as *Eletriptan*, *Frovatriptan*, and *Zolmitriptan*) in which their targets are potentially associated with HCoV-associated host proteins in the human interactome.

Anti-inflammatory agents

Inflammatory pathways play essential roles in viral infections [73]. As a biogenic amine, melatonin (N-acetyl5-methoxytryptamine) plays a key role in various biological processes, and offers a potential strategy in the management of viral infections [74,75]. Viral infections are often associated with immune-inflammatory injury, in which the level of oxidative stress increases significantly and leaves negative effects on the function of multiple organs [76]. The antioxidant effect of melatonin makes it a putative candidate drug to relieve patients' clinical symptoms in antiviral treatment, even though melatonin cannot eradicate or even curb the viral replication or transcription [77]. In addition, the application of melatonin may prolong patients' survival time, which may provide a chance for patients' immune systems to recover and eventually eradicate the virus. Melatonin indirectly targets several HCoV cellular targets, including ACE2, BCL2L1, JUN, and IKBKB. Eplerenone, an aldosterone receptor antagonist, is reported to have a similar anti-inflammatory effect as melatonin. By inhibiting mast-cell-derived proteinases and suppressing fibrosis, eplerenone can improve survival of mice infected with encephalomyocarditis virus [78]. In summary, our network proximity analyses offer multiple candidates repurposable drugs that target diverse cellular pathways for potential prevention and treatment of 2019-nCoV/SARS-CoV-2. However, further preclinical experiments, and clinical trials are required to verify the clinical benefits of these network-predicted candidates before clinical use [21].

Toremifene plus emodin

Toremifene is among the approved first-generation nonsteroidal SERMs for the treatment of metastatic breast cancer [79]. SERMs (including toremifene) inhibited Ebola virus infection, by interacting with and destabilizing the Ebola virus glycoprotein [80,81]. *In vitro* assays have demonstrated that toremifene inhibited growth of MERSCoV, and SARA-CoV [82,83]. Emodin, an anthraquinone derivative extracted from the roots of *rheum tanguticum*, has been reported to have various anti-virus effects. Specifically, emodin inhibited SARSCoV-associated 3a protein, and blocked an interaction between the SARS-CoV spike protein and ACE2 [84,85]. Altogether, network analyses and published experimental data suggested that combining toremifene and emodin offered a potential therapeutic approach for 2019-nCoV/ SARS-CoV-2.

DISCUSSION

Mercaptopurine plus melatonin

Targets of both mercaptopurine and melatonin showed strong

network proximity with HCoV associated host proteins in the human interactome network. Recent *in vitro* and *in vivo* studies identified mercaptopurine as a selective inhibitor of both SARS-CoV and MERS-CoV by targeting papain like protease [86,87]. Melatonin was reported in potential antiviral infection via its anti-inflammatory and antioxidant effects [74,77]. Melatonin indirectly regulates ACE2 expression, a key entry receptor involved in viral infection of HCoVs, including 2019-nCoV/SARS-CoV-2 [1]. Specifically, melatonin was reported to inhibit calmodulin and calmodulin interacts with ACE2 by inhibiting shedding of its ectodomain, a key infectious process of SARS-CoV [88,89]. JUN, also known as c-Jun, is a key host protein involving in HCoV infectious bronchitis virus [90-114]. Mercaptopurine and melatonin may synergistically block c-Jun signaling by targeting multiple cellular targets.

CONCLUSION

In conclusion, combination of mercaptopurine and melatonin may offer a potential combination therapy for 2019nCoV/SARS-CoV-2 by synergistically targeting papainlike protease, ACE2, c-Jun signaling, and anti-inflammatory pathways. However, further experimental observations on ACE2 pathways by melatonin in 2019-nCoV/SARS-CoV-2 are highly warranted.

FUNDING

Not applicable.

ACKNOWLEDGMENT

I would like to acknowledge University of Gondar.

CONFLICT OF INTEREST

The authors declare that they have no competing interests.

REFERENCES

- Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579(1):270-273.
- Wu JT, Leung K, Leung GM. Nowcasting and forecasting the potential domestic and international spread of the 2019-nCoV outbreak originating in wuhan, china: a modelling study. *The Lancet*. 2020;395(10225):689-697.
- The LID. Challenges of coronavirus disease 2019. *Lancet Infect Dis*. 2020;20(3):261.
- Burki T. Outbreak of coronavirus disease 2019. *Lancet Infect Dis*. 2020;20(3):292-293.
- Vickers NJ. Animal communication: when i'm calling you, will you answer too? *Current Biology*. 2017;27(14):R713-R715.
- Zhu Rf, Gao Rl, Robert SH, Gao JP, Yang SG, Zhu C. Systematic review of the registered clinical trials of coronavirus diseases 2019 (COVID-19). *Med Rxiv*. 2020.
- Anthony SJ, Johnson CK, Greig DJ, Kramer S, Che X, Wells H, et al. Global patterns in coronavirus diversity. *Virus Evol*. 2017;3(1).
- Su S, Wong G, Shi W, Liu J, Lai AC, Zhou J, et al. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. *Trends Microbiol*. 2016;24(6):490-502.
- Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in china, 2019. *N Engl J Med*. 2020;382(8):727-733.
- Tang B, Bragazzi NL, Li Q, Tang S, Xiao Y, Wu J. An updated estimation of the risk of transmission of the novel coronavirus (2019-nCoV). *Infect Dis Model*. 2020;5(1):248-255.
- Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in wuhan, china. *JAMA*. 2020;323(11):1061-1069.
- Waters E, Doyle J. Systematic reviews of public health in developing countries are in train. *Bmj*. 2004;328(7439):585.
- Liu C, Zhou Q, Li Y, Garner LV, Watkins SP, Carter LJ, et al. Research and development on therapeutic agents and vaccines for covid-19 and related human coronavirus diseases. *ACS Cent Sci*. 2020;6(3):315-331.
- Kampen KRV, Shi Z, Gao P, Zhang J, Foster KW, Chen DT, et al. Safety and immunogenicity of adenovirus-vectored nasal and epicutaneous influenza vaccines in humans. *Vaccine*. 2005;23(8):1029-1036.
- Lu H. Drug treatment options for the 2019-new coronavirus (2019-nCoV). *Biosci Trends*. 2020;14(1):69-71.
- Kumar V, Jung YS, Liang PH. Anti-SARS coronavirus agents: a patent review (2008-present). *Expert Opin Ther Pat*. 2013;23(10):1337-1348.
- Richardson P, Griffin I, Tucker C, Smith D, Oechsle O, Phelan A, et al. Baricitinib as potential treatment for 2019-nCoV acute respiratory disease. *The Lancet*. 2020;395(10223):e30-e31.
- Qiu R, Wei X, Zhao M, Zhong C, Zhao C, Hu J, et al. Outcome reporting from protocols of clinical trials of Coronavirus Disease 2019 (COVID-19): a review. *Med Rxiv*. 2020.
- Maxmen A. More than 80 clinical trials launch to test coronavirus treatments. *Nature*. 2020;578(7795):347-348.
- Mifsud EJ, Hayden FG, Hurt AC. Antivirals targeting the polymerase complex of influenza viruses. *Antiviral Res*. 2019;169(1):104545.
- Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) *in vitro*. *Cell Res*. 2020;30(3):269-271.
- Wang Z, Chen X, Lu Y, Chen F, Zhang W. Clinical characteristics and therapeutic procedure for four cases with 2019 novel coronavirus pneumonia receiving combined chinese and western medicine treatment. *BioSci Trends*. 2020;14(1):64-68.
- Raj VS, Mou H, Smits SL, Dekkers DH, Müller MA, Dijkman R, et al. Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC. *Nature*. 2013;495(7440):251-254.
- Li W, Moore MJ, Vasilieva N, Sui J, Wong SK, Berne MA, et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature*. 2003;426(6965):450-454.
- Zhang N, Jiang S, Du L. Current advancements and potential strategies in the development of MERS-CoV vaccines. *Expert Rev Vaccines*. 2014;13(6):761-774.

26. Wang X, Xiong W, Ma X, Wei M, Chen Y, Lu L, et al. The conserved residue Arg46 in the N-terminal heptad repeat domain of HIV-1 gp41 is critical for viral fusion and entry. *Plos One*. 2012;7(9).
27. Zhou Y, Yang Y, Huang J, Jiang S, Du L. Advances in MERS-CoV vaccines and therapeutics based on the receptor-binding domain. *Viruses*. 2019;11(1):60.
28. Agrawal AS, Tao X, Algaissi A, Garron T, Narayanan K, Peng BH, et al. Immunization with inactivated middle east respiratory syndrome coronavirus vaccine leads to lung immunopathology on challenge with live virus. *Hum Vaccin Immunother*. 2016;12(9):2351-2356.
29. Fett C, DeDiego ML, Regla-Nava JA, Enjuanes L, Perlman S. Complete protection against severe acute respiratory syndrome coronavirus-mediated lethal respiratory disease in aged mice by immunization with a mouse-adapted virus lacking e protein. *J Virol*. 2013;87(12):6551-6559.
30. Lokugamage KG, Yoshikawa-Iwata N, Ito N, Watts DM, Wyde PR, Wang N, et al. Chimeric coronavirus-like particles carrying severe acute respiratory syndrome coronavirus (SCoV) S protein protect mice against challenge with SCoV. *Vaccine*. 2008;26(6):797-808.
31. Liu YV, Massare MJ, Barnard DL, Kort T, Nathan M, Wang L, et al. Chimeric severe acute respiratory syndrome coronavirus (SARS-CoV) S glycoprotein and influenza matrix 1 efficiently form virus-like particles (VLPs) that protect mice against challenge with SARS-CoV. *Vaccine*. 2011;29(38):6606-6613.
32. Volz A, Kupke A, Song F, Jany S, Fux R, Shams-Eldin H, et al. Protective efficacy of recombinant modified vaccinia virus Ankara delivering Middle East respiratory syndrome coronavirus spike glycoprotein. *J Virol*. 2015;89(16):8651-8656.
33. Munster VJ, Wells D, Lambe T, Wright D, Fischer RJ, Bushmaker T, et al. Protective efficacy of a novel simian adenovirus vaccine against lethal MERS-CoV challenge in a transgenic human DPP4 mouse model. *npj Vaccines*. 2017;2(1):1-4.
34. Du L, Ma C, Jiang S. Antibodies induced by receptor-binding domain in spike protein of SARS-CoV do not cross-neutralize the novel human coronavirus hCoV-EMC. *J Infect*. 2013;67(4):348-350.
35. Chi H, Zheng X, Wang X, Wang C, Wang H, Gai W, et al. DNA vaccine encoding Middle East respiratory syndrome coronavirus S1 protein induces protective immune responses in mice. *Vaccine*. 2017;35(16):2069-2075.
36. Martin JE, Louder MK, Holman LA, Gordon IJ, Enama ME, Larkin BD, et al. A SARS DNA vaccine induces neutralizing antibody and cellular immune responses in healthy adults in a Phase I clinical trial. *Vaccine*. 2008;26(50):6338-6343.
37. Modjarrad K, Roberts CC, Mills KT, Castellano AR, Paolino K, Muthumani K, et al. Safety and immunogenicity of an anti-Middle East respiratory syndrome coronavirus DNA vaccine: a phase 1, open-label, single-arm, dose-escalation trial. *Lancet Infect Dis*. 2019;19(9):1013-1022.
38. Liu WJ, Zhao M, Liu K, Xu K, Wong G, Tan W, et al. T-cell immunity of SARS-CoV: Implications for vaccine development against MERS-CoV. *Antiviral Res*. 2017;137:82-92.
39. Yuan Y, Cao D, Zhang Y, Ma J, Qi J, Wang Q, et al. Cryo-EM structures of MERS-CoV and SARS-CoV spike glycoproteins reveal the dynamic receptor binding domains. *Nat Commun*. 2017;8(1):15092.
40. Du L, Jiang S. Middle East respiratory syndrome: current status and future prospects for vaccine development. *Expert Opin Biol Ther*. 2015; 15(11):1647-1651.
41. Wang L, Shi W, Chappell JD, Joyce MG, Zhang Y, Kanekiyo M, et al. Importance of neutralizing monoclonal antibodies targeting multiple antigenic sites on the middle east respiratory syndrome coronavirus spike glycoprotein to avoid neutralization escape. *J Virol*. 2018;92(10):e02002-e02017.
42. Olsen CW. A review of feline infectious peritonitis virus: molecular biology, immunopathogenesis, clinical aspects, and vaccination. *Vet Microbiol*. 1993;36(1):1-37.
43. Long BR, Sandza K, Holcomb J, Crockett L, Hayes GM, Arens J, et al. The impact of pre-existing immunity on the non-clinical pharmacodynamics of AAV5-based gene therapy. *Mol Ther-Meth Clin D*. 2019;13(1):440-452.
44. He Y, Zhou Y, Liu S, Kou Z, Li W, Farzan M, et al. Receptor-binding domain of SARS-CoV spike protein induces highly potent neutralizing antibodies: implication for developing subunit vaccine. *Biochem Biophys Res Commun*. 2004;324(2):773-781.
45. He Y, Li J, Li W, Lustigman S, Farzan M, Jiang S. Cross-neutralization of human and palm civet severe acute respiratory syndrome coronaviruses by antibodies targeting the receptor-binding domain of spike protein. *J Immunol*. 2006;176(10):6085-6092.
46. Zeng LP, Ge XY, Peng C, Tai W, Jiang S, Du L, et al. Cross-neutralization of SARS coronavirus-specific antibodies against bat SARS-like coronaviruses. *Sci China Life Sci*. 2017;60(12):1399-1402.
47. Chen WH, Du L, Chag SM, Ma C, Tricoche N, Tao X, et al. Yeast-expressed recombinant protein of the receptor-binding domain in SARS-CoV spike protein with deglycosylated forms as a SARS vaccine candidate. *Hum Vaccin Immunother*. 2014;10(3):648-658.
48. Jiang S, Lin K, Strick N, Neurath AR. HIV-1 inhibition by a peptide. *Nature*. 1993;365(6442):113.
49. Liu S, Xiao G, Chen Y, He Y, Niu J, Escalante CR, et al. Interaction between heptad repeat 1 and 2 regions in spike protein of SARS-associated coronavirus: implications for virus fusogenic mechanism and identification of fusion inhibitors. *Lancet*. 2004;363(9413):938-947.
50. Lu L, Liu Q, Zhu Y, Chan K-H, Qin L, Li Y, et al. Structure-based discovery of Middle East respiratory syndrome coronavirus fusion inhibitor. *Nat Commun*. 2014;5(1):1-12.
51. Xia S, Yan L, Xu W, Agrawal AS, Algaissi A, Tseng C-TK, et al. A pan-coronavirus fusion inhibitor targeting the HR1 domain of human coronavirus spike. *Sci Adv*. 2019;5(4):eaav4580.
52. Buchholz UJ, Bukreyev A, Yang L, Lamirande EW, Murphy BR, Subbarao K, et al. Contributions of the structural proteins of severe acute respiratory syndrome coronavirus to protective immunity. *Proc Natl Acad Sci U S A*. 2004;101(26):9804-9809.
53. Olsen CW, Corapi WV, Jacobson RH, Simkins RA, Saif LJ, Scott FW. Identification of antigenic sites mediating antibody-dependent enhancement of feline infectious peritonitis virus infectivity. *J Gen Virol*. 1993;74(4):745-749.
54. Bakker AB, Marissen WE, Kramer RA, Rice AB, Weldon WC, Niezgoda M, et al. Novel human monoclonal antibody combination effectively neutralizing natural rabies virus variants and individual *in vitro* escape mutants. *J Virol*. 2005;79(14):9062-9068.

55. Qiu H, Sun S, Xiao H, Feng J, Guo Y, Tai W, et al. Single-dose treatment with a humanized neutralizing antibody affords full protection of a human transgenic mouse model from lethal middle east respiratory syndrome (MERS)-coronavirus infection. *Antiviral Res.* 2016;132:141-148.
56. Pascal KE, Coleman CM, Mujica AO, Kamat V, Badithe A, Fairhurst J, et al. Pre-and postexposure efficacy of fully human antibodies against spike protein in a novel humanized mouse model of MERS-CoV infection. *Proc Natl Acad Sci U S A.* 2015;112(28):8738-8743.
57. Coughlin MM, Prabhakar BS. Neutralizing human monoclonal antibodies to severe acute respiratory syndrome coronavirus: target, mechanism of action, and therapeutic potential. *Rev Med Virol.* 2012;22(1):2-17.
58. Tang X-C, Agnihothram SS, Jiao Y, Stanhope J, Graham RL, Peterson EC, et al. Identification of human neutralizing antibodies against MERS-CoV and their role in virus adaptive evolution. *Proc Natl Acad Sci U S A.* 2014;111(19):E2018-E2026.
59. Sui J, Aird DR, Tamin A, Murakami A, Yan M, Yammanuru A, et al. Broadening of neutralization activity to directly block a dominant antibody-driven SARS-coronavirus evolution pathway. *Plos Pathog.* 2008;4(11): e1000197.
60. Omrani AS, Saad MM, Baig K, Bahloul A, Abdul-Matin M, Alaidaroos AY, et al. Ribavirin and interferon alfa-2a for severe Middle East respiratory syndrome coronavirus infection: a retrospective cohort study. *Lancet Infect Dis.* 2014;14(11):1090-1095.
61. Sheahan TP, Sims AC, Graham RL, Menachery VD, Gralinski LE, Case JB, et al. Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses. *Sci Transl Med.* 2017;9(396).
62. Gross AE, Bryson ML. Oral ribavirin for the treatment of noninfluenza respiratory viral infections: a systematic review. *Ann Pharmacother.* 2015;49(10):1125-1135.
63. Chu C, Cheng V, Hung I, Wong M, Chan K, Chan K, et al. Role of lopinavir/ritonavir in the treatment of SARS: initial virological and clinical findings. *Thorax.* 2004;59(3):252-256.
64. Chan JF, Chan KH, Kao RY, To KK, Zheng BJ, Li CP, et al. Broad-spectrum antivirals for the emerging middle east respiratory syndrome coronavirus. *J Infect.* 2013;67(6):606-616.
65. Loutfy MR, Blatt LM, Siminovitch KA, Ward S, Wolff B, Lho H, et al. Interferon alfacon-1 plus corticosteroids in severe acute respiratory syndrome: a preliminary study. *JAMA.* 2003;290(24):3222-3228.
66. Dyall J, Gross R, Kindrachuk J, Johnson RF, Olinger GG, Hensley LE, et al. Middle East respiratory syndrome and severe acute respiratory syndrome: current therapeutic options and potential targets for novel therapies. *Drugs.* 2017;77(18):1935-1966.
67. Belshaw R, Gardner A, Rambaut A, Pybus OG. Pacing a small cage: mutation and RNA viruses. *Trends Ecol Evol.* 2008;23(4):188-193.
68. Moskowitz DW, Johnson FE. The central role of angiotensin I-converting enzyme in vertebrate pathophysiology. *Curr Top Med Chem.* 2004;4(13):1431-1452.
69. Erlandson KM, Kitch D, Wester CW, Kalayjian RC, Overton ET, Castillo-Mancilla J, et al. The impact of statin and angiotensin-converting enzyme inhibitor/angiotensin receptor blocker therapy on cognitive function in adults with human immunodeficiency virus infection. *Clin Infect Dis.* 2017;65(12):2042-2049.
70. Wang X-j, Hu W, Zhang T-y, Mao Y-y, Liu N-n, Wang S-q. Irbesartan, an FDA approved drug for hypertension and diabetic nephropathy, is a potent inhibitor for hepatitis B virus entry by disturbing Na(+)-dependent taurocholate cotransporting polypeptide activity. *Antiviral Res.* 2015;120(1):140-146.
71. Ko C, Park WJ, Park S, Kim S, Windisch MP, Ryu WS. The FDA-approved drug irbesartan inhibits HBV-infection in HepG2 cells stably expressing sodium taurocholate co-transporting polypeptide. *Antivir Ther.* 2015;20(8):835-842.
72. Hong M, Li W, Wang L, Jiang L, Liu L, Zhao H, et al. Identification of a novel transcriptional repressor (HEPIS) that interacts with nsp-10 of SARS coronavirus. *Viral Immunol.* 2008;21(2):153-162.
73. Rainsford K. Influenza ("Bird Flu"), inflammation and anti-inflammatory/analgesic drugs. *Inflammopharmacology.* 2006;14(2):2-9.
74. Silvestri M, Rossi GA. Melatonin: its possible role in the management of viral infections-a brief review. *Ital J Pediatr.* 2013;39(1):61.
75. Srinivasan V, Mohamed M, Kato H. Melatonin in bacterial and viral infections with focus on sepsis: a review. *Recent Pat Endocr Metab Immune Drug Discov.* 2012;6(1):30-39.
76. Tan DX, Korkmaz A, Reiter RJ, Manchester LC. Ebola virus disease: potential use of melatonin as a treatment. *J Pineal Res.* 2014;57(4):381-384.
77. Galano A, Tan DX, Reiter RJ. On the free radical scavenging activities of melatonin's metabolites, AFMK and AMK. *J Pineal Res.* 2013;54(3):245-257.
78. Xiao J, Shimada M, Liu W, Hu D, Matsumori A. Antinflammatory effects of eplerenone on viral myocarditis. *Eur J Heart Fail.* 2009;11(4):349-353.
79. Zhou W-B, Ding Q, Chen L, Liu X-A, Wang S. Toremifene is an effective and safe alternative to tamoxifen in adjuvant endocrine therapy for breast cancer: results of four randomized trials. *Breast Cancer Res Treat.* 2011;128(3):625-631.
80. Johansen LM, Brannan JM, Delos SE, Shoemaker CJ, Stossel A, Lear C, et al. FDA-approved selective estrogen receptor modulators inhibit ebola virus infection. *Sci Transl Med.* 2013;5(190):79.
81. Zhao Y, Ren J, Harlos K, Jones DM, Zeltina A, Bowden TA, et al. Toremifene interacts with and destabilizes the ebola virus glycoprotein. *Nature.* 2016;535(7610):169-172.
82. Cong Y, Hart BJ, Gross R, Zhou H, Frieman M, Bollinger L, et al. MERS-CoV pathogenesis and antiviral efficacy of licensed drugs in human monocyte-derived antigen-presenting cells. *Plos One.* 2018;13(3): e0194868.
83. De Wilde AH, Jochmans D, Posthuma CC, Zevenhoven-Dobbe JC, Van Nieuwkoop S, Bestebroer TM, et al. Screening of an FDA-approved compound library identifies four small-molecule inhibitors of middle east respiratory syndrome coronavirus replication in cell culture. *Antimicrob Agents Chemother.* 2014;58(8):4875-4884.
84. Schwarz S, Wang K, Yu W, Sun B, Schwarz W. Emodin inhibits current through SARS-associated coronavirus 3a protein. *Antiviral Res.* 2011;90(1):64-69.
85. Ho TY, Wu SL, Chen JC, Li CC, Hsiang CY. Emodin blocks the SARS coronavirus spike protein and angiotensin-converting enzyme 2 interaction. *Antiviral Res.* 2007;74(2):92-101.

86. Chen X, Chou CY, Chang GG. Thiopurine analogue inhibitors of severe acute respiratory syndrome-coronavirus papain-like protease, a deubiquitinating and deisgylating enzyme. *Antiviral Chem Chemother.* 2009;19(4):151-156.
87. Cheng KW, Cheng SC, Chen WY, Lin MH, Chuang SJ, Cheng IH, et al. Thiopurine analogs and mycophenolic acid synergistically inhibit the papain-like protease of middle east respiratory syndrome coronavirus. *Antiviral Res.* 2015;115(1):9-16.
88. Lambert DW, Clarke NE, Hooper NM, Turner AJ. Calmodulin interacts with angiotensin converting enzyme2 (ACE2) and inhibits shedding of its ectodomain. *FEBS Lett.* 2008;582(2):385-390.
89. Dai J, Inscho EW, Yuan L, Hill SM. Modulation of intracellular calcium and calmodulin by melatonin in MCF7 human breast cancer cells. *J Pineal Res.* 2002;32(2):112-119.
90. Fung TS, Liu DX. Activation of the c-Jun NH 2-terminal kinase pathway by coronavirus infectious bronchitis virus promotes apoptosis independently of c-Jun. *Cell Death Dis.* 2017;8(12):3215.
91. Zhang L, Liu Y. Potential interventions for novel coronavirus in China: a systematic review. *J Med Virol.* 2020;92(5):479-490.
92. Sui J, Li W, Murakami A, Tamin A, Matthews LJ, Wong SK, et al. Potent neutralization of severe acute respiratory syndrome (SARS) coronavirus by a human mAb to S1 protein that blocks receptor association. *Proc Natl Acad Sci U S A.* 2004;101(8):2536-2541.
93. Brink ENVD, Meulen JT, Cox F, Jongeneelen MA, Thijsse A, Throsby M, et al. Molecular and biological characterization of human monoclonal antibodies binding to the spike and nucleocapsid proteins of severe acute respiratory syndrome coronavirus. *J Virol.* 2005;79(3):1635-1644.
94. Shanmugaraj B, Siri wattananon K, Wangkanont K, Phoolcharoen W. Perspectives on monoclonal antibody therapy as potential therapeutic intervention for Coronavirus disease-19 (COVID-19). *Asian Pac J Allergy Immunol.* 2020; 38(1):10-18.
95. Ter Meulen J, Van Den Brink EN, Poon LL, Marissen WE, Leung CS, Cox F, et al. Human monoclonal antibody combination against SARS coronavirus: synergy and coverage of escape mutants. *Plos Med.* 2006;3(7):e237.
96. Berry JD, Hay K, Rini JM, Yu M, Wang L, Plummer FA, et al. Neutralizing epitopes of the SARS-CoV S-protein cluster independent of repertoire, antigen structure or mAb technology. *Mabs.* 2010;2(1):53-66.
97. Zhu Z, Chakraborti S, He Y, Roberts A, Sheahan T, Xiao X, et al. Potent cross-reactive neutralization of SARS coronavirus isolates by human monoclonal antibodies. *Proc Natl Acad Sci U S A.* 2007;104(29):12123-12128.
98. Lip K-M, Shen S, Yang X, Keng C-T, Zhang A, Oh H-LJ, et al. Monoclonal antibodies targeting the HR2 domain and the region immediately upstream of the HR2 of the S protein neutralize *in vitro* infection of severe acute respiratory syndrome coronavirus. *J Virol.* 2006;80(2):941-950.
99. Greenough TC, Babcock GJ, Roberts A, Hernandez HJ, Thomas Jr WD, Coccia JA, et al. Development and characterization of a severe acute respiratory syndrome-associated coronavirus-neutralizing human monoclonal antibody that provides effective immunoprophylaxis in mice. *J Infect Dis.* 2005;191(4):507-514.
100. Elshabrawy HA, Coughlin MM, Baker SC, Prabhakar BS. Human monoclonal antibodies against highly conserved HR1 and HR2 domains of the SARS-CoV spike protein are more broadly neutralizing. *Plos One.* 2012;7(11):e50366.
101. Walls AC, Xiong X, Park Y-J, Tortorici MA, Snijder J, Quispe J, et al. Unexpected receptor functional mimicry elucidates activation of coronavirus fusion. *Cell.* 2019;176(5):1026-1039.
102. Yi Y, Lagniton PN, Ye S, Li E, Xu RH, Zhong BL, et al. COVID-19: what has been learned and to be learned about the novel coronavirus disease. *Int J Biol Sci.* 2020;16(10):1753-1766.
103. Ying T, Li H, Lu L, Dimitrov DS, Jiang S. Development of human neutralizing monoclonal antibodies for prevention and therapy of MERS-CoV infections. *Microbes Infect.* 2015;17(2):142-148.
104. Yu X, Zhang S, Jiang L, Cui Y, Li D, Wang D, et al. Structural basis for the neutralization of MERS-CoV by a human monoclonal antibody MERS-27. *Sci Rep.* 2015;5(1):13133.
105. Zhang S, Zhou P, Wang P, Li Y, Jiang L, Jia W, et al. Structural definition of a unique neutralization epitope on the receptor-binding domain of MERS-CoV spike glycoprotein. *Cell Rep.* 2018;24(2):441-452.
106. Li Y, Wan Y, Liu P, Zhao J, Lu G, Qi J, et al. A humanized neutralizing antibody against MERS-CoV targeting the receptor-binding domain of the spike protein. *Cell Res.* 2015;25(11):1237-1249.
107. Doremalen NV, Falzarano D, Ying T, Wit E, Bushmaker T, Feldmann F, et al. Efficacy of antibody-based therapies against Middle East respiratory syndrome coronavirus (MERS-CoV) in common marmosets. *Antiviral Res.* 2017;143(1):30-37.
108. Ying T, Du L, Ju TW, Prabakaran P, Lau CC, Lu L, et al. Exceptionally potent neutralization of Middle East respiratory syndrome coronavirus by human monoclonal antibodies. *J Virol.* 2014;88(14):7796-7805.
109. Wang L, Shi W, Joyce MG, Modjarrad K, Zhang Y, Leung K, et al. Evaluation of candidate vaccine approaches for MERS-CoV. *Nat Commun.* 2015;6(1):7712.
110. Pallesen J, Wang N, Corbett KS, Wrapp D, Kirchdoerfer RN, Turner HL, et al. Immunogenicity and structures of a rationally designed prefusion MERS-CoV spike antigen. *Proc Natl Acad Sci U S A.* 2017;114(35):E7348-E7357.
111. Niu P, Zhao G, Deng Y, Sun S, Wang W, Zhou Y, et al. A novel human mAb (MERS-GD27) provides prophylactic and postexposure efficacy in MERS-CoV susceptible mice. *Sci China Life Sci.* 2018;61(10):1280-1282.
112. Niu P, Zhang S, Zhou P, Huang B, Deng Y, Qin K, et al. Ultrapotent human neutralizing antibody repertoires against Middle East respiratory syndrome coronavirus from a recovered patient. *J Infect Dis.* 2018;218(8):1249-1260.
113. Wit ED, Rasmussen AL, Falzarano D, Bushmaker T, Feldmann F, Brining DL, et al. Middle East respiratory syndrome coronavirus (MERS-CoV) causes transient lower respiratory tract infection in rhesus macaques. *Proc Natl Acad Sci U S A.* 2013;110(41):16598-16603.
114. Zhou H, Chen Y, Zhang S, Niu P, Qin K, Jia W, et al. Structural definition of a neutralization epitope on the N-terminal domain of MERS-CoV spike glycoprotein. *Nat Commun.* 2019;10(1):1-13.