

# Co-resilience: A Novel Therapeutic Approach to Resilience-Building of the Host and Gut Microbiota

Takumi Tochio<sup>1\*</sup>, Ayako Watanabe<sup>2</sup>, Yasuyuki Kitaura<sup>2</sup>, Koji Kawano<sup>3</sup>, Yasuhiro Koga<sup>4</sup>, Senju Hashimoto<sup>1</sup>, Ryoji Miyahara<sup>1</sup>, Naoto Kawabe<sup>1</sup>, Teiji Kuzuya<sup>1</sup>, Kazunori Nakaoka<sup>1</sup>, Takuji Nakano<sup>1</sup>, Yoshiki Hirooka<sup>1</sup>

<sup>1</sup>Department of Gastroenterology and Gastroenterological Oncology, Fujita Health University Toyoake, Aichi, Japan; <sup>2</sup>Department of Applied Biosciences, Graduate School of Bioagricultural Sciences, Nagoya University, Nagoya, Aichi, Japan; <sup>3</sup>Tokyo Animal Allergy Center, Adachi-ku, Tokyo, Japan; <sup>4</sup>Department of Infectious Diseases, Tokai University School of Medicine, Isehara, Kanagawa, Japan

## ABSTRACT

The resilience of the gut microbiota is a feature of healthy gut microbiota. Since associations between inflammatory-related diseases, such as type 2 diabetes or early-life allergic diseases, and the gut microbiota have been increasingly recognized in humans, besides lifestyle interventions, microbial interventions to increase the resilience of the gut microbiota, including probiotics, prebiotics, and fecal microbial transplantations, have been utilized clinically. However, inconsistent results and limitations are present in practice partially due to the intricate role of the gut microbiota in the disease. Thus, we explore the idea of "co-resilience," which emphasizes the importance of the resilience of the gut microbiota and that of the hosts to fitness with outlining the current evidence showing that gut microbial interventions influence host fitness for resilience-building management of the gut microbiota. We also propose a co-resilience-building approach that integrates gut microbial interventions with conventional therapy. The combined effects can render host-gut microbiota interactions beneficial, and such a novel co-resilience approach is expected to have a potent impact on host fitness.

**Keywords:** Gut microbiota; Resilience; Fitness; Dysbiosis; Gut microbial interventions; Co-Resilience

## INTRODUCTION

Accumulating studies have indicated links between the gut microbiota and health and diseases. The emerging interest in intestinal microbes involves altered microbial communities that are associated with a specific disease, particularly in inflammatory-related diseases (e.g., allergies and metabolic syndrome). This microbial alteration is often described as dysbiosis caused by extrinsic insults such as a disruptive diet and antibiotic medications. Experimental animal models have provided evidence for the role of the gut microbiota in the diseases [1,2], and restoration of the microbiota from an altered state by colonization with beneficial microbes mitigates physiological symptoms [3].

In this review, we consider evidence for resilience-building management of the gut microbiota that involves returning the microbiota to the pre-dysbiotic state with gut microbial interventions. Then, we argue that the idea of "co-resilience," which combines the resilience of the host with gut microbiota, is essential for a deeper understanding of our fitness. Finally, we propose that a co-resilience-building approach may lead to efficacious therapy for diseases.

## RESILIENCE-BUILDING MANAGEMENT OF THE GUT MICROBIOTA

### The resilience of the gut microbiota

The resilience of the gut microbiota is an expansionary concept derived from ecology, and is defined as "the amount of stress or perturbation that a system can tolerate before its trajectory changes towards a different equilibrium state" [4,5]. In this concept, although an acute disruptive force temporarily affects the gut microbial compositions, the gut microbiota can revert to the initial state due to resilience. For example, although a high-fat diet impacts the microbial communities in humans, a high-fat diet is less disruptive to the communities in the short term interventions than a regular long-term diet consumed over a year [6]. In other words, although what constitutes a "healthy human gut microbiota" is controversial [7] and remains to be defined as a single idealized set of microbial community organisms [8], resilience to disturbance is a feature of the healthy gut microbiota [7].

On the other hand, loss of resilience means that the gut microbiota loses its capacity to tolerate perturbation and thus become susceptible to damage to the gut microbial community. Namely, dysbiosis,

**Correspondence to:** Takumi Tochio, Department of Gastroenterology and Gastroenterological Oncology, Fujita Health University, Toyoake, Aichi, Japan, E-mail: t-tochio@bfsci.co.jp

**Received:** July 7, 2021; **Accepted:** July 21, 2021; **Published:** July 28, 2021

**Citation:** Tochio T, Watanabe A, Kitaura Y, Kawano K, Koga Y, Hashimoto T, et al. (2021) Co-resilience: A Novel Therapeutic Approach to Resilience-Building of the Host and Gut Microbiota. Clin Microbiol. 10: 214.

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described in the context of diseases, occurs due to external forces, such as a chronic disruptive diet or antibiotic use that overcome the internal resilience of the gut microbiota. For example, the gut microbiota of patients with Type 2 Diabetes (T2D) is relatively enriched with Bacteroidetes, Proteobacteria [9], and the imidazole propionate-producers, *Eggerthella lenta*, and *Streptococcus mutans* [10]. An increase in *Escherichia* is associated with Nonalcoholic Steatohepatitis (NASH) [11]. Also, recent studies have shown that neurodegenerative diseases such as Parkinson's Disease (PD) and Alzheimer's Disease (AD) may be linked to dysbiosis triggered by the accumulation of extrinsic insults such as aging, diets, and stress [12,13]. For example, the abundance of Prevotellaceae is reduced in feces of Parkinson's disease (PD) patients. Alzheimer's disease (AD) patients have an increased abundance of Bacteroidetes and a decreased abundance of Firmicutes [13]. Although "dysbiotic patterns" may differ between individuals due to multifactorial causes such as the long-term impact of lifestyle factors, these studies show clinically relevant changes in the gut microbial compositions in association with the loss of resilience of the gut microbiota.

The microbiota in infants is relatively unsettled over the first 3 years of life compared to adults [14]. In early-life development, an infant's microbial community is mostly shaped by maternal microbiota via prenatal and neonatal events such as birth mode and feeding [15]. In this context, the resilience of the gut microbiota of an infant is established during a critical period, and indeed, dysbiosis of the gut microbiota of both mothers and infants is considered to be co-dependently associated with health risk of infants. For example, in humans, *Bifidobacterium breve* (*B. breve*) and *Bifidobacterium infantis* in the mother's feces are associated with bifidobacteria counts and their diversity in the neonate's feces [16]. Moreover, decreased colonization with *Lactobacillus*, *Bifidobacterium*, and the butyrate-producer *Anaerostipes caccae* has been linked to food allergy and atopic diseases [3,17,18], whereas increased abundance of *Clostridium* and *Ruminococcus gnavus* is linked to allergy-related diseases [17-19]. These studies suggest that infants need to form a resilient microbial community during this critical time.

**Table 1:** Clinical studies of gut microbial interventions.

Gut microbial interventions	Participants	Main results	Refs
<b>Obesity</b>			
<i>Bifidobacterium adolescentis</i> IVS-1, <i>Bifidobacterium lactis</i> BB-12 with or without galactooligosaccharide for 3 weeks.	151 obese adults (BMI $\geq 30$ kg/m <sup>2</sup> )	Colonization with <i>B. adolescentis</i> IVS-1	[28]
Oligofructose for 12 weeks.	48 overweight or obese adults and older adults (BMI $\geq 25$ kg/m <sup>2</sup> )	Reduced body weight Suppressed ghrelin and enhanced peptide YY	[29]
<i>Lactobacillus salivarius</i> Ls-33 for 12 weeks.	51 obese adolescents (BMI $\geq 30$ kg/m <sup>2</sup> )	Altered fecal microbiota	[30]
<i>Bifidobacterium animalis</i> with or without polydextrose for 6 months.	225 obese adults (BMI 28-34.9 kg/m <sup>2</sup> )	Altered fecal microbiota Reduced body fat mass by administration of synbiotics	[31] [32]
Whole grain pasta containing barley $\beta$ -glucans and <i>Bacillus coagulans</i> for 12 weeks.	41 overweight and obese adults (BMI $\geq 25$ kg/m <sup>2</sup> )	No effect	[33]
Oligofructose-enriched inulin for 16 weeks.	42 overweight and obese children (BMI >85th percentile)	Reduction in body fat Increase in <i>Bifidobacterium</i> spp.	[34]
Inulin-type fructans (ITF) or whey protein with or without ITF for 12 weeks.	125 overweight and obese adults (BMI $\geq 25$ kg/m <sup>2</sup> )	Reduction in appetite Increase in fecal <i>Bifidobacterium</i>	[35]
Inulin plus resistant maltodextrin with energy-restricted diet for 12 weeks.	116 overweight and obese adults (BMI $\geq 28$ kg/m <sup>2</sup> )	No effect on weight loss. Increase in fecal Parabacteroides and <i>Bifidobacterium</i> .	[36]

## The resilience-building management of the gut microbiota

Gut microbial interventions (probiotics, prebiotics, and fecal microbiota transplantation) constitute resilience-building management of the gut microbiota. The association between dysbiosis and disease is still unclear regarding whether dysbiosis is a cause or consequence of diseases [20]; however, given that the gut microbiota largely contributes to host fitness, rectifying altered microbiota through gut microbial interventions is reasonable. For example, when a mixture of *Bifidobacterium* and *Lactobacillus* strains was ingested by healthy obese subjects for 6 months, a reduction in body weight and improved quality of life were observed [21]. Also, in patients with the chronic liver disease nonalcoholic fatty liver disease, the alanine aminotransferase concentration was reduced following administration of a mixture of *Bifidobacterium* and *Lactobacillus* strains combined with Fructooligosaccharides (FOS) [22]. Psychometric tests were improved following treatment with *Bifidobacterium* and Fructooligosaccharides (FOS) in patients with hepatic encephalopathy associated with liver disease [23]. Administration of *Lactobacillus rhamnosus* (*Lactobacillus* GG) or *B. breve* and *Bifidobacterium longum* to prenatal mothers and their neonates, respectively, more effectively prevented atopic dermatitis relative to the placebo group [24,25].

Dietary interventions using prebiotics are also feasible for resilience-building management because they provide fermentable resources for the selected microbiota. Prebiotic intervention in children with genetic and diet-induced obesity changes the gut microbiota compositions and improves health outcomes [26]. Similarly, substantial evidence suggests that prebiotics improve allergic diseases early in life. Therapeutic use of prebiotic 1-kestose, a component of Fructooligosaccharides (FOS), improves the clinical symptoms of atopic dermatitis in infants [27]. Table 1 summarizes the clinical evidence for gut microbial interventions. Thus, resilience-building management of the gut microbiota successfully increases the beneficial health outcomes in clinical trials [28-33].

Fructooligosaccharides (FOS) with or without multistrain probiotics ( <i>Lactobacillus paracasei</i> , <i>Lactobacillus rhamnosus</i> , <i>Lactobacillus acidophilus</i> , and <i>B. lactis</i> ) for 15 days.	9 obese adults scheduled for Roux-en-Y gastric bypass (BMI >40 kg/m <sup>2</sup> or >35 kg/m <sup>2</sup> with at least one comorbidity)	Reduced body weight with FOS	[37]
Arabinoxylans for 6 weeks.	47 overweight and obese adults (BMI 28-35 kg/m <sup>2</sup> )	Increased fecal SCFAs	[38]
<i>L. paracasei</i> or flaxseed mucilage for 6 weeks.	58 obese postmenopausal women (BMI 30-45 kg/m <sup>2</sup> )	Improved insulin sensitivity with flaxseed mucilage Altered fecal microbiota with flaxseed mucilage	[39]
<i>L. rhamnosus</i> for 24 weeks.	125 obese adults (BMI 29-41 kg/m <sup>2</sup> )	Reduced body weight in women	[40]
Multistrain probiotic ( <i>L. acidophilus</i> , <i>L. plantarum</i> , <i>Bifidobacterium bifidum</i> , and <i>B. animalis</i> subsp. <i>lactis</i> ) for 6 months.	220 overweight and obese adults (BMI 25-34.9 kg/m <sup>2</sup> )	Reduced body weight Improved quality of life	[21]
<b>Nonalcoholic Fatty Liver Disease (NAFLD)</b>			
Multistrain probiotics ( <i>L. plantarum</i> , <i>L. deslbrueckii</i> , <i>L. acidophilus</i> , <i>L. rhamnosus</i> , and <i>B. bifidum</i> ) for 6 months.	20 adult and older adult patients with histology-proven NASH	Reduced intrahepatic triglyceride content Decreased serum aspartate aminotransferase	[41]
Synbiotics ( <i>L. casei</i> , <i>L. rhamnosus</i> , <i>S. thermophilus</i> , <i>B. breve</i> , <i>L. acidophilus</i> , <i>B. longum</i> , and <i>L. bulgaricus</i> plus FOS) for 28 weeks.	52 adult and older adult patients with NAFLD (presence of steatosis, ALT >60 U/L)	Decreased in serum alanine aminotransferase (ALT)	[22]
Multistrain probiotics ( <i>S. thermophilus</i> , <i>B. breve</i> , <i>B. infantis</i> , <i>B. longum</i> , <i>L. acidophilus</i> , <i>L. plantarum</i> , <i>L. paracasei</i> , and <i>L. delbrueckii</i> subsp. <i>bulgaricus</i> ) for 4 months.	48 obese children with NAFLD (BMI >85th percentile)	Improvement in fatty liver Decrease in BMI	[42]
<i>Lactobacillus rhamnosus</i> GG for 8 weeks.	20 obese children (BMI >95th percentile)	Decrease in serum ALT	[43]
<b>Type 2 Diabetes (T2D)</b>			
Xylooligosaccharides (XOS) for 8 weeks.	13 prediabetic adults and older adults (BMI 27-35 kg/m <sup>2</sup> , fasting glucose level >100 mg/dL)	Altered fecal microbiota	[44]
Multistrain probiotics ( <i>B. bifidum</i> , <i>B. lactis</i> , <i>L. acidophilus</i> , <i>L. brevis</i> , <i>Lactobacillus casei</i> , <i>L. salivarius</i> , and <i>Lactococcus lactis</i> ) for 6 months.	96 adult and older adult patients with T2D	Decrease in HOMA-IR	[45]
Multistrain probiotics ( <i>Lactobacillus</i> , <i>Lactococcus</i> , <i>Bifidobacterium</i> , <i>Propionibacterium</i> , and <i>Acetobacter</i> ) for 8 weeks.	53 adult and older adult patients with T2D	Improved insulin resistance	[46]
<b>Metabolic Syndrome (MetS)</b>			
Intestinal microbial transfer from lean donor	18 male obese patients with MetS (BMI >30kg/m <sup>2</sup> , fasting plasma glucose level >5.6 mmol/L)	Improved insulin sensitivity 6 weeks after infusion. Increased butyrate-producing intestinal microbiota.	[47]
<i>L. casei</i> Shirota for 12 weeks.	30 adult patients with MetS	Increase in fecal Parabacteroides	[48]
<b>Hypercholesterolemia</b>			
<i>L. plantarum</i> for 12 weeks.	49 adults (total cholesterol 5.16-7.64 mmol/L)	Reduction in total cholesterol in high-cholesterol group	[49]
<i>L. reuteri</i> for 9 weeks.	127 hypercholesterolemic adults and older adults (LDL-cholesterol >3.4 mmol/L)	Reduction in LDL-Cholesterol	[50]
<b>Cystic Fibrosis (CF)</b>			
<i>L. reuteri</i> for 6 months.	61 pediatric and adult patients with CF	Decreased risk of pulmonary exacerbations. Reduced upper respiratory tract infections.	[51]
<b>Chronic Kidney Disease (CKD)</b>			
Synbiotics ( <i>L. plantarum</i> , <i>L. rhamnosus</i> , <i>L. gasseri</i> , <i>B. infantis</i> , <i>B. longum</i> , <i>L. acidophilus</i> , <i>L. salivarius</i> , <i>Lactobacillus sporogenes</i> , and <i>S. thermophilus</i> plus inulin) for 4 weeks.	30 adult and older adult participants with CKD (stage 3-4 CKD)	Lowered plasma total p-cresol concentrations	[52]
<b>Psychosis</b>			
Galactooligosaccharides for 12 weeks.	39 adult patients with psychotic disorder	Improved cognitive assessment	[53]

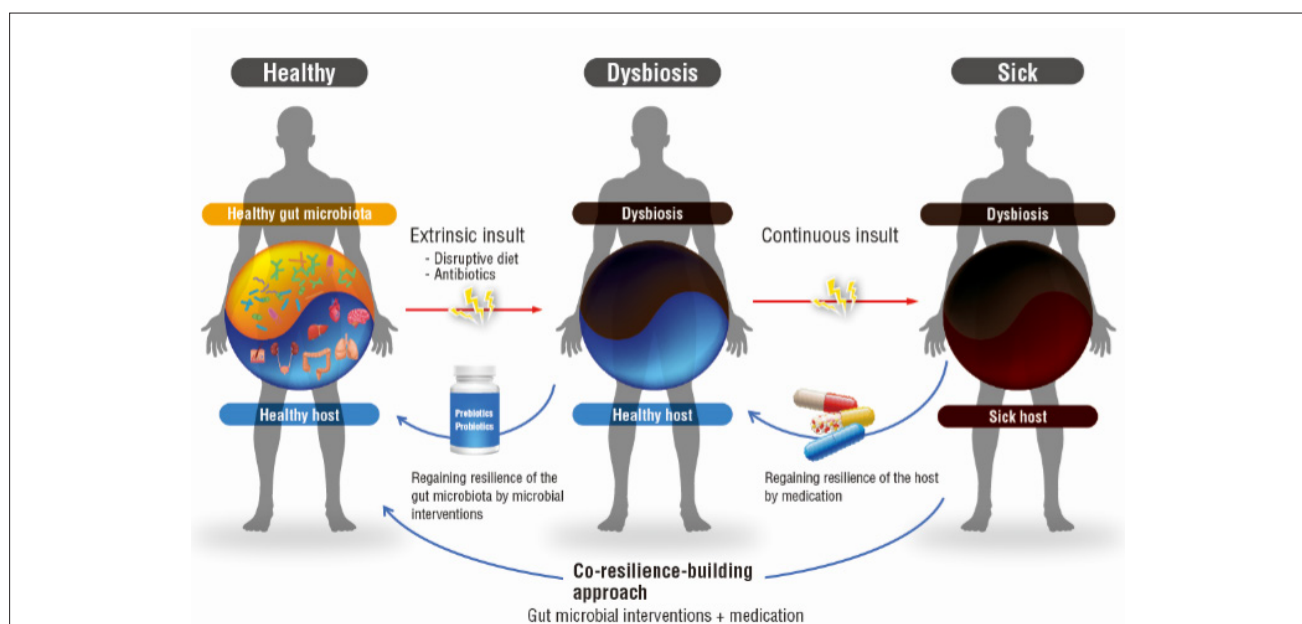
Autism			
Galactooligosaccharide for 6 weeks.	26 autistic children	Improved antisocial behavior by exclusion diet and prebiotics	[54]
<i>L. acidophilus</i> for 2 months.	22 autistic children	Decreased D-Arabinitol (DA) and the ratio of D-/L-arabinitol (DA/LA) in the urine	[55]
Atopic Dermatitis (AD)			
Synbiotics ( <i>B. breve</i> plus galacto-/fructooligosaccharide mixture) for 12 weeks.	90 infants with AD	No effect on AD severity Altered fecal microbiota	[56]
Cyanotic Congenital Heart Disease (CCHD)			
<i>B. lactis</i> plus inulin for 8 weeks.	100 infants with CCHD	Reduced incidence of nosocomial sepsis, necrotizing enterocolitis, and death	[57]

**Abbreviations:** BMI: Body Mass Index; ITF: Inulin-Type Fructans; FOS: Fructooligosaccharides; SCFA: Short-Chain Fatty Acid; NAFLD: Nonalcoholic Fatty Liver Disease; NASH: Non-Alcoholic Steatohepatitis; ALT: Alanine aminotransferase; T2D: Type 2 Diabetes; XOS: Xylooligosaccharides; HOMA-IR: Homeostatic Model Assessment of Insulin Resistance; MetS: Metabolic Syndrome; LDL: Low-Density Lipoproteins; CF: Cystic Fibrosis; CKD: Chronic Kidney Disease; AD: Atopic Dermatitis; DA: D-Arabinitol; LA: L-arabinitol; CCHD: Cyanotic Congenital Heart Disease.

## Co-resilience

The studies mentioned above emphasize the implications of being a human-microbe hybrid, i.e., a superorganism (Figure 1), and suggest the idea of “co-resilience” of the superorganism [34-39]. Conventional therapies target host tissues/organs that are susceptible to damage. Although tissues/organs can recover from damage, which is defined as resilience of the host, the damage results in severe diseases when the resilience of the host is lost [40-45]. When resilience is lost, drugs, exercise, and other interventions

support the recovery of tissues/organs to the pre-disease state by repairing damage [46-51]. For this reason, treatment for Type 2 Diabetes (T2D) and atopic dermatitis often include metformin and topical corticosteroids [52-57], which target glucose metabolism in the liver and skin inflammation, respectively [58,59]. However, because altered gut microbiota is linked to diseases both in adults and infants, guiding the establishment of health-promoting microbiota is essential for regaining host fitness. Such “co-resilience” may be promising for improving physiological outcomes.



**Figure 1:** Co-resilience-building approach. Humans are human-microbe hybrids, also called superorganisms (Left). In a healthy host, human tissues/organs and gut microbiota maintain fitness and a co-resilient state. External insults such as disruptive diet cause dysbiosis of the gut microbiota. The pre-dysbiotic state can be restored by gut microbial interventions (Middle). In a sick host, both human tissues/organs and the gut microbiota are damaged (Right). Conventional therapy such as medications is used as intervention in a sick host. However, treatments targeting the host and the gut microbiota, known as a “co-resilience-building approach”, will lead to the restoration of the superorganism fitness.

## Co-resilience-building approach

Combining conventional therapy with gut microbial interventions will require reconsideration of how we manage fitness. As mentioned, “Co-resilience” involving recovery from clinical disorders entails correction of damage to the gut microbial community via gut microbial interventions in addition to conventional therapy. Such a novel

approach may provide more efficacious therapy than conventional therapy alone. Thus, we propose a co-resilience-building approach (Figure 1). Evidence supporting the efficacy of this co-resilience-building approach is sparse at present (Table 2). A good example illustrating this approach is treatment for constipation, which is common in patients with Parkinson’s Disease (PD) [60,61].

**Table 2:** Tentative co-resilience-building approach.

Gut microbial interventions	Drugs	Participants	Main results	Ref.
<b>Constipation in Parkinson’s Disease (PD)</b>				
Multistrain probiotics ( <i>Streptococcus salivarius</i> subsp <i>thermophilus</i> , <i>Enterococcus faecium</i> , <i>Lactobacillus rhamnosus</i> GG, <i>Lactobacillus acidophilus</i> , <i>Lactobacillus plantarum</i> , <i>Lactobacillus paracasei</i> , <i>L. paracasei</i> , <i>delbrueckii</i> subsp <i>bulgaricus</i> , <i>Bifidobacterium breve</i> , and <i>animalis</i> subsp <i>lactis</i> plus FOS) for 4 weeks.	Laxatives, Dopamine agonist	120 patients with PD	Improved constipation	[60]
<b>Small-intestinal ulcers (adverse effect of acetylsalicylic acid)</b>				
After 6-week acetylsalicylic acid intake, <i>B. breve</i> for 8 weeks.	Acetylsalicylic acid	75 healthy adults	Lowered Lewis score (validation for ulcers)	[62]
<b>Type 2 Diabetes (T2D) with Non-Alcoholic Fatty Liver Disease (NAFLD)</b>				
Multistrain probiotics ( <i>Lactobacillus</i> , <i>Lactococcus</i> , <i>Bifidobacterium</i> , <i>Propionibacterium</i> , and <i>Acetobacter</i> plus omega-3 fatty acids) for 8 weeks.	Metformin, Sulfonylurea, Insulin	48 adult patients with T2D and NAFLD	Reduced fatty liver index. Improved serum lipids	[63]
<b>Candidemia</b>				
<i>Lactobacillus reuteri</i> or <i>L. rhamnosus</i> for 6 weeks.	Liposomal Amphotericin B, Antibiotics	249 pre-term infants	Lowered candida stool colonization	[64]

\*Note: The list includes clinical trials that can be considered to use a co-resilience-building approach.

Abbreviations: PD: Parkinson’s Disease; FOS: Fructooligosaccharides; T2D: Type 2 Diabetes; NAFLD: Non-Alcoholic Fatty Liver Disease.

## DISCUSSION

In a clinical trial, constipation improved in patients with Parkinson’s disease (PD) following administration of multistrain probiotics for 4 weeks. Because medications are unavoidable in Parkinson’s disease (PD), patients in the trial were allowed to take dopamine agonists and laxatives. Although how probiotics improve constipation in PD is still to be determined, this trial indicates that targeting tissues/organs and gut microbiota is effective in PD. Likewise, in T2D rats, the bioavailability of the drug, gliclazide, was enhanced by administration of probiotics, suggesting that probiotics have an adjuvant function with gliclazide [62-65]. Thus, rethinking conventional therapy and broadening the potential of the co-resilience-building approach will be worthwhile.

A potent co-resilience-building approach involves two considerations; one is an adjuvant role, and the other is minimizing the adverse effects of drugs. The former was described above regarding PD trials [60]. For the latter, probiotic *Bifidobacterium* intake, for example, reduces the number of ulcers, which is an adverse effect of acetylsalicylic acid [62], implying the potency of a co-resilience-building approach to minimize the adverse effects of drugs. Diseases can be treated with conventional medications and fine-tuned with gut microbial interventions, and thus, such a co-resilience approach may be efficacious (Figure 1) [66].

## CONCLUSION

The evidence presented here is not comprehensive, and the adverse effects of drug-microbiota interactions need to be considered. For example, non-antibiotic drugs have anticomensal activity, and 24% of screened drugs demonstrate inhibition of at least one species *in vitro*. Because the gut microbiota is seldom binary and intricate, studying how these alterations caused by gut microbial interventions affect the host will be essential. Thus, our proposal needs to be tested

rigorously in animal models and clinical trials, and the co-resilience-building approach should be implemented with the right combination of drug-microbiota interactions. With a shift in the co-resilience-building approach, treatment of a disease may improve the outcome for patients, and continuous active interventions will help create and sustain a healthy state for both the host and the gut microbiota.

## ACKNOWLEDGMENT

We thank Hidenobu Shinohara for illustrations.

## CONFLICT OF INTEREST

The authors declare that they have no competing interests.

## REFERENCES

- Ridaura VK, Faith JJ, Rey FE, Cheng J, Duncan AE, Kau AL, et al. Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science*. 2013; 341(6150):1241-1244.
- Cani PD, Neyrinck AM, Fava F, Knauf C, Burcelin RG, Tuohy KM, et al. Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia. *Diabetologia*. 2007; 50(11):2374-2383.
- Feehley T, Plunkett CH, Bao RY, Hong SMC, Cullen E, Belda-Ferre P, et al. Healthy infants harbor intestinal bacteria that protect against food allergy. *Nat Med*. 2019; 25(3):448-453.
- Folke C, Carpenter S, Walker B, Scheffer M, Elmqvist T, Gunderson L, et al. Regime shifts, resilience, and biodiversity in ecosystem management. *Annual Review of Ecology, Evolution, and Systematics*. 2004; 35(1):557-581.

5. Lozupone CA, Stombaugh JI, Gordon JI, Jansson JK, Knight R. Diversity, stability and resilience of the human gut microbiota. *Nature*. 2012; 489(7415):220-230.
6. Wu GD, Chen J, Hoffmann C, Bittinger K, Chen YY, Keilbaugh SA, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science*. 2011; 334(6052):105-108.
7. Bäckhed F, Fraser CM, Ringel Y, Sanders ME, Sartor RB, Sherman PM, et al. Defining a healthy human gut microbiome: Current concepts, future directions, and clinical applications. *Cell Host Microbe*. 2012; 12(5):611-622.
8. Mcburney MI, Davis C, Fraser CM, Schneeman BO, Huttenhower C, Verbeke K, et al. Establishing what constitutes a healthy human gut microbiome: State of the science, regulatory considerations, and future directions. *J Nutr*. 2019; 149(11):1882-1895.
9. Larsen N, Vogensen FK, van den Berg FWJ, Nielsen DS, Andreasen AS, Pedersen BK, et al. Gut microbiota in human adults with Type 2 Diabetes differs from non-diabetic adults. *PLoS ONE*. 2010; 5(2):e9085.
10. Koh A, Molinaro A, Ståhlman M, Khan MT, Schmidt C, Mannerås-Holm L, et al. Microbially produced imidazole propionate impairs insulin signaling through mTORC1. *Cell*. 2018; 175(4):947-61.e17.
11. Zhu LX, Baker SS, Gill C, Liu WS, Alkhoury R, Baker RD, et al. Characterization of gut microbiomes in Nonalcoholic Steatohepatitis (NASH) patients: A connection between endogenous alcohol and NASH. *Hepatology*. 2013; 57(2):601-609.
12. Scheperjans F, Aho V, Pereira PA, Koskinen K, Paulin L, Pekkonen E, et al. Gut microbiota are related to Parkinson's disease and clinical phenotype. *Mov Disord*. 2015; 30(3):350-358.
13. Vogt NM, Kerby RL, Dill-McFarland KA, Harding SJ, Merluzzi AP, Johnson SC, et al. Gut microbiome alterations in alzheimer's disease. *Sci Rep*. 2017; 7(1):13537.
14. Yatsunenkov T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, et al. Human gut microbiome viewed across age and geography. *Nature*. 2012; 486(7402):222-227.
15. Forgie AJ, Drall KM, Bourque SL, Field CJ, Kozyrskyj AL, Willing BP. The impact of maternal and early life malnutrition on health: A diet-microbe perspective. *BMC Med*. 2020; 18(1):135.
16. Mikami K, Takahashi H, Kimura M, Isozaki M, Izuchi K, Shibata R, et al. Influence of maternal bifidobacteria on the establishment of bifidobacteria colonizing the gut in infants. *Pediatr Res*. 2009; 65(6):669-674.
17. Bjorksten B, Naaber P, Sepp E, Mikelsaar M. The intestinal microflora in allergic estonian and Swedish 2-Year-Old children. *Clin Exp Allergy*. 1999; 29(3):342-346.
18. Kalliomaki M, Kirjavainen P, Eerola E, Kero P, Salminen S, Isolauri E. Distinct patterns of neonatal gut microflora in infants in whom atopy was and was not developing. *J Allergy Clin Immunol*. 2001; 107(1):129-134.
19. Chua HH, Chou HC, Tung YL, Chiang BL, Liao CC, Liu HH, et al. Intestinal dysbiosis featuring abundance of *Ruminococcus gnavis* associates with allergic diseases in infants. *Gastroenterology*. 2018; 154(1):154-167.
20. Schmidt TSB, Raes J, Bork P. The Human gut microbiome: From association to modulation. *Cell*. 2018; 172(6):1198-1215.
21. Michael DR, Jack AA, Masetti G, Davies TS, Loxley KE, Kerry-Smith J, et al. A randomised controlled study shows supplementation of overweight and obese adults with lactobacilli and bifidobacteria reduces bodyweight and improves well-being. *Sci Rep*. 2020; 10(1).
22. Eslamparast T, Poustchi H, Zamani F, Sharafkhan M, Malekzadeh R, Hekmatdoost A. Synbiotic supplementation in nonalcoholic fatty liver disease: A randomised, double-blind, placebo-controlled pilot study. *American J Clin Nutr*. 2014; 99(3):535-542.
23. Malaguarnera M, Gargante MP, Malaguarnera G, Salmeri M, Mastrojeni S, Rampello L, et al. *Bifidobacterium* combined with fructooligosaccharide versus lactulose in the treatment of patients with hepatic encephalopathy. *Eur J Gastroenterol Hepatol*. 2010; 22(2):199-206.
24. Kalliomaki M, Salminen S, Arvilommi H, Kero P, Koskinen P, Isolauri E. Probiotics in primary prevention of atopic disease: A randomised placebo-controlled trial. *Lancet*. 2001; 357(9262):1076-1079.
25. Enomoto T, Sowa M, Nishimori K, Shimazu S, Yoshida A, Yamada K, et al. Effects of bifidobacterial supplementation to pregnant women and infants in the prevention of allergy development in infants and on fecal microbiota. *Allergol Int*. 2014; 63(4):575-85.
26. Zhang C, Yin A, Li H, Wang R, Wu G, Shen J, et al. Dietary modulation of gut microbiota contributes to alleviation of both genetic and simple obesity in children. *EBioMedicine*. 2015; 2(8):968-984.
27. Shibata R, Kimura M, Takahashi H, Mikami K, Aiba Y, Takeda H, et al. Clinical effects of kestose, a prebiotic oligosaccharide, on the treatment of atopic dermatitis in infants. *Clin Exp Allergy*. 2009; 39(9):1397-1403.
28. Krumbeck JA, Rasmussen HE, Hutkins RW, Clarke J, Shawron K, Keshavarzian A, et al. Probiotic *bifidobacterium* strains and galactooligosaccharides improve intestinal barrier function in obese adults but show no synergism when used together as synbiotics. *Microbiome*. 2018; 6(1):121.
29. Parnell JA, Reimer RA. Weight loss during oligofructose supplementation is associated with decreased ghrelin and increased peptide YY in overweight and obese adults. *Am J Clin Nutr*. 2009; 89(6):1751-1759.
30. Larsen N, Vogensen FK, Gøbel RJ, Michaelsen KF, Forssten SD, Lahtinen SJ, et al. Effect of *lactobacillus salivarius* Ls-33 on fecal microbiota in obese adolescents. *Clin Nutr*. 2013; 32(6):935-940.
31. Stenman LK, Lehtinen MJ, Meland N, Christensen JE, Yeung N, Saarinen MT, et al. Probiotic with or without fiber controls body fat mass, associated with serum zonulin, in overweight and obese adults—randomised controlled trial. *EBioMedicine*. 2016; 13:190-200.
32. Hibberd AA, Yde CC, Ziegler ML, Honore AH, Saarinen MT, Lahtinen S, et al. Probiotic or synbiotic alters the gut microbiota and metabolism in a randomised controlled trial of weight management in overweight adults. *Benef Microbes*. 2019; 10(2):121-135.
33. Angelino D, Martina A, Rosi A, Veronesi L, Antonini M, Mennella I, et al. Glucose- and lipid-related biomarkers are affected in healthy obese or hyperglycemic adults consuming a whole-grain pasta enriched in prebiotics and probiotics: A 12-week randomised controlled trial. *J Nutr*. 2019; 149(10):1714-1723.
34. Nicolucci AC, Hume MP, Martinez I, Mayengbam S, Walter J, Reimer RA. Prebiotics reduce body fat and alter intestinal microbiota in children who are overweight or with obesity. *Gastroenterology*. 2017; 153(3):711-722.
35. Reimer RA, Willis HJ, Tunnicliffe JM, Park H, Madsen KL, Soto-Vaca A. Inulin-type fructans and whey protein both modulate appetite but only fructans alter gut microbiota in adults with overweight/obesity: A randomised controlled trial. *Mol Nutr Food Res*. 2017; 61(11):1700484.
36. Hess AL, Benítez-Páez A, Blädel T, Larsen LH, Iglesias JR, Madera C, et al. The effect of inulin and resistant maltodextrin on weight loss during energy restriction: A randomised, placebo-controlled, double-blinded intervention. *Eur J of Nutr*. 2020; 59(6):2507-2524.

37. Fernandes R, Beserra BTS, Mocellin MC, Kuntz MGF, Da Rosa JS, De Miranda RCD, et al. Effects of prebiotic and synbiotic supplementation on inflammatory markers and anthropometric indices after Roux-en-Y gastric bypass. *J Clin Gastroenterol.* 2016; 50(3):208-217.
38. Salden BN, Troost FJ, Wilms E, Truchado P, Vilchez-Vargas R, Pieper DH, et al. Reinforcement of intestinal epithelial barrier by arabinoxylans in overweight and obese subjects: A randomised controlled trial. *Clin Nutr.* 2018; 37(2):471-480.
39. Brahe LK, Le Chatelier E, Prifti E, Pons N, Kennedy S, Blädel T, et al. Dietary modulation of the gut microbiota – A randomised controlled trial in obese postmenopausal women. *Br J Nutr.* 2015; 114(3):406-417.
40. Sanchez M, Darimont C, Drapeau V, Emady-Azar S, Lepage M, Rezzonico E, et al. Effect of *Lactobacillus rhamnosus* CGMCC1.3724 supplementation on weight loss and maintenance in obese men and women. *Br J Nutr.* 2014; 111(8):1507-1519.
41. Wong VWS, Wong GLH, Chim AML, Chu WCW, Yeung DKW, Li KCT, et al. Treatment of nonalcoholic steatohepatitis with probiotics. A proof-of-concept study. *Ann Hepatol.* 2013; 12(2):256-262.
42. Alisi A, Bedogni G, Baviera G, Giorgio V, Porro E, Paris C, et al. Randomised clinical trial: The beneficial effects of VSL#3 in obese children with non-alcoholic steatohepatitis. *Alimentary pharmacology & therapeutics.* 2014; 39(11):1276-1285.
43. Vajro P, Mandato C, Licenziati MR, Franzese A, Vitale DF, Lenta S, et al. Effects of *Lactobacillus rhamnosus* strain GG in pediatric obesity-related liver disease. *J Pediatr Gastroenterol Nutr.* 2011; 52(6):740-743.
44. Yang J, Summanen PH, Henning SM, Hsu M, Lam H, Huang J, et al. Xylooligosaccharide supplementation alters gut bacteria in both healthy and prediabetic adults: A pilot study. *Front Physiol.* 2015; 6:216.
45. Sabico S, Al-Mashharawi A, Al-Daghri NM, Wani K, Amer OE, Hussain DS, et al. Effects of a 6-month multi-strain probiotics supplementation in endotoxemic, inflammatory and cardiometabolic status of T2DM patients: A randomised, double-blind, placebo-controlled trial. *Clin Nutr.* 2019; 38(4):1561-1569.
46. Kobylak N, Falalyeyeva T, Mykhalchyshyn G, Kyriienko D, Komissarenko I. Effect of alive probiotic on insulin resistance in Type 2 Diabetes patients: Randomised clinical trial. *Diabetes Metab Syndr.* 2018; 12(5):617-624.
47. Vrieze A, Van Nood E, Holleman F, Salojärvi J, Kootte RS, Bartelsman JF, et al. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology.* 2012; 143(4):913-6.e7.
48. Stadlbauer V, Leber B, Lemesch S, Trajanoski S, Bashir M, Horvath A, et al. *Lactobacillus casei* shirota supplementation does not restore gut microbiota composition and gut barrier in metabolic syndrome: A randomised pilot study. *PLoS One.* 2015; 10(10):e0141399.
49. Costabile A, Buttarazzi I, Kolida S, Quercia S, Baldini J, Swann JR, et al. An *in vivo* assessment of the cholesterol-lowering efficacy of *Lactobacillus plantarum* ECGC 13110402 in normal to mildly hypercholesterolaemic adults. *PLoS One.* 2017; 12(12):e0187964.
50. Jones MJ, Martoni CJ, Prakash S. Cholesterol lowering and inhibition of sterol absorption by *Lactobacillus reuteri* NCIMB 30242: A randomised controlled trial. *Eur J Clin Nutr.* 2012; 66(11):1234-1241.
51. Di Nardo G, Oliva S, Menichella A, Riccardo Pistelli R, Valerio De Biase R, Patriarchi F, et al. *Lactobacillus reuteri* ATCC55730 in cystic fibrosis. *J Pediatr Gastroenterol Nutr.* 2014; 58(1):81-86.
52. Guida B, Germanò R, Trio R, Russo D, Memoli B, Grumetto L, et al. Effect of short-term synbiotic treatment on plasma p-cresol levels in patients with chronic renal failure: A randomised clinical trial. *Nutr Metab Cardiovasc Dis.* 2014; 24(9):1043-1049.
53. Kao AC, Safarikova J, Marquardt T, Mullins B, Lennox BR, Burnet PWJ. Pro-cognitive effect of a prebiotic in psychosis: A double blind placebo controlled cross-over study. *Schizophr Res.* 2019; 208:460-461.
54. Grimaldi R, Gibson GR, Vulevic J, Giallourou N, Castro-Mejía JL, Hansen LH, et al. A prebiotic intervention study in children with Autism Spectrum Disorders (ASDs). *Microbiome.* 2018;6(1).
55. Kaluzna-Czaplinska J, Blaszczyk S. The level of arabinitol in autistic children after probiotic therapy. *Nutrition.* 2012; 28(2):124-126.
56. Van Der Aa LB, Heymans HS, Van Aalderen WM, Sillevs Smitt JH, Knol J, Amor KB, et al. Effect of a new synbiotic mixture on atopic dermatitis in infants: A randomised-controlled trial. *Clin Exp Allergy.* 2010.
57. Dilli D, Aydin B, Zenciroglu A, Ozyazici E, Beken S, Okumus N. Treatment outcomes of infants with cyanotic congenital heart disease treated with synbiotics. *Pediatrics.* 2013;132(4):e932-e938.
58. Rena G, Hardie DG, Pearson ER. The mechanisms of action of metformin. *Diabetologia.* 2017; 60(9):1577-1585.
59. Hoare C, Li Wan Po A, Williams H. Systematic review of treatments for atopic eczema. *Health Technol Assess.* 2000; 4(37):1-191.
60. Barichella M, Pacchetti C, Bolliri C, Cassani E, Iorio L, Pusani C, et al. Probiotics and prebiotic fiber for constipation associated with Parkinson's disease: An RCT. *Neurology.* 2016; 87(12):1274-1280.
61. Fasano A, Visanji NP, Liu LWC, Lang AE, Pfeiffer RF. Gastrointestinal dysfunction in Parkinson's disease. *The Lancet Neurol.* 2015; 14(6):625-639.
62. Mortensen B, Murphy C, O'Grady J, Lucey M, Elsafi G, Barry L, et al. *Bifidobacterium breve* Bif195 protects against small-intestinal damage caused by acetylsalicylic acid in healthy volunteers. *Gastroenterology.* 2019; 157(3):637-46.e4.
63. Kobylak N, Abenavoli L, Falalyeyeva T, Mykhalchyshyn G, Boccuto L, Kononenko L, et al. Beneficial effects of probiotic combination with omega-3 fatty acids in NAFLD: A randomised clinical study. *Minerva Med.* 2018; 109(6):418-428.
64. Romeo MG, Romeo DM, Trovato L, Oliveri S, Palermo F, Cota F, et al. Role of probiotics in the prevention of the enteric colonization by candida in preterm newborns: Incidence of late-onset sepsis and neurological outcome. *J Perinatol.* 2011; 31(1):63-69.
65. Al-Salami H, Butt G, Fawcett JP, Tucker IG, Golocorbin-Kon S, Mikov M. Probiotic treatment reduces blood glucose levels and increases systemic absorption of gliclazide in diabetic rats. *Eur J Drug Metab Pharmacokinet.* 2008; 33(2):101-106.
66. Maier L, Pruteanu M, Kuhn M, Zeller G, Telzerow A, Anderson EE, et al. Extensive impact of non-antibiotic drugs on human gut bacteria. *Nature.* 2018; 555(7698):623-628.