(April-June, 2015)



GLOBAL JOURNAL OF BIOLOGY, AGRICULTURE & HEALTH SCIENCES (Published By: Global Institute for Research & Education)

## www.gifre.org

# Biological and Nanocomposite Control of *Fusarium Wilt* of Potato Caused by *Fusarium Oxysporum* f. sp. *Tuberosi*

Abeer H. Makhlouf<sup>(1)</sup> & Rehab Abdeen<sup>(2)</sup> <sup>(1)</sup>Faculty of Agriculture, Minufiya University <sup>(2)</sup>Faculty of science, Tanta university

## Abstract

The present study indicated the antagonists activity of different biocontrol agents against a soil – born *pathogen Fusarium oxysporum*, that causes wilt disease of potato (Hermis and Spunta cvs.). *In vitro*, the effect of antagonists by the modified montmorillonite particles (Mod- MMT) as a fungicide with concentration (1,3 and 5%), and some strains of fungus *Trichoderma* and *Pseudomonas* revealed the presence of clear antagonistic action against *Fusarium oxysporum* f. sp. *tuberosi*. The highest mean inhibition values were 93.33% and 90% with Mod-MMT 5% and Mod-MMT 3% combined with *Trichoderma harzianum* respectively, while the lowest effect was investigated with *Pseudomonas fluorescens* only 51.11%. Under green house conditions the results proved that soil treatments was more effective than tuber treatments against pre and post wilting disease. The highest protection against the pathogen cleared with Mod-MMT 5% and Mod-MMT 5% and Mod-MMT 5% and Mod-MMT 3% combined with *Trichoderma harzianum* respectively as soil treatment. This treatment was superior to Mod-MMT 5% and Mod-MMT 3% combined with *Trichoderma harzianum* respectively as soil treatment. This treatment was superior to Mod-MMT 3% combined with *Trichoderma harzianum* respectively as soil treatment. This treatment was superior to infested control. All biological treatments also significantly decreased the incidence of disease specially Mod-MMT 5% and Mod-MMT 3% combined with *Trichoderma*. On the other hand the all bio- control agents reduced the population of *Fusarium oxysporum* gradually up to 8weeks after sowing. Also all biocontrol agents increased plant hight(cm./plant), fresh and dry weight(gm./ plant), number of tubers/ plant and weight of tubers (gm./ plant).

Key words: biological control -nanocomposite- potato - Fusarium wilt- Trichoderma- Pseudomonas

### Introduction

Potato (*Solanum tuberosum* L.) is a crop of major economic importance worldwide (Tsegaw, 2005; FAO, 2008). It is the world's fourth-largest food crop, following rice, wheat and maize (Harris, 1992; FAO, 2008). It widely cultivated, and could contribute to reducing worldwide food shortages, So it is a useful crop that provide high yields of carbohydrate and protein from their tubers so it has an important role in human nutrition (OrzoLek *et al.*2010). The potato tubers could be easily invaded by pests and pathogens such as *Fusarium oxysporum* (Agrios, 1997).

*Fusarium* wilt is a worldwide important disease of potato and can be transmitted by seed cuts (Powelson and Rowe, 1993). This disease caused by *F. oxysporum tuberosi* which is a soil-borne hyphomycete and causes vascular wilts of flowering plants (Domsch *et al.* 1980; Nelson *et al.* 1983). It considers one of the most destructive and economically damaging diseases of potato that lead to yield Losses (Stevenson *et al.*,2001 and Haggag,2008). This disease, characterized by generally symptoms such as wilting, chlorosis, necrosis, premature leaf drop, browning of the vascular system, stunting, and damping-off. The most important of these is vascular wilt., frequently results in the death of infected plants (Jones *et al.*,1991).

In view of difficulties and problems associated with chemical control of soil borne plant pathogens and environmental pollution, employment of biocontrol agents for plant disease management is considered as a good alternative (Blond 1990).

Using *Trichoderma* species as biocontrol agents was recognized for the first time by (Weindling, 1932). During recent decades, attention has been paid to this group of fungi and subsequently they have been applied successfully as biocontrol agents against several plant diseases in commercial agriculture (Howell, 2003 and Farkhondeh *et al* .,2013). *Trichoderma* spp. is an unique ability to control the plant pathogens by using the mechanism of mycoparasitism, antibiosis, competition, siderophore production, induction of systemic resistance etc (Chet, 1987; Dennis and Webster 1971, Upadhyay and Mukhopadhyay, 1986; Howell, 2003 ). Many studies cleared that *Trichoderma* spp.have antagonistic and biologically control potential against a diversity of soil borne pathogens.(Grondona *et al.*, 1997; Hanson and Howell, 2004; Bajwa *et al.*, 2004). Antagonistic ability of different isolates of *Trichoderma* on different formae specials of *Fusarium oxysporum* is well documented (Kerkeni *et al.*, 2007; Dohroo, 1995; Orole and Adejumo, 2009 ; Sivan and Chet, 1989; Pawan and Vijay 2011).

*Pseudomonas* spp. are suitable for application as agricultural bio-control agents since they can use many exudates compounds as a nutrient source. It has been widely used as antagonists against fusarium wilt of several agriculture important crops (Alabouvette and Steinberg, 1995; Aeshah *et al.*, 2011). Certain members of the *P. fluorescens* have been shown to be potential agents for the bio-control which suppress plant diseases by protecting the seeds and roots from fungal infection. They are known to enhance plant growth promotion and reduce severity of many fungal diseases (Hoffland *et al.* 1996, Wei *et al.* 1996). This effect is the result of the production of a number of secondary metabolites including antibiotics, siderophores and hydrogen cyanide (O'Sullivan and O'Gara 1992). Hass and Defago (2005) reviewed the mechanisms by which *P. fluorescens* control pathogenic microorganisms in detail. Competitive exclusion of pathogens as the result of rapid colonization of the rhizosphere by *P. fluorescens* may also be an important factor in disease control (Girija and Manoj 2005).

(April-June, 2015)

Montmorillonite (MMT), a bioinert clay mineral with fine grain and large inter-planner spacing in the (001) plane, has superior capability to intercalate large molecules into the interlayer space of the (001) plane. The structure of MMT is an octahedral laminated sheet, sandwiched between tetrahedral silicate layers (Bremner, 1982 and Boyd et al., 1988). The expandable layered aluminosilicate structure of clay minerals, consisting of stacks of plate-like layers of ~1 nm thickness, separated by an interlayer distance of  $\sim 1$  nm. The platelets with an aspect ratio in the range of 20-100 nm have an extremely large surface area of ~750 m2 g-1. The layers are not stacked regularly enough to form true crystal but are grouped in small numbers not exceeding dozen-forming stacks, inside which the only degree of organization is a parallelism sufficient to result in selective d001 reflection. Stacks of parallel layers form primary particle and number of primary particles form aggregate and microaggregate (0.1 -10 µm) ( Mering, 1946). It has some excellent properties such as good water absorption, swelling, absorbability, cation exchange and drug carrying capability (Macter et al., 1990), that are considered beneficial from the viewpoint of synthesis of pharmaceutical products as both inactive and active substances (Ricka et al. 1984 and Hirokawa et al., 1984). However, in spite of the beneficial effects of clay minerals, there are some inherent drawbacks associated with their use for drug delivery. Under physiological conditions clay dispersions are unstable and tend to flocculate and precipitate in ion containing solution, because of high salt concentration and the presence of polyelectrolytes such as proteins. Stability of dispersion is an important requirement for drug carriers because it plays a determining role with regard to adsorption and bioavailability. Furthermore, the ability of clay particles to adsorb negatively charged or neutral drugs is low, restricting their application as carriers of negatively charged or neutral drugs (Aguzzi et al., 2007). MMT is a natural biodegradable copolymer of N-acetylglucosamine and D-glucosamine that is usually obtained from exoskeletons of shellfish and insects. With regard to its unique properties such as biocompatibility, biodegradability, non-toxicity and non allergenic with antimicrobial effect, it is widely used in fields like biotechnology, biomedical, pharmaceutics, cosmetics, textile and agriculture (Mazzarelli, 1973 and Darder et al., 2003).

The present study was undertaken to find out environmental friendly methods of controlling *Fusarium oxysporum* f. sp. *Tuberosi*, using modified montmorillonite nanocomposite with different ratio (Mod-MMT), *Trichoderma harzianum* and *Pseudomonas fluorescens* individually or combination.

### **Materials and Methods**

## Isolation and identification of *F.oxysporum*

Potato plants showing wilt symptoms were collected from different locations at Kafr El-Sheikh and Minufiya governorates, Egypt at flowering stage. Small pieces of diseased specimens (stem vascular tissue) were grown on Potato Dextrose Agar plates (PDA), then kept in incubator at 25°C for 3days .After purification, isolates were identified according to their morphological ,microscopical and pathological characteristics with the help of standard key (Gilman1957; Burnett and Hunter 1972; Nelson *et al.*, 1983).

#### **Pathogenicity tests**

For pathogenicity tests ten isolates of *F. oxysporum* from thirty two isolates were selected. Pathogenicity of the isolates toward potato plants (Hermis and Spunta cvs.) were estimated on potted potato plants according to Khalifa (1991). Nine days old *F. oxysporum* isolates grown on potato dextrose agar (PDA) at 25 °C were transferred to 500 ml Erlenmayer flasks containing 250 ml PDB (potato dextrose liquid broth) and the flasks were incubated in shaker at 25 °C for 4 days. Cultures were passed through cheesecloth to separate the mycelia from the spores and the final concentration of the spore suspension was adjusted to $10^8$  spores/ml. Twenty ml of these suspensions was added to each pot in which potato tubers were grown. Three pots were used for each isolate and plants were observed for appearance of disease symptoms Pathogens were re-isolated from randomly selected diseased plants, the Koch's postulates were followed in order to confirm the pathogenicity of *F. oxysporum* isolates.

#### Determine the index of leaf damage

A scale of 0-4  $\alpha$  according to (Beye and Lafay ,1985) was used to determine the disease severity every weak 0: none symptoms on leaf, 1: leaf wilted, 2:leaf with yellowing, 3:leaf with necrosis and 4: dead leaf.

ILD=  $\sum$  notes / Max

ILD : Index of leaf damage

 $\sum$  notes : Total notes

Max : 4 times of developed leaf number

#### Isolation and identification of antagonists:

From soil rhizosphere samples (20 cm deep) of healthy potato plants antagonists were isolated from different fields at Kafr El - Sheikh and Minufiya governorates, Egypt. The used bioagents *Pseudomonas fluorescens* and *Trichoderma harzianum* were isolated on selected media according to the methods recommended by (Burgess *et al* .1994); King *et al* . 1954 ; Davet 1979 and Turner *et al*. 1998 ) five isolates from each antagonist were used. After proper growth they were incubated at 25 °C., isolates were purified and identified on the basis of their morphological characteristics( Buchanan and Gibbons, 1974; and Rifai, 1969).

#### **Praperatiom of modified MMT**

The clay mineral used in this study was sodium MMT (Colloid BP) from Southern Clay Products Inc (Gonzales, Texas, USA) with cation exchange capacity (CEC) of 114.8 meq/100g. Chitosan (CS) with the deacetylation degree of 70 % and the molecular weight (Mw) of 400.000 was purchased from Aldrich chemicals. Triphenyl phosphine and chloroacetylchloride were purchased from Aldrich chemicals and were used as received without further purification. The structure of MMT is shown in Fig. 1.

G.J.B.A.H.S., Vol.4(2):151-163 (April-June, 2015) В 🔿 Al, Fe, Ma, Li CH2-OH 0 🕀 Li. Na. Rb. Cs OH O NH 1 O CCH<sub>2</sub>Ph<sub>3</sub>P<sup>+</sup>MMT<sup>-</sup>

## Fig 1: (A) Structure of MMT and (B) Structure of Mod-MMT

### Chloroacetylation of chitosan.

Under magnetic stirring drops of pyridine (75.2 ml, 99.6 mmol) were added to 1 g chitosan. The mixture was cooled in an ice-salt bath and chloroacetyl chloride (7.965 ml, 99.6 mmol) was added drop wise with stirring. The reaction mixture was stirred overnight at 40 °C for three days. The chloroacetylated chitosan was precipitated by addition of dilute HCl (1N), filtered off and washed with distilled water several times and the product was dried to give 0.95 g (N. Salahuddin,2013).

## Synthesis of Triphenyl-chloro acetylated chitosan phosphonium salt (mod-CS).

2 g of chloro acetylatyed chitosan was dissolved in 20 ml dry DMF followed by addition of 5.33 g (20 mmol) triphenyl phosphine. The reaction mixture was stirred for four days at 80 °C in an oil bath. The product was filtered off, washed with DMF to give 1.5 g (N. Salahuddin, 2013).

### Preparation of Triphenyl- (chloro acetylated chitosan) phosphonium salt-montmorillonite intercalates (Mod/MMT).

1 g of Na-MMT was swelled in 30 ml water by stirring several hours at 40°C. A solution of 100 g mod-CS dissolved in 100 ml DMF was added and stirred for 24 hr. The product (Mod-MMT 1%) was filtered, washed several times with water and collected by a filter press and dried at 80 °C in vacuum oven for 10 hr. The same procedure was done using 3 g and 5 g of mod-CS to prepare (Mod-/MMT 3%) and (Mod-MMT 5%) respectively (Table 1). Table 1. Composition data of mod-MMT with different concentrations.

Antagonistic effect in vitro: Organo/clay clay (wt) Condition of reaction Mod-Yield % Cs(wt) Support Mod-MMT (1%) (1)Solvent method at room (100 g)75 temp. Mod-MMT (3%) Solvent method at room 82 (3g) (100 g)temp. Mod-MMT (5%) Solvent method at room (5g) (100 g)89 temp.

## Antagonism of pseudomonas flourescens

Discs (5mm) of 7days old mycelium of Fusarium oxysporum f. sp. tuberose.

Were placed in the one side of Petri plates with PDA. The streaking colony of *pseudomonas flourescens* were placed equidistant on the other side from plates. After 7days of inoculation at 25°C the percentages of antagonistic colonies of bacteria inhibit growth of Fusarium oxysporum f. sp. tuberosi mycelium in the whole population was assessed.

## Antagonism of Trichoderma harzianum

The antagonistic effect of Trichoderma harzianum against Fusarium oxysporum f. sp. tuberosi in vitro was examined on Petri plates (9 cm. in diameter) containing potato dextrose agar (PDA) medium . Discs (5mm) of 7days old mycelium of Fusarium oxysporum f. sp. tuberosi was transferred to one side of Petri plates and the other side was inoculated with mycelia disc from Trichoderma harzianum. Five plates were inoculated with the pathogen only as a control . Plates were incubated at 25°C for seven days .

## Antagonism of Montmorillonite particles

(Mod-MMT) solutions of 10, 30 and 50 ppm were prepared . 5 ml of each solution was spread aseptically on the surface of an inoculated PDA Petri dishes with mycelium of Fusarium oxysporum f. sp. tuberosi. Five plates were inoculated with the pathogen only as a control . All Petri dishes were incubated at 25°C for one week in the dark (due to the photocatalytic nature of nanocomposite). Five plates were inoculated with the pathogen only as a control.

The radial growth inhibited percentage of the pathogen was calculated using the equation (Frolich, 1979) of all bio-control agents.

I.P % =  $C - T / C \times 100$  where I.P= inhibition %

C = Radial growth of control

T= Radial growth of treatment

The experiment included the following treatments in laboratory:

- 1- Control
- 2- Trichoderma harzianum
- 3- Pseudomonas fluorescens
- 4- MMT1%
- 5- MMT3%
- 6- MMT5%
- 7- MMT1% + Trichoderma harzianum
- 8- MMT1% + Pseudomonas fluorescens
- 9- MMT3% + Trichoderma harzianum
- 10- MMT3% + Pseudomonas fluorescens
- 11- MMT5% + Trichoderma harzianum
- 12- MMT5% + Pseudomonas fluorescens
- 13- MMT1% + Trichoderma harzianum + Pseudomonas fluorescens
- 14- MMT3% + Trichoderma harzianum + Pseudomonas fluorescens
- 15- MMT5% + Trichoderma harzianum + Pseudomonas fluorescens

## Green house experiment

#### Plant materials

We selected a commercial potato cultivars (Hermis and Spunta cvs.), which are susceptible to *Fusarium* potato wilt. Tubers were surface sterilized with 1% sodium hypochlorite (NaOCl) for 2 min, then triple rinsed with sterilized distilled water.

### **Pot experiment:**

A pot experiment was designed under greenhouse conditions using plastic pots

(15 cm, diameter) each pot containing 8 kg. of sterilized sandy clay soil. Soil was infested with *Fusarium oxysporum f. sp. tuberosi* grown on barely grain at the rate of 5g/kg soil before sowing. Infested pots were irrigated for 5 days before sowing. Three potato tubers (Hermis and Spunta cvs.) were sown in each pot, six replicate pots were specified for each treatment in completely randomized experimental design. The experiment included the treatments as laboratory.

#### The bio- control agents were applied in two methods

#### **First : Tuber treatment**

Spores with along mycelia fragments of *Trichoderma harzianum* and colonies of *Pseudomonas fluorescens* were collected from the surface of agar cultures after 7 days of inoculation by adding 10 ml of sterile distilled water . After collecting by brush, blended the mycelia was filtrated and centrifuged at 3000 rpm for 10 minutes. The spore suspension was adjusted to  $5 \times 10^9$  conidia / ml using haemocytometer for *Trichoderma harzianum* and ( $10^8$  cfu/mL) for *Pseudomonas fluorescens*. potato tubers were wounded and put in each suspensions of previous treatments for half an hour and sown in pathogen inoculated soil. Tubers with no treatments as control.

#### Second : Soil treatments

Suspensions inoculum of the antagonists was mixed with the soil at the rat of 10 ml / pot at planting time. The percentages of pre and post – emergence damping –off were recorded after 20 and 40 days from planting ,respectively, fresh weight of plant, plant height, weight of tubers/ plant and number of tuber / plant.

### Population of Fusarium oxysporum f. sp. tuberosi

To determine the Population of *Fusarium oxysporum* f. sp. *tuberosi* pots and tubers infested with all bio- control agents individually or combine and compare the effect of them on the Population of *Fusarium*. Dilution plate technique were used and the population was expressed as colony forming units (cfu)/gm soil

## Statistical analysis:

The obtained data were statistically analysed according to the method of Gomez and Gomez (1984).

## **Results and Discussion**

## Isolation and identification of Fusarium oxysporum f. sp. tuberosi

The isolated organism were including (10) pathogen isolates belonging to the genus *Fusarium* as cleared by preliminary microscopic examination. The isolated *Fusarium* were identified as *Fusarium* oxysporum f. sp. tuberosi according to (Gilman1957; Burnett and Hunter 1972; Nelson et al., 1983).

## Pathogenicity tests of Fusarium oxysporum f. sp. tuberosi

All isolates of *Fusarium oxysporum f. sp. tuberosi* were pathogenic to all potato cultivars (Hermis and Spunta cvs.) and typical symptoms of *Fusarium* wilt disease were appeared. The symptoms appeared for 25 days after inoculation at 22-25°C. Inoculated potato plants showed yellow color of lower leaves, browning in the vascular stem, ascending wilt symptoms and inoculated plants completely wilted eighty days after planting (Fig 2), this results were similar to (Fakher *et al.*, 2006).

(April-June, 2015)







Fig (2) Pathogenicity tests of Fusarium oxysporum f. sp. tuberosi under greenhouse conditions (A: healthy potato plant and B: inoculated plant by pathogen at 60 days after planting).

### Determine the index of leaf damage ( ILD)

There were significant reduced in index of leaf damage of potato plants( Hermis c.v) that treated with all biocontrol agents compared with control (F. oxysporum f. sp. tuberosi). The index of leaf damage of potato plants treated with Mod-MMT 5% combine with T. harzianum decreased significantly the severity of Fusarium wilt disease and it was 0.15, Mod-MMT 3% combine with T. harzianum 0.36, Mod-MMT 1% combine with T. harzianum 0.46, Mod-MMT 5% combine with P. fluorescens 0.67, MMT3% combine with P. fluorescens 0.91, Mod-MMT1% combine with P. fluorescens 0.96. In the other hand the index of leaf damage of potato plants treated with all biocontrol agents individually were also effective but its effects were lowest than the combined one . The ILD of potato treated with Mod-MMT5 %, Mod-MMT3%, Mod-MMT1%, T. harzianum and P. fluorescens were 1, 1.02,1.03,1.06 and 1.08 respectively compared with control (F. oxysporum f. sp. tuberosi) 3.2. Asimilar results were obtained with Spunta c.v.

### Evaluation antagonistic effect in vitro:

 $T_{111}$  (0)  $T_{111}$ 

The antagonistic activities of all biocontrol agents fungal, bacteria strains and nanocomposites were evaluated on Petri dishes containing PDA medium against F. oxysporum f. sp. tuberosi in vitro. Data in table (2) show that all concentration of nanocomposite (Mod-MMT) and bioagent strains succeeded in reducing the radial growth of F. oxysporum f. sp. tuberosi . Mod-MMT with concentration 5% was more active than Mod-MMT with concentration 3, 1% for reducing the radial growth of F. oxysporum f. sp. tuberosi (0.6, 0.9 and 1 cm respectively) and these value equal to 88.88, 81.88 and 76.33 inhibition percentage respectively. Moreover, T. harzianum inhibited the over growth of F. oxysporum f. sp. tuberosi, by 56.33% comparing with P. fluorescens 51.11%. Both of nano particles and bioagents strains reduced growth percentage comparing with control of F. oxysporum f. sp. tuberosi. These results are agreement with (Howell, 2003)  $(\mathbf{A} \mathbf{A} \mathbf{A} \mathbf{T}) = \mathbf{1} \mathbf{1}$ 

Table (2) Effect of composite (Mi	MT) and bloagents strains on the radial growt	n of F. oxysporum J. sp. tuberosi in vitro.
<b>Bio- agents</b>	Radial growth (cm)	Inhibition (%)

Bio- agents	Radial growth (cm )	Inhibition (%)
T. harzianum	3.93	56.33
P.fluorescens	4.40	51.11
MMT1%	3.0	66.66
MMT3%	2.13	76.33
MMT5%	2.0	77.77
MMT1%+ T. harzianum	1.00	88.88
MMT1% + P.fluorescens	1.9	78.88
MMT3%+ T. harzianum	0.9	90
MMT3% + P.fluorescens	1.8	80
MMT5%+T. harzianum	0.6	93.33
MMT5%+ P.fluorescens	1.8	80
MMT1% + T.harzianum +	- 3.79	57.88
P.fluorescens	3.42	62
MMT1% + T.harzianum +	- 3.21	64.33
P.fluorescens	9.00	00.00
MMT1% + T.harzianum +	-	
P.fluorescens Control		
LSD = 0.5017 at alpha 0.05		

T = Trichoderma

P.= Pseudomonas

MMT = montmorillonite particles

## Green house experiment

## Effect of bio control agents on Fusarium wilt disease

From table (3) we can indicate that, in tuber treatments adding tubers in suspensions of all concentrations of MMT combine with T. harzianum in both two cultivars Hermis and Spunta gave the lowest percentage of pre and post emergence damping off specially with concentration 5% (5.14,1.83, 6.72 and 3.55%) followed by concentration 3% (5.59, 3.19, 7.12 and 5.44) respectively. Also all concentrations of Mod-MMT combine with P. fluorescens reduced the pre and post emergence damping off specially with concentration 5% (10.38, 6.24, 12.71, and 8.12) followed by concentration 3% (14.31,7.31,17.48 and 9.25) compared to control (F. oxysporum f. sp. tuberosi). Whereas P. fluorescens individually gave the highest percentage of damping off in either Hermis and Spunta cultivars (27.01, 18.57

(April-June, 2015)

,31.22 and 19.1 % respectively ) compared to all biocontrol agents. Data in table (4) showed that soil treatments with all tested biocontrol agents gave higher protection against *Fusarium* wilt disease than tuber treatments in both Hermis and Spunta cultivars and significantly reduced the percentage of pre and post emergence damping off from 33.17 and 20.89 to 1.66 and 0 % in Hermis cultivar and from 37.27 and 23.97 to 3.36 and 0% in Spunta cultivar. Also all biocontrol agents increased the percentage of the survivals .The highest percentage of survival was 98.34% and 96.64 % with treatment MMT 5% combine with *T. harzianum* but the lowest one was 64.04 and 58.65 % with treatment *P. fluorescens* individually in Hermis and Spunta cultivars respectively .

 Table (3) Effect of tuber treatments with all tested antagonists on *Fusarium* wilt disease caused by *F. oxysporum f. sp. tuberosi* under greenhouse conditions.

	Tuber treatmen	its				
Antagonists	Hermis cultivar	•		Spunta cultiva	ır	
	Pre-emerg.	Post-emerg.	Survival ]	Pre-emerg.	Post-emerg	Survival
T. harzianum	23.64 c*	16.45 c	59.91 h	27.52 c	16.8 c	55.68 g
P.fluorescens	27.01 b	18.57 b	54.42 h	31.22 b	19.1 b	49.68 h
Mod-MMT1%	22.43 cd	16 c	61.57 f	23.68 de	16.24 cd	60.08 f
Mod-MMT3%	23.27 с	15.5 c	61.23 f	23.08 e	15.8 de	61.12 f
Mod-MMT5%	23.03 cd	15.43 c	61.54 f	22.87 e	15.11 e	62.02 f
Mod-MMT1%+ T. harzianum	8.27 fg	5.04 g	86.69 b	10.53 h	7.39 h	82.08 b
Mod-MMT1%+ P.fluorescens	15.52 e	10.04 e	74.44 e	17.51 f	12.52 f	69.97 e
MMT3%+ T. harzianum	5.59 gh	3.19 h	91.22 a	7.12 i	5.44 i	87.44 a
MMT3%+ P.fluorescens	14.31 e	7.31 f	78.37 d	17.48 f	9.25 g	73.27 d
MMT5%+ T. harzianum	5.14 h	1.83 h	93.03 a	6.72 i	3.55 i	89.73 a
MMT5%+ P.fluorescens	10.38 f	6.24 fg	83.37 c	12.71 g	8.12 h	79.17 с
MMT1%+ T. harzianum	21.66 cd	15.56 c	62.78 f	23.2 e	13.21 f	63.59 f
MMT3%+ T. harzianum + P.fluorescens	21.31 cd	12.48 d	66.21 g	24.63 d	13.35 f	62.02 f
MMT5%+ T. harzianum + P.fluorescens	20.34 d	12.07 d	67.59 g	23.11 e	13.08 f	63.81 f
Control	40.17 a	26.07 a	33.76 i	44.45 a	28.76 a	26.79 i

\*: There are significant differences between carrying different litters.

 Table (4) Effect of soil treatments with all tested antagonists on *Fusarium* wilt disease caused by *F. oxysporum f. sp. tuberosi* under greenhouse conditions

Son treatments													
Antagonists	Ĩ	He	rmis cultiv	var				Spun	ta cultiva	r			
		Pre-en	nerg.	Post-e	merg.	Surviv	al	Pre-em	erg.	Post-e	merg	Surviva	al
T. harzianum		17.44	d*	10.59	cd	71.97	de	22.22	e	15.56	b	62.22	f
P.fluorescens		21.74	b	14.22	b	64.04	e	25.09	b	16.26	b	58.65	g
Mod-MMT1%		19.14	c	8.56	de	73.3	d	20.14	f	11.67	cd	68.19	d
Mod-MMT3%		17.82	d	7.66	e	74.52	d	18.68	g	9.39	e	71.93	d
Mod-MMT5%		15.28	f	5.32	fg	79.4	c	18.49	g	6.23	f	75.28	bc
Mod-MMT1%+ harzianum	Т.	4.51	j	0	h	95.49	a	6.16	j	5.25	fg	88.59	b
Mod-MMT1%+ P.fluorescens		12.42	g	5.13	fg	82.45	bc	16.47	h	6.35	f	77.18	c
Mod-MMT3%+ harzianum	Т.	3.39	j	0	h	96.61	a	4.11	k	0	h	95.89	a
Mod-MMT1%+ P.fluorescens		9.36	h	3.45	g	87.19	b	14.92	i	5.57	fg	79.51	с
Mod-MMT5%+ harzianum	Т.	1.66	k	0	h	98.34	a	3.36	1	0h	g	96.64	a
Mod-MMT5%+		7.73	i	3.41	g	88.86	b	14.5	i	4.74	с	80.76	с

G.J.B.A.H.S., Vol.4	(2):1	151-163	}		(Ar	oril-June	э, 2015)				ISSN: 231	19 – 558	4
P.fluorescens Mod-MMT1%+ harzianum P.fluorescens	T. +	15.2	f	11.49	с	73.31 (	d	23.79	c	12.83	d	63.38 1	f
Mod-MMT3%+ harzianum P.fluorescens	<i>T</i> . +	16.03	ef	11.11	с	72.86	d	23.05	d	10.85	e	66.1 e	
Mod-MMT5%+ harzianum P.fluorescens	<i>T</i> . +	16.63	de	6.8	ef	76.57	с	22.15	e	8.16	a	69.69	d
Control		33.17	a	20.89	a	45.94	f	37.27	a	23.97		38.76	h

\_ . . . . . . .

\*: There are significant differences between carrying different litters.

## **Growth parameters:**

. . . . . . . . . . . . . . . .

### Effect of antagonists on fresh, dry weight and plant height of potato plants

The effect of antagonists on plant growth that presented in Table (5 and 6) reveal that, soil treatments with the antagonists increased the fresh, dry weight and plant height of potato plants more than tuber treatment of both Hermis and Spunta cultivars.

Data showed in Table (5) indicated low values of growth parameters, (fresh, dry weight of plants and plant height, ) with the control treatment (6.45, 8.36, 1.89, 1.95, 21.86 and 26.78) in both two cultivars Hermis and Spunta in comparison with other treatment. The growth parameters of potato plants were significantly increased with the dual inoculation of treatment MMT combine with either *T. harzianum* and *P. fluorescens* compared with the individual one. Data in table (6) revealed that treating the soil with all concentration of MMT5 combine with *T. harzianum* followed by the same treatment with *P. fluorescens* were the most effective in improving fresh, dry weight and plant height from 32.83 to 29.77, 25.41, 24.68, 22.3 and 19.73 gm/ plant of fresh weight respectively in Hermis cultivar; from 11.36, 10.23, 10.11, 9.36, 9.11 and 8.38 gm/ plant of dry weight respectively in Hermis cultivar; from 11.39 to 11.06, 10.93, 8.67, 7.98 and 7.88 gm/ plant of dry weight respectively in Spunta cultivar; from 11.39 to 11.06, 10.93, 8.67 in Spunta cultivar. Treatment of *P. fluorescens* individually was the least effective in this respect, while the other antagonists fall in between.

Table (5): Effect of antagonists on fresh, dry weight/ plant (gm) and plant height (cm) in tuber treatments experiment under greenhouse conditions

Antagonists	Mean of fresh gm)	weight/plant(	Mean of plant(gm)	dry weight/	Mean of pl cm)	lant height (
	Hermis	Spunta	Hermis	Spunta	Hermis	Spunta
T. harzianum	11.36 f*	14.26 g	2.81 c	3.71 d	30.25 f	40.85 f
P.fluorescens	8.67 h	9.64 i	2.22 c	2.81 d	27.64 h	35.56 h
Mod-MMT1%	11.26 f	14.0 g	2.87 c	3.88 d	28.24 g	39.25 g
Mod-MMT3%	13.37 e	14.21 g	3.21 c	3.88 d	29.14 g	42.17 f
Mod-MMT5%	13.36 e	15.36 f	3.16 c	3.87 d	31.25 f	42.36 f
Mod-MMT1%+ T. harzianum	18.66 c	24.37 с	5.11 a	6.87 c	43.58 c	52.36 c
Mod-MMT1%+ <i>P.fluorescens</i>	15.22 d	17.21 e	3.64 b	4.74 b	37.22 e	46.12 e
Mod-MMT3%+ T. harzianum	21.86 b	27.98 b	5.28 a	7.22 a	47.36 b	57.69 b
Mod-MMT3%+ <i>P.fluorescens</i>	15.58 d	17.68 e	3.88 b	5.01 b	41.18 d	46.58 e
Mod-MMT5%+ T. harzianum	23.95 a	31.29 a	6.39 a	8.38 a	51.23 a	59.59 a
Mod-MMT5%+ <i>P.fluorescens</i>	17.36 c	20.69 d	4.65 b	5.63 b	41.21 d	50.34 d
Mod-MMT1%+ T. harzianum +	10.36 g	13.98 g	2.01 d	3.55 d	27.56 h	38.16 g
P.fluorescens						
Mod-MMT3%+ T. harzianum +	10.89 g	13.57 gh	2.66 c	3.57 d	29.48 g	38.15 g
P.fluorescens						
Mod-MMT5%+ T. harzianum +	11.06 f	14.59 g	2.66 c	3.88 d	29.56 g	40.81 f
P.fluorescens						
Control	6.45 i	8.36 i	1.89 e	1.95 e	21.86 i	26.78 i

\*: There are significant differences between carrying different litters.

(April-June, 2015)

Table (6): Effect of antagonists on fresh ,dry weight/ plant (gm) and plant height (cm) in soil treatments experimentunder greenhouse conditions.AntagonistsMean of fresh weight/plant(Mean of fresh weight/plant(Mean of plant height (

	gm)		plant(gm)		cm)	
	TT ·	<b>a</b> .	· ·	C .		<b>a</b> .
	Hermis	Spunta	Hermis	Spunta	Hermis	Spunta
T. harzianum	18.42 f*	22.02 f	7.23 bc	6.32 c	39.54 g	57.36 g
P.fluorescens	14.26 h	16.32 h	4.51 d	3.82 d	30.34 ј	51.34 j
Mod-MMT1%	16.31 g	19.36 g	6.0 c	5.72 c	33.86 i	55.31 h
Mod-MMT3%	17.15 de	21.56 f	6.11 c	5.88 c	36.71 h	57.34 g
Mod-MMT5%	18.14 f	22.18 f	7.25 b	5.62 c	41.82 f	59.57 f
Mod-MMT1%+ T. harzianum	25.41 с	36.57 c	10.11 a	10.93 a	54.14 c	78.63 b
Mod-MMT1%+ <i>P.fluorescens</i>	19.73 e	27.68 e	8.38 b	7.88 bc	44.98. e	64.25 e
Mod-MMT3%+ T. harzianum	29.77 b	40.67 b	10.23 a	11.06 a	59.24 b	79.16 b
Mod-MMT3%+ <i>P.fluorescens</i>	22.38 d	31.48 d	9.11 b	7.98 b	50.32 d	68.45 d
Mod-MMT5%+ T. harzianum	32.83 a	43.89 a	11.36 a	11.39 a	62.78 a	83.18 a
Mod-MMT5%+ <i>P.fluorescens</i>	24.86 c	31.58 d	9.36 b	8.67 b	55.71 c	72.94 c
Mod-MMT1%+ T. harzianum +	17.85 d	19.46 g	5.68 cd	4.95 d	34.24 i	53.48 i
P.fluorescens						
Mod-MMT3%+ T. harzianum +	16.56 g	19.12 g	6.83 c	4.68 d	34.52 i	55.14 h
P.fluorescens						
Mod-MMT5%+ T. harzianum +	18.06 f	21.15 f	6.35 c	5.25 cd	38.82 g	56.57 g
P.fluorescens						
Control	5.43 i	10.36 i	2.12 e	2.97 e	20.14 k	31.54 k

\*: There are significant differences between carrying different litters.

### Effect of antagonists on number and weight of potato plants

Data in table (7 and 8) demonstrate that the application of all antagonists to soil gave the maximum yield as average number and weight of tubers compared with tuber treatments in both Hermis and Spunta cultivars. Treating soil with Mod- MMT5% + *T. harzianum* increased the yield as average number of tubers / plant to 13.68 and 6.32 compared with control 1.79 and 1.04 tubers /plant in Hermis and Spunta, respectively and average weight of tubers to 63.85 and 68.34 gm/plant in Hermis and Spunta, respectively. In this respect *P.fluorescens* gave the minimum yield , whereas other antagonists fall in between.

Table (7) Effect of antagonists on average number of tubers /plant(gm) and average weight of tubers/ plant(gm) in tuber treatments experiment under greenhouse conditions.

Antagonists	Ave. number of	f tubers /plant(	Ave. weight	of tubers/
	gm)		plant(gm)	
	Hermis	Spunta	Hermis	Spunta
T. harzianum	2.34 g*	2.23 g	27.31 de	32.26 de
P.fluorescens	1.4 h	1.167 h	14.39 g	18.57 g
Mod-MMT1%	2.43 g	2.05 g	21.58 f	25.28 f
Mod-MMT3%	3.01 fg	2.29 g	23.15 e	26.27 с
Mod-MMT5%	3.01 fg	2.77 ef	25.39 d	30.25c e
Mod-MMT1%+ T. harzianum	5.51 c	5.11 c	35.36 b	40.97 b
Mod-MMT1%+ <i>P.fluorescens</i>	3.54 ef	3.05 de	28.38 d	33.18 d
Mod-MMT3%+ T. harzianum	6.43 b	5.12 b	36.15 b	42.29 a
Mod-MMT3%+ <i>P.fluorescens</i>	4.04 de	3.23 de	31.19 c	37.28 с
Mod-MMT5%+ T. harzianum	8.2 a	5.12 a	39.26 a	42.36 a
Mod-MMT5%+ P.fluorescens	4.79 cd	3.33 d	31.26 c	39.26 b
Mod-MMT1%+ T. harzianum +	2.69 g	2.37 fg	24.39 d	26.38 с
P.fluorescens				
Mod-MMT3%+ T. harzianum +	2.42 g	2.37 fg	24.26 d	30.56 e
P.fluorescens				
Mod-MMT5%+ T. harzianum +	2.81 fg	2.06 g	23.15 e	32.58 de
P.fluorescens				
Control	1.43 h	1.06 h	8.26 h	9.35 h
		14.00		

\*: There are significant differences between carrying different litters.

(April-June, 2015)

. 1 . 6 . 1 . . . . . .

Table (8) Effect of antagonists on average number of tubers /plant(gm) and average weight of tubers/ plant(gm) in soil treatments experiment under greenhouse conditions.

Antagonists	Ave. number gm)	of tubers /plant(	Ave. weight of t	tubers/ plant(gm)
	Hermis	Spunta	Hermis	Spunta
T. harzianum	6.58 d*	3.89 b	44.39 ef	50.37 f
P.fluorescens	3.66 f	2.167 c	32.69 i	36.57 i
Mod-MMT1%	5.28 e	3.08 b	36.16 h	40.98 h
Mod-MMT3%	6.54 d	3.19 b	40.86 g	45.39 g
Mod-MMT5%	6.38 d	3.26 b	45.19 f	49.37 f
Mod-MMT1%+ T. harzianum	10.26 b	5.01 a	56.37 c	59.36 c
Mod-MMT1%+ <i>P.fluorescens</i>	7.26 d	4.0 a	50.39 e	52.39 e
Mod-MMT3%+ T. harzianum	11.68 b	5.01 a	60.28 b	62.38 b
Mod-MMT3%+ <i>P.fluorescens</i>	8.28 c	4.0 a	51.13 d	53.39 e
Mod-MMT5%+ T. harzianum	13.68 a	6.32 a	63.85 a	68.34 a
Mod-MMT5%+ <i>P.fluorescens</i>	9.34 c	4.16 a	51.37 d	55.12 d
Mod-MMT1%+ T. harzianum +	4.99 e	3.0 b	39.25 ef	41.86 h
P.fluorescens				
Mod-MMT3%+ T. harzianum +	- 5.12 e	3.01 b	41.29 g	47.38 g
P.fluorescens			-	-
Mod-MMT5%+ T. harzianum +	5.68 e	3.16 b	44.35 ef	49.38 f
P.fluorescens				
Control	1.79 g	1.04 d	9.68 j	10.28 j

\*: There are significant differences between carrying different litters.

Effect of tuber and soil treatment with some antagonists on the soil population of Fusarium oxysporum f. sp. tuberosi

Data in Figures (3 to 6) showed that, treating the tubers and soil with all bio control agents significantly decreased the population of Fusarium oxysporum f. sp. tuberosi by time compared with control (Fusarium oxysporum f. sp. tuberosi) in both Hermis and Spunta cultivars. Reduction in Fusarium oxysporum f. sp. tuberosi was investigated when MMT5%+T. harzianum and MMT3%+ P.fluorescens were added to the soil. These bioagents reduced the population of Fusarium oxysporum f. sp. tuberosi from 31.25 and 35.25 to1.15 and 0.5 (cfu)× 10<sup>6</sup> gm/soil after 8 weeks of planting in Hermis and Spunta cultivars respectively .On the other hand the highest values of population was with P.fluorescens only compared with other bioagents . While the population was increased by time in the untreated control from 27.25 and 23.25 to 41.15 and 36.15 (cfu)  $\times 10^6$  gm/soil after 8 weeks of planting in Hermis and Spunta cultivars respectively



Fig (3) Effect of antagonists on Fusarium oxysporum f. sp. tuberosi ( $cfu \times 10^6$  gm/soil) in tubers (Hermis c.v) treatments at different intervals week



Fig (4) Effect of antagonists on Fusarium oxysporum f. sp. tuberosi (cfu× 10<sup>6</sup> gm/soil) in tubers (Spunta c.v) treatments at different intervals week



Fig (5) Effect of antagonists on *Fusarium oxysporum* f. sp. *tuberosi* ( $cfu \times 10^6$  gm/soil) in soil(Spunta c.v) treatments at different intervals week.



(6) Effect of antagonists on *Fusarium oxysporum* f. sp. *tuberosi* ( $cfu \times 10^6$  gm/soil) in soil (Spunta c.v) treatments at different intervals week

```
C=Control
Т.
    =Trichoderma harzianum
Р.
    =Pseudomonas fluorescens
M1 = Mod-MMT1\%
M3 = Mod-MMT3\%
M5 = Mod-MMT5\%
M1+T. = Mod-MMT1% + Trichoderma harzianum
M1 + P. = Mod-MMT1% + Pseudomonas fluorescens
M3+T. = Mod-MMT3% + Trichoderma harzianum
M3 + P. = Mod-MMT3% + Pseudomonas fluorescens
M5+T. = Mod-MMT5% + Trichoderma harzianum
M5 + P. = Mod-MMT5\% + Pseudomonas fluorescens
M1+T.+P. = Mod-MMT1% + Trichoderma harzianum + Pseudomonas fluorescens
M3+T.+P. = Mod-MMT3% + Trichoderma harzianum + Pseudomonas fluorescens
M5+T.+P. = Mod-MMT5% + Trichoderma harzianum + Pseudomonas fluorescens
```

## Discussion

Bio-agents have remarkable capacity of multiplication, thus, when applied; they multiply in exponential ratio and even can overcome stress condition by forming thick walled spores. Hence, bioagents are the solution for safer environmental issues and needs proper attention for seed treatment (Bharath *et al.*, 2005). There are three strategies in considering biological control with introduced biocontrol agents : (a) to reduce the population of the pathogens , (b) to prevent the pathogen to infect the plant , and (c) to limit the disease development after infection (Cook, 1993). *Trichoderma* spp. are among the most - promising bio control agents that can be used against many fungal plant pathogens. *T. harzianum* has multiple mechanisms of action (Benitez *et al.*, 2004), including antibiotics, competition, coparasitism via production of chitinases,  $\beta$ -1-3 glucanases and  $\beta$ - 4 glucanases, , solubilisation of inorganic plant nutrients, induced resistance and inactivation of the pathogen's enzymes involved in the infection process (Sivan and

(April-June, 2015)

Chet, 1993; Altomare *et al.*, 1999; Elad and Kapat, 1999; Morsy, 2005 and Harman, 2006. Our results are agreement with those recorded by(Ercole *et al.*, 1993 and Karunanithi and Usman, 1999). They suggested that the dual culture in Petri-dishes may be useful for detecting the micro-organism as biocontrol agent. The antagonistic effect of *Trichoderma spp.* may be due to faster mycelia growth than pathogenic fungi (Wei *et al.*, 1999). In addition to its produced the non-volatile compounds of ethylene and formic aldehyde (Karunanithi & Usman, 1999).

Also, our results indicat that *P. fluorescens* has great potential to be used as biocontrol agents for the management of the fusarium wilt of potato. *P. fluorescens* is consider from one of PGPR bacteria that have great potential in both phytostimulation and the biocontrol of plant pathogens (Akhtar and Sidiqui, 2009). The results from both soil and tubers treatments that were treated with *P. fluorescens* strains showed varying degrees of bioprotection that of fusarium wilt of potato (Kloepper *et al.*, 1980; Maurhofer *et al.*, 1995; Raaijmakers and Weller, 1998). PGPR antagonize plant pathogenic fungi, mainly by the production of antimicrobial metabolites but also by the competition for iron or rhizosphere niches (Keel et al., 1992) and the stimulation of the host defenses (induced systemic resistance) (Van Loon *et al.*, 1998). Other mechanisms are involved directly in the promotion of plant growth and modulate the biocontrol activity of the bacteria (Lemanceau and Alabouvette, 1993; De Werra *et al.*, 2009).

In this study, MMT was modified by modifies chitosan with different ratio producing nanocomposites with positive charge. The positive charges of these nanocomposites enhance their ability to adsorb bacteria through electrostatic interactions. In addition, there may be hydrophobic interaction between mod-chitosan and lipophilic components of the bacterial cell walls such as lipoproteins, liposaccarides and phospholipids. Therefore, microorganisms in this system can be adsorbed and immobilized on the surface of the nanocomposites and then biocide exerts its effect. And this agree with our result give the highest mean inhibition values (N. Salahuddin,2013)..

The obtained results of this study suggest that the tested microorganisms proved to be an effective bioagents in controlling the tested potato pathogenic fungi and it can reduce the environmental pollution in controlling plant diseases.

#### Conclusion

From the results of present study it is concluded that, although all bio control agents applied individually reduced disease incidence, under laboratory and greenhouse conditions. Species *Trichoderma.harzianum* and *Pseudomonas fluorescens* combine with nanocomposites Mod-MMT5% and Mod-MMT3% have great potential to control *Fusarium* wilt disease caused by *Fusarium oxysporum* f. sp. *tuberosi*.

#### References

Agrios, G.N. 1997. Plant Pathology, 4thEd. Academic Press, San Diego, CA, USA.

Bell, D.;Wells,H.D.and Markham,C.R.1982.In vitro antagonism of Trichoderma species against six fungal plant pathogens. Phytopathology, 72 : 379 – 382.

Alabouvette, C. and C. Steinberg, 1995. Suppressiveness of soils to Invading 15.

microorganisms. In: Biological Control: Benefits and Risks. H.T. Hikkanen and M. Lynch (Eds), Cambridge` University Press, Cambridge, pp: 3-1

Aeshah Mohammed, Laith K.T. AL-Ani, Lyazzat Bekbayeva and Baharuddin Salleh ,2011. Biological Control of Fusarium oxysporum f. sp.cubense by *Pseudomonas fluorescens* and BABA in vitro. World Applied Sciences Journal 15 (2): 189-191, 2011.

Akhtar MS, Siddiqui ZA (2009). Use of plant growth promoting rhizobacteria for the biocontrol of root-rot disease complex of chickpea. Austral. Plant Pathol. 38:44-50.

Altomare C, Norvell W A, Bjbrkman T, Harman GE (1999). Solubilization of phosphates and micronutrients by the Plant growth promoting and biocontrol fungus *Trichoderma harzianum* Rifai 1295 - 22. Appl . Environ. Microbiol.J. 65 2926 – 2933.

Aguzzi C., Cerezo P., Viseras C., Caramella C., Use of clays as drug delivery systems: possibilities and limitations, Appl Clay Sci. 36 (2007) 22-36.

Buchanan, R.E. and N.E. Gibbons, 1974.Bergey's manual of determinative bacteriology. (Eighth edition), the Williams and Wilkins Co., Baltimore, pp: 747-842

Blond, G.j. (1990). Biological control of plant disease with fungal antagonists : challenges and opportunities. Can. J. plant pathology, 12:290-299

Bajwa R, Mukhtar I, AnjumT, 2004. In vitro biological control of *Fusarium solani* cause of wilt in Dalbergia sissoo Roxb. Mycopath, 2(1):11-14

Benitez, T., A.M. Rincon., M. Carmenlimon and A.C. Condon.2004. Biocontrol mechanisms of *Trichoderma* strains. Int. Micro., 7: 249-260.

Bharath, B.G., S. Lokesh and H.S. Shetty. 2005. Effects of fungicides and bioagents on seed mycoflora, growth and yield of watermelon. Integ. Biol. Sci., 9: 75-78.

Beye, I. and J.F. Lafay, 1985. Study of selection criteria for the general resistance in *verticillium* wilt of tomato. Agronomy, 5: 305-311.

Burnett, H.I. and Hunter, B.B. 1972. Illustrated genera of imperfect fungi. Burgess Pub co. Minneapolis, Minnesota, USA pp:241.

Burgess, L.W.; Summerell, B.A.; Bullock, S. G. and Backhouse. K. P. D. 1994: Laboratory Manual for Fusarium Research. 3 rd Ed. University of Sydney. 133 pp.

Bremner J.M., Methods of soil analysis Madison, WI: American Society of Agronomy Inc. Part 2. Agronomy 9 (1982) 1149-1225.

Boyd S.A., Lee J.F., Mortland M.M., Attenuating organic contaminant mobility by soil modification, Nature 333 (1988) 345-47.

Chet, I. (1987). Trichoderma application, mode of action, and potential as a biocontrol agent of soil - borne plant pathogenic fungi. Chet, 1<sup>st</sup> Ed. Innovative Approaches to Plant Disease Control. Wiley InterScience, New York. 137-160.

Cook, R.J. (1993). Making great use of introduced microorganisms for biological control of plant pathogens. The American phytopathological Society: St. Paul, Minnesota, U.S.A

Domsch, K. H., Gams, W., Anderson, T. H., 1980. Compendium of Soil Fungi, Vol. 1. Academic Press, New York

Dennis, C.and Webster, J. (1971). Antagonistic properties of species group of

Trichoderma II. Production of non - volatile antibiotics. Trans. Brit. Mycol. Soc.57: 41-48.

Dohroo, N.P. (1995). Integrated management of vellows of ginger. Indian Phytopath. 48: 90 - 92. Garibaldi, A., Bertetti, D. and Gulinno, M. L. (2009). Susceptibility of chrysanthemum and paris daisy varieties to several isolates of Fusarium oxysporum f. sp. chrysanthemi. Commun Agric Appli Biol Sci. 47: 651 - 657.

De Werra P, Péchy-Tarr M, Keel C, Maurhofer M (2009). Role of Gluconic Acid Production in the Regulation of Biocontrol Traits of Pseudomonas fluorescens CHA0. Appl. Environ. Microbiol. 75:4162-4174.

Davet, P. 1979. Technique analyze for populations of Trichoderma and Gliocladium virens. Annals of Phytopathology, 11 (4): 529-533.

Darder M., Collilla M., Ruiz-Hitzky E., Biopolymer-clay nanocomposites based on chitosan intercalated in montmorillonite, Chem Mater. 15 (2003) 3774-80.

Ercole, D. N., P. Nipoti., D. Manzall and L. Di-Pillo, 1993. In vitro and in vivo activity of Trichoderma spp. against some seeding diseases of horticultural plants. Colture Protette, 22 (1): 63-65.

Elad Y, Kapat A (1999). The role of Trichoderma harzianum protease in the biocontrol of Botrytis cinerea . Eur . J . Plant Pathol. 105 (1999) 177-189.

FAO (2008) International year of the potato [Online]. Available at http://www.potato2008.org (Accessed 10 March 2010). Food and Agriculture Organisation of the United Nations, Rome

Fakher, A., Mejda, D., Hayfa, J., and Mohamed , M., (2006). Potato vascular Fusarium wilt in Tunisia : Incidence and biocontrol by Trichoderma. Plant pathology journal 5(1):92-98.

Farkhondeh Ommati , Masoud Zaker and Alireza Mohammadi . 2013. Biological control of Fusarium wilt of potato (Fusarium oxysporum f. sp. Tuberosi ) by Trichoderma isolates under field condition and their effect on yield. J. Crop Prot, (4): 435-442.

Grondona I, Hermosa R, TejadaM, GomisMD, Mateos PF, Bridge PD, Monte E,

Garcia-AchaI, 1997Physiological and biochemical characterization of Trichoderma harzianum, a biological controlagent against soil-borne fungal plant pathogens. Appl Environ Microbiol63:3189–3198.

Girija G. and A. Manoj, 2005. Pseudomonas fluorescens, a potential bacterial antagonist to control plant diseases. Journal of Plant Interactions, September 2005; 1(3): 123-134.

Gomez, K.A. and Gomez, A. A. 1984. Statistical Procedures for Agricultural

Research . 2 nd E d. John Wiley and Sons, pp. 229 – 308. Gilman, C.J. 1957. Amanual of soil fungi . 2<sup>nd</sup> ed. Lowa state college press. USA pp: 450.

Harris, P. M. 1992. Mineral nutrition. In: Harris, P. M. (Ed.), The Potato Crop: The Scientific Basis for Improvement, second ed. Chapman & Hall, London, pp. 163-213

Haggag H. E. Karima ,2008.Effect of some fungicides on Fusarium solani and Rhizoctonia solani in bean plants. Egyptian Journal of Applied Sciences, 23 (10B); 668-687.

Howell, C. R. 2003. Mechanism employed by Trichoderma species in the biological control of plant diseases: The history and evolution of current concepts. Plant Disease, 87 (1): 4-10

Hanson LE, Howell CR, 2004. Elicitors of plant defense responses from biological control strains of Trichoderma virens. Phytopathology., 94: 171 – 176.

Hoffland E, Halilinen J, Van Pelt JA. 1996. Comparison of systemic resistance induced by avirulant and nonpathogenic Pseudomonas species. Phytopathology 86:757 762.

Hass D, Defago G. 2005. Biological control of soil born pathogens by fluorescent Pseudomonads. Nature Rev Microbiol 3:307 319.

Harman G E (2006) .Overview of mechanisms and uses of Trichodema

spp. Phytopathology.96.190-194.

Hirokawa Y., Tanaka T., Volume phase transition in a nonionic gel, J. Chem Phys 81 (1984) 6379-80.

Jones JB, Jones JP, Stall RE, Zitter TA 1991: Compendium of Tomato Diseases, American Phytopathological Society, St. Paul, MN.

King, E.O., M.K. Ward and D.E. Raney, 1954. Lab. Clin. Med., 44: 301-307.

Kerkeni, A., Mejda, D.R., Neji, T.and Ben, K.M. (2007). In vitro and in vivo

Supp ression of Fusaruim oxysporum f. sp. Radicis - lycopersici the causal agent of fusarium crown and root rot of tomato by some compost fungi. Int. J. Agric. Res. 2: 1022-1029.

Karunanithi, K. and K. M. Usman, 1999. Screening of Trichoderma spp. against Fusarium oxysporium f.sp. sesame causing wilt in seasmum. Crop Research (Histar), 18 (1): 127 - 130.

Keel C, Schnider U, Maurhofer M, Voisard C, Laville J, Burger U, Wirthner P, Haas D, Défago G (1992). Suppression of root diseases by Pseudomonas fluorescens CHA0: importance of the bacterial secondary metabolite 2,4diacetylphloroglucinol. Mol. Plant-Microbe Interact. 5:4-13.

Kloepper JW, Leong J, Teintze M, Schroth MN (1980). Enhanced plant growth by siderophores produced by plant growthpromoting rhizobacteria. Nature 286:885-886.

Khalifa, E. Z. 1991. Biological control of tomato Fusarium wilt by Trichoderma harzianum. Minufiya J. Agric. Res., 16 (2): 1247 - 1259.

Lemanceau P, Alabouvette C (1993). Suppression of fusarium wilts by fluorescent pseudomonads: mechanism and applications. Biocontrol Sci. Technol. 3:219-234.

Maurhofer M, Keel C, Haas D, Défago G (1995). Influence of plant species on disease suppression by Pseudomonas fluorescens strain CHA0 with enhanced antibiotic production. Plant Pathol. 44:40-50.

Morsy, Ebtsam M. 2005. Role of growth promoting substances producing

microorganisms on tomato plant and control of some root rot fungi.

Ph.D. Thesis, Fac. of Agric. Ain shams Univ., Cairo.

Mazzarelli R. A. A., Natural chelating polymers, Oxford: pergamon press; 1973.

Macter W.J., Broussean R., The A.C. response of hardened cement paste, Cement concrete Res. 20 (1990) 891-900.

Mering J., On the hydration of montmorillonite, Trans. Faraday Soc. 42 (1946) 205-219.

Nelson, P. E., Toussoun, T. A. and Marasas, W. F. O. 1983. Fusarium species. An illustrated manual for identification. University Park, The Pennsylvania state university Press, 193 pp.

N. Salahuddin, R. Abdeen , 2013, Drug Release Behavior and Antitumor Efficiency of 5-ASA Loaded Chitosan-Layered Silicate Nanocomposites, J Inorg Organomet Polym, 23:1078-1088

(April-June, 2015)

OrzoLek, M.d., Greaser, G.L., & Harper, J.k.2010.Commercial Vegetable Production Guide.Penn state cooperative extension Agricultural Alternatives: The Pennsylvania state University

O'Sullivan DB, O'Gara F. 1992. Traits of fluorescent Pseudomonas spp. involved in suppression of plant root pathogens. Microbiol Rev 56:662 676.

Orole, O.O and Adejumo T.O. (2009). Activity of fungal endophyte against four maize wilt pathogen. African J. Microbiol. Res. 3: 969-973.

Powelson, M. L. and Rowe, R. C. 1993. Biology and management of early dying of potato. Annual Review of Phytopathology, 31: 111-126.

Pawan and Vijay. (2011). Biological Control of Fusarium wilt of Chrysanthemum with Trichoderma and Botanicals. Journal of Agricultural Technology 2011 Vol. 7(6): 1603-1613.

Rifai, M. A. 1969. A revision of the genus Trichoderma . Mycological Papers, 116: 1-156.

Raaijmakers JM, Weller DM (1998). Natural plant protection by 2,4- diacetylphloroglucinol producing Pseudomonas spp. in take all decline soils. Mol. Plant-Microbe Interact. 11: 144-152.

Ricka J., Tanaka T., Swelling of ionic gels: quantitative performance of the Donnan theory, Macromolecules 17 (12) (1984) 2916-21.

Stevenson W, Loria R, Franc GD, Weingartner DP, 2001. Compendium of Potato Diseases.APS Press, St. Paul, Minnesota, USA

Sivan, A.and Chet, I. (1989). The possible role of competition between

Trichoderma harzianum and Fusarium oxysporum on rhizosphere colonization. Phytopathology 79: 198 - 203.

Sivan A ,Chet I (1993) . Integrated control of Fusarium crown and root of tomato with Trichoderma harzianum in combination with methyl bromide or soil solarization. Crop Protection . 12 . 380 - 386.

Tsegaw T (2005) Response of potato to paclobutrazol and manipulation of reproductive growth under tropical conditions. Ph.D. Thesis., University of Pretoria, South Africa.

Turner, D.; Kovacs, W.; Kuhls, K.; Lieckfeldt, E.; Peter, B.; Arisan Atac, I.; Strauss, J.; Samuels, G.J.; Borner, T. and Kubicek, C. P. 1998. Biogeography and phenotypic variation in Trichoderma sect. 1 ongibraclatum and associated Hypocrea species. Mycol. Res., 101:449-549.

Upadhyay, J.P.and Mukhopadhyay, A.N. (1986). Biological control of

Sclerotium rolfsii by Trichoderma harzianum in sugarbeet. Trop. Pest Management 32: 215 - 220.

Van Loon LC, Bakker PAHM, Pieterse CMJ (1998). Systemic resistance induced by rhizosphere bacteria. Annu. Rev. Phytopathol. 36:453-483.

Weindling, R. 1932. Trichoderma lignorum as a parasite of other soil fungi. Phytopathology, 22: 838-845

Wei G, Kloepper JW, Tuzun S. 1996. Induced systemic resistance to cucumber diseases and increased plant growth by plant growth promoting rhizobacteria under field condition. Phyto-pathology 86:221-224.

Wei, M.W., C. J. Nal, S. Y. Tang, Z. Zhaottai and S. X. Yu, 1999.

Apreliminary study on the growth inhibition effects of Trichoderma spp on six species of soil borne plant pathogenic fungi . Chinese Journal of Biological Control ,15 (13): 142 - 143.