

Bioactive Compounds Derived from Microalgae Showing Antimicrobial Activities

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Abstract

Microalgae have been explored for their bioactive compounds with promising applications encompassing antibacterial, antiviral, antifungal and anti-algal activities. Considering the present status of widely used treatment therapies and their limitations to tackle their adverse effects, the application of bioactive compounds derived from algae will prove beneficial and much more effective as compared with traditional treatment methods. Due to the emerging infectious diseases, viral infections (epidemic and pandemic) and rise in antibiotic resistant bacteria, there is an urgent need for development of alternative treatment therapies against infectious diseases. Present work provides a brief introduction about the algal bioactive compounds and their activities against various pathogens.

Keywords: Microalgae; Antibacterial; Antifungal; Antiviral; Antimicrobial

Introduction

Algae has wide span of ecosystems contributes to the innumerable chemical compounds that they are able to synthesize. A number of antimicrobial compounds have been identified in microalgae as well as macroalgae [1]. More than 18,000 new compounds have been isolated from marine sources, yet majority of them have not yet been obtained nor characterized [2]. Therefore, microalgae represent a unique opportunity to discover novel metabolites. The rate of finding metabolites already obtained from other biological sources is less in microalgae as compared with other microorganisms [3]. Due to their metabolic plasticity under stressed vs. nonstressed conditions

microalgae possess the extra advantage of triggering secondary metabolism [4]. As microalgae were potentially explored only after 1950s, they were not considered previously for therapeutic purposes. Extensive search is presently undergoing to find novel therapeutically useful agents [5-7]. Microalgae have meanwhile been found to produce antibiotics. A large number of microalgal extracts and/or extracellular products have proven antibacterial, antifungal, antiprotozoal and antiplasmodial [6-13]. Efforts to identify the compounds directly responsible for those antimicrobial features have been made, but are still embryonic.

We have been working with algae like *Chlorella* and *Chlamydomonas* (Figure 1) isolated, maintained and extracted as described by Salem et al. [14]. These extracts were later used for antibacterial assay and determination of minimum inhibition concentration (MIC). Antibacterial activity of algal extracts determines the MIC of algae used in this study in vitro [14].

Algal cell-free extracts are already being tested [15-17]. Our aim is to provide information about the recent trends in the discovery of bioactive compounds derived from algae which have shown their potential as antimicrobial agents. We have briefly summarized the recent works carried out by the researchers globally in the field of algal antimicrobial activities.

Antimicrobial activities of algal extracts

Antibacterial activity of algae: The needs for development of alternative antibiotic agent were investigated since the emergence of antibiotic resistant microbes. Due to the emergence of drug-resistant pathogens they endanger people in affluent, industrial societies like the United States, as well as in less-developed nations.

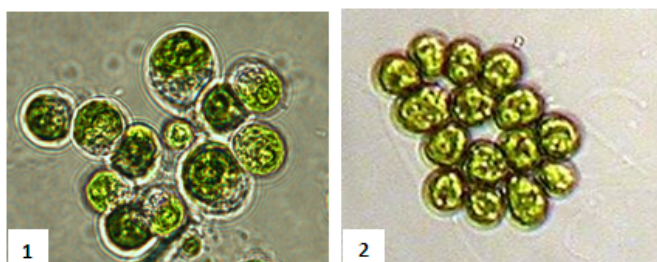


Figure 1: The microscopic images of *Chlorella* sp. (1) and *Chlamydomonas* sp. (2).

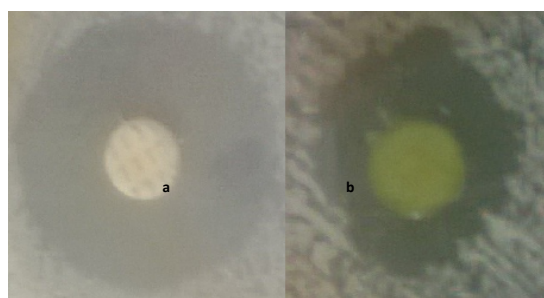


Figure 2: The antibacterial activity of *Chlorella vulgaris* ethanol extract b) against *Staphylococcus* Sp. as compared with the control antibiotic streptomycin (a).

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Examples of clinically important microbes that include *Streptococcus pneumoniae*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella* or *E. coli* and infections transmitted healthcare malpractices enterococci, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Klebsiella* spp [18]. The development in the field of algal therapeutic research has made it possible by their bioactive compounds which have been found effective against most of the pathogens (Figures 1 and 2). The list of some of the algal bioactive compounds is summarized in the Table 1 [19-24].

Antiviral activity of algae: The viruses have been the cause of mass epidemic and pandemic outbreaks of potentially harmful and deadly diseases like influenza, hepatitis, etc. Due to the unavailability of proper treatment facilities and precautionary measures they have been causing a great panic worldwide. Considering the present situation the discovery of antiviral compounds which were derived from algal bioactive compounds provide us a great relief. These compounds, which are tabulated in Table 2 [25-28] has a great prospective in the future.

A number of infectious diseases caused by viruses have re-emerged in recent years, new antiviral measures are necessary for those who are not exposed to them previously. Due to this microalgae have received a strong attention to be explored for potential antiviral agents [29].

Antifungal activity of algae: The study of resistance to antifungal agents has lagged far behind that of antibacterial resistance likely because fungi were not recognized as important pathogens [30,31]. The associated increase in fungal infections prompted search for newer and safer agents to combat fungal infections [32] and a few noteworthy results encompassing microalgae are listed in Table 3 [19,24,33].

Antialgal activity of algae: Inhibitory phenomena between microalgal cells have been reported in the past; Bagchi et al. [34] originally proposed that natural algaecides could effectively be applied in control of toxic algal blooms like *Isochrysis galbana* from cell-free filtrates *Dunaliella salina*, *Platymonas elliptica*, *C. vulgaris*, *Chaetoceros muelleri*, *Chlorella gracilis*, *Nitzschia closterium* and *P. tricorutum* [35].

Algal species	Extract source	Target bacteria	Reference
<i>Pithophora oedogonium</i>	Ethanol extract	<i>Salmonella</i> , <i>Staphylococcus</i> sp., 4978	[19]
<i>Rivularia bullata</i> , <i>Nostoc spongiaeforme</i> , <i>Codium fragile</i> , <i>Colpomenia peregrina</i> Sauvageau, <i>Cystoseira barbata</i> , <i>Zanardinia typus</i>	Methanol Chloroform Diethylether Dichloromethane Ethanol	Gram negative and Gram positive bacteria	[20]
<i>Sargassum wightii</i> , <i>Chaetomorpha linum</i> , <i>Padina gymnospora</i> .	Acetone, methanol	<i>P. aeruginosa</i> (ATCC27853), <i>S. typhi</i> -B, <i>Erwinia amylovora</i> (MTCC2760) (<i>E. amylovora</i>), <i>Enterobacter aerogenes</i> (MTCC111) (<i>E. aerogenes</i>), <i>Proteus vulgaris</i> (MTCC1771) (<i>P. vulgaris</i>), <i>Klebsiella pneumonia</i> (ATCC15380) (<i>K. pneumonia</i>) and <i>E. coli</i> (ATCC25922). gram-positive bacterial strains were Methicillin resistant <i>S. aureus</i> ,	[21]
<i>Asparagopsis taxiformis</i>	Ethanol extract	<i>Vibrio alginolyticus</i> , <i>Vibrio vulnificus</i> and <i>Aeromonas salmonicida</i> subsp. <i>salmonicida</i> <i>Photobacterium damselae</i> subsp. <i>Damselae</i> and <i>Photobacterium damselae</i> subsp. <i>piscicida</i> , <i>Salmonella</i> sp., <i>Vibrio cholerae</i> , <i>Vibrio harveyi</i> and <i>Vibrio parahaemolyticus</i>	[22]
<i>Chlorococcum humicola</i>	Bioactive compounds	<i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Salmonella typhimurium</i> , <i>Klebsiella pneumoniae</i> , <i>Vibrio cholerae</i> , <i>Staphylococcus aureus</i> , <i>Bacillus subtilis</i> , <i>Candida albicans</i> , <i>Aspergillus niger</i> and <i>Aspergillus flavus</i> .	[23]
<i>Gloeocapsa</i> sp. <i>Synechocystis</i> sp. <i>Anabaena</i> sp. <i>Aphanizomenon flos-aquae</i> . <i>Nostoc</i> sp. <i>Nostoc entophyllum</i> , <i>Nostoc muscorum</i> , <i>Scytonema ocellatum</i> , <i>Arthrospira fusiformis</i> (Voronich), <i>Scenedesmus obliquus</i> , <i>Coelastrella</i> sp. <i>Chlorella</i> sp. <i>Rhodella violacea</i> , <i>Porphyridium cruentum</i> (AG.) NAG	Ethanol extract	<i>Staphylococcus aureus</i> 209, <i>Streptococcus pyogenes</i> 981, <i>Bacillus cereus</i> 2421 <i>Escherichia coli</i> 3702, <i>Pseudomonas aeruginosa</i> 1396, <i>Salmonella typhimurium</i> 123, and <i>Yersinia enterocolitica</i> 623	[24]

Table 1: Antibacterial activity of selected compounds from microalgae.

Algal species	Extract source	Target Virus	Reference
<i>Haematococcus Pluvialis</i> and <i>Dunaliella salina</i>	Pressurized liquid extraction	Herpes simplex virus type 1	[25]
<i>Gyrodinium Impudium</i> (sulfated polysaccharide, p-KG03)	Sulfated polysaccharide	Influenza virus	[26]
<i>Navicula directa</i>	Polysaccharide	HSV1 & 2, Influenza A virus	[27]
<i>Gyrodinium impudicum</i>	p-KG03 exopolysaccharides	Encephalomyocarditis Virus	[28]

Table 2: Antiviral activities of selected compounds from microalgae.

Algal species	Extract source	Target fungi	Reference
<i>Pithophora oedogonium</i>	Ethanol extract	<i>Penicillium viridicatum</i> 1101, <i>Fusarium solini</i> 1127	[19]
<i>Gloeocapsa</i> sp.	Exopolysaccharides	<i>Candida albicans</i>	[24]
<i>Haematococcus pluvialis</i>	Butanoic acid and methyl lactate	<i>Candida albicans</i>	[33]

Table 3: Antifungal activities of selected compounds from microalgae.

However, Pratt [36] was the first to report that growth of *C. vulgaris* was depressed by a compound (chlorellin) that was produced and excreted into the medium - and several other extracellular metabolites able to inhibit their own growth and the growth of other species have meanwhile been reported [37].

Antiprotozoal and Antiplasmodial activity of algae: The antiprotozoal activities algal extracts have recently been discovered against *Trypanosoma brucei rhodesiense*, *Trypanosoma cruzi* and *Leishmania donovani* and were found effective. The development of antiprotozoal algal extracts may prove effective in controlling various protozoan diseases and their preventive measures [38]. The crude seaweed extracts from green marine algae *Cladophora rupestris*, *Codium fragile ssp. tomentosoides*, *Ulva intestinalis* and *Ulva lactuca* have shown anti protozoan activity against *Trypanosoma brucei rhodesiense*, *Trypanosoma cruzi*, *Leishmania donovani* [39]. Ciau et al. [40] studied the antiprotozoal activity of brown alga *Lobophora variegata* against *Giardia intestinalis*, *Entamoeba histolytica* and *Trichomonas vaginalis*. They have extracted antiprotozoal compound by chloroform, the major compounds included -O-palmitoyl-2-O-myristoyl-3-O-(6"-sulfo-a-D-quinovopyranosyl)-glycerol; 1,2-di-O-palmitoyl-3-O-(6"-sulfo-a-D-quinovopyranosyl)-glycerol and a new compound identified as 1-O-palmitoyl-2-O-oleoyl-3-O-(6"-sulfo-a-D-quinovopyranosyl)-glycerol [40]. The ethanolic extracts of freshwater macrophytes *Potamogeton perfoliatus*, *Ranunculus tricophyllus* and *Cladophora glomerata* as well as marine macroalgae *Dictyota dichotoma*, *Halopteris scoparia*, *Posidonia oceanica*, *Scinaia furcellata*, *Sargassum natans* and *Ulva lactuca* are assayed for their in vitro antiprotozoal activity against *Trypanosoma brucei rhodesiense*, *Trypanosoma cruzi*, *Leishmania donovani* and *Plasmodium falciparum* [41].

Trypanosomiasis is one of the most important parasitic diseases worldwide. The undesirable side effects and low efficacy of classical trypanocidal drugs underline the necessity of the development of new drugs from natural products. Although marine algae have been recognized as attractive sources of known and novel bioactive compounds, very little research has been focused on antiprotozoal activity. Aqueous and organic extracts of Rhodophyta, Phaeophyta and Chlorophyta were evaluated for their antiprotozoal activity in vitro against *Trypanosoma cruzi* trypomastigotes. The organic extracts from *Dictyota caribea*, *Lobophora variegata*, *Turbinaria turbinata Linnaeus*, and *Laurencia microcladia Kützing* possess promising in vitro activity against *T. cruzi* trypomastigotes. *Laurencia microcladia* is effective against *Artemia salina* and the high cytotoxicity exhibited by *T. turbinata* is required to be investigated further [42].

Red alga from genus *Chondria* produces cyclic polysulfides, terpenoids, amino acids and amines. Domoic acid derivatives from *Chondria armata* show larvicidal and blood pressure lowering activity [43]. The algal extracts have also been explored for their antiplasmodial activities, [38]. The *P. falciparum* (Erythrocytic stages), *T. cruzi* (Trypomastigotes) and *L. donovani* (Axenic amastigotes) are growth inhibited with the ethanol and ethyl acetate extract of algae belonging to Chlorophyta, Heterokontophyta and Rhodophyta. Antimalarial leads from marine algae, four metabolites, sargaquinoic acid, sargahydroquinoic acid, sargaquinal and fucoxanthin, were isolated from the *Sargassum heterophyllum*. Fucoxanthin and sargaquinal showed good antiplasmodial activity toward a chloroquine-sensitive strain of *Plasmodium falciparum* [44] Ethylacetate (EtOAc) extract of *Sargassum swartzii* and *Chondria dasyphylla* were investigated for larvicidal activities in larvae of malaria vector *Anopheles stephensi* and the mortality rate of *Anopheles stephensi* was 96 and 95%, respectively

[45]. The endemic marine red alga *Plocamium cornutum* (Turner) Harvey show antiplasmodial activity in organic extracts. Interestingly, compounds bearing the 7-dichloromethyl substituent showed significantly higher antiplasmodial activity toward a chloroquine sensitive strain of *Plasmodium falciparum* [46].

Conclusion

We have screened the antibacterial activities of organic extracts of isolated culture of algal species and had evaluated them by agar well diffusion method. Methanol extract and ethyl acetate extract of algae were effective against few bacterial species including *Staphylococcus spp.* and *E. coli*. Methanol extracts were more effective as compared with ethyl acetate extract of algae. The antibacterial and antifungal activities were seen predominantly from the *chlorella* sp. as well as *Chlamydomonas* sp. Our work clearly summarizes the importance of microalgal extracts which have potential implication as antibacterial, antiviral, antifungal, antimicrobial antiprotozoal as well as antiplasmodial agents. This information can prove very helpful in further research and discovery of new drugs. The work briefly explains the work carried out by various researchers, clearly elaborating the important implications of algal bioactive compounds for the application against infectious diseases and as an antimicrobial therapy.

References

1. Tandeau-de-Marsac, HJ (1993) Adaptation of cyanobacteria to environmental stimuli: new steps towards molecular mechanisms. FEMS Microbiology Reviews 104: 119-190.
2. Blunt JW, Copp BR, Hu WP, Munro MH, Northcote PT, et al. (2008) Marine natural products. Nat Prod Rep 25: 35-94.
3. Olaizola M (2003) Commercial development of microalgal biotechnology: from the test tube to the marketplace. Biomol Eng 20: 459-466.
4. Guedes AC, Amaro HM, Malcata FX (2011) Microalgae as sources of high added-value compounds—a brief review of recent work. Biotechnol Prog 27: 597-613.
5. Mendes RL, Nobre BP, Cardoso MT, Pereira AP, Palavra AF (2003) Supercritical carbon dioxide extraction of compounds with pharmaceutical importance from microalgae. Inorganica Chimica Acta 356: 328-334.
6. Mayer AMS, Hamann MT (2005) Marine pharmacology in 2001–2002: marine compounds with antihelmintic, antibacterial, anticoagulant, antidiabetic, antifungal, anti-inflammatory, antimalarial, antiplatelet, antiprotozoal, antituberculosis, and antiviral activities; affecting the cardiovascular, immune and nervous systems and other miscellaneous mechanisms of action. Comp Biochem Physiol C Toxicol Pharmacol 140: 265-286.
7. Cardozo KHM, Guaratini T, Barros MP, Falcão VR, Tonon AP, et al. (2007) Metabolites from algae with economical impact. Comp Biochem Physiol C Toxicol Pharmacol 146: 60-78.
8. Kellan SJ, Walker JM (1989) Antibacterial activity from marine microalgae. British Journal of Phycology 23: 41-44.
9. Ozemir G, Karabay NU, Dalay MC, Pazarbasi B (2004) Antibacterial activity of volatile components and various extracts of *Spirulina platensis*. Phytother Res 18: 754-757.
10. Herrero M, Ibañez E, Cifuentes A, Reglero G, Santoyo S (2006) Dunaliella salina microalga pressurized liquid extracts as potential antimicrobials. J Food Prot 69: 2471-2477.
11. Ghasemi Y, Yazdi MT, Shafiee A, Amini M, Shokravi S, et al. (2004) Parsiguine, a novel antimicrobial substance from *Fischerella ambigua*. Pharmaceutical Biology 42: 318-322.
12. Mendiola JA, Torres CF, Martín-Alvarez PJ, Santoyo S, Toré A, et al. (2007) Use of supercritical CO₂ to obtain extracts with antimicrobial activity from *Chaetoceros muelleri* microalga. A correlation with their lipidic content. Eur Food Res Technol 224: 505-510.
13. Metting B, Pyne JW (1986) Biologically active compounds from microalgae. Enzyme and Microbial Technology 8: 386-394.

14. Salem WM, Galal H, Nasr El-deen F (2011) Screening for antibacterial activities in some marine algae from the red sea (Hurghada, Egypt). African Journal of Microbiology Research 5: 2160-2167.
15. Tramper J, Battershill C, Brandenburg W, Burgess G, Hill R, et al. (2003) What to do in marine biotechnology? Biomol Eng 20: 467-471.
16. Trias J, Gordon EM (1997) Innovative approaches to novel antibacterial drug discovery. Curr Opin Biotechnol 8: 757-762.
17. Guedes AC, Barbosa CR, Amaro HM, Pereira CI, Malcata FX (2011) Microalgal and cyanobacterial cell extracts for use as natural antibacterial additives against food pathogens. International Journal of Food Science and Technology 46: 862-870.
18. Interagency Task Force on Antimicrobial Resistance (2011) 2011 Progress Towards Implementation of: A Public Health Action Plan to Combat Antimicrobial Resistance.
19. Danyal A, Mubeen U, Malik KA (2013) Investigating Two Native Algal Species to Determine Antibiotic Susceptibility Against some Pathogens. Curr Res J Biol Sci 5: 70-74
20. Akgul R, Suerdem TB, Akgul F (2013) Antimicrobial Activities of Some Marine Algae and Some Cyanobacteria from Canakkale J. Algal Biomass Utln 4: 35-40
21. Rosaline XD, Sakthivelkumar S, Rajendran K, Janarthanan S (2012) Screening of selected marine algae from the coastal Tamil Nadu, South India for antibacterial activity. Asian Pacific Journal of Tropical Biomedicine S140-S146
22. Genovese G, Faggio C, Gugliandolo C, Torre A, Spanò A, et al. (2012) In vitro evaluation of antibacterial activity of *Asparagopsis taxiformis* from the Straits of Messina against pathogens relevant in aquaculture. Mar Environ Res 73: 1-6.
23. Bhagavathy S, Sumathi P, Bell JS (2011) Green algae *Chlorococcum humicola*-a new source of bioactive compounds with antimicrobial activity. Asian Pacific Journal of Tropical Biomedicine 1: S1-S7.
24. Najdenski HM, Gigova LG, Iliev II, Pilarski PS, Lukavsky J, et al. (2013) Antibacterial and antifungal activities of selected microalgae and cyanobacteria. International Journal of Food Science and Technology 48: 1533-1540.
25. Santoyo S, Jaime L, Plaza M, Herrero M, Rodriguez-Meizoso I, et al. (2012) Antiviral compounds obtained from microalgae commonly used as carotenoid sources. Journal of Applied Phycology 24: 731-741.
26. Kim M, Yim JH, Kim SY, Kim HS, Lee WG, et al. (2012) In vitro inhibition of influenza A virus infection by marine microalga-derived sulfated polysaccharide p-KG03. Antiviral Res 93: 253-259.
27. Lee JB, Hayashi K, Hirata M, Kuroda E, Suzuki E, et al. (2006) Antiviral sulfated polysaccharide from *Navicula directa*, a diatom collected from deep-sea water in Toyama Bay. Biol Pharma Bull 29: 2135-2139.
28. Yim JH, Kim SJ, Ahn SH, Lee CK, Rhie KT, et al. (2004) Antiviral effects of sulfated exo-polysacchride from the marine microalga *Gyrodinium impudicum* strain KG03. Marine Biotechnol 6: 17-25.
29. Borowitzka MA (1995) Microalgae as sources of pharmaceuticals and other biologically active compounds. J Applied Phycol 7: 65-68.
30. Anaissie EJ, Bodey GP (1989) Nosocomial fungal infections - old problems and new challenges. Infectious Disease Clinics of North America 3: 867-882.
31. Wey SB, Mori M, Pfaller M, Woolson RF, Wenzel RP (1988) Hospital-acquired candidemia. The attributable mortality and excess length of stay. Arch Intern Med 148: 2642-2645.
32. Sánchez F, Fernández JM, Acien FG, Rueda A, Perez-Parra J, et al. (2008) Influence of culture conditions on the productivity and lutein content of the new strain *Scenedesmus almeriensis*. Process Biochemistry 43: 398-405.
33. Santoyo S, Rodríguez-Meizoso I, Cifuentes A, Jaime L, García-Blairsy Reina G, et al. (2009) Green processes based on the extraction with pressurized fluids to obtain potent antimicrobials from *Haematococcus pluvialis* microalgae. LWT - Food Science and Technology 42: 1213-1218.
34. Bagchi SN, Palod A, Chauhan VS (1990) Algicidal properties of a bloom-forming blue-green alga, *Oscillatoria* sp. J Basic Microbiol 30: 21-29.
35. Yingying S, Changhai W, Jing C (2008) Growth inhibition of eight species of microalgae by growth inhibitor from the culture of *Isochrysis galbana* and its isolation and identification. J Appl Phycol 20: 315-321.
36. Pratt R (1942) Studies on *Chlorella vulgaris*. V. Some properties of the growth-inhibitor formed by *Chlorella* cells. Amer J Bot 29: 142-148.
37. Hellebust JA (1974) Extracellular products: Algal Physiology and Biochemistry. (Stewart WDP edn), University of California Press, Oxford, 838-863.
38. Vonthron-Sénécheau C, Kaiser M, Devambeiz I, Vastel A, Mussio I, et al. (2011) Antiprotozoal Activities of Organic Extracts from French Marine Seaweeds. Mar Drugs 9: 922-933.
39. Spavieri J, Kaiser M, Casey R, Hingley-Wilson S, Lalvani A, et al. (2010) Antiprotozoal, antimycobacterial and cytotoxic potential of some british green algae. Phytother Res 24: 1095-1098.
40. Cantillo-Ciau Z, Moo-Puc R, Quijano L, Freile-Pelegriñ Y (2010) The Tropical Brown Alga *Lobophora variegata*: A Source of Antiprotozoal Compounds. Marine Drugs 8: 1292-1304.
41. Orhan I, Sener B, Atici T, Brun R, Perozzo R, et al. (2014) Turkish freshwater and marine macrophyte extracts show in vitro antiprotozoal activity and inhibit FabI, a key enzyme of *Plasmodium falciparum* fatty acid biosynthesis. The Free Library.
42. Leon-Deniz LV, Dumontel E, Moo-Puc R, Freile-Pelegriñ Y (2009) Antitrypanosomal in vitro activity of tropical marine algae extracts. Pharmaceutical Biology 47: 864-871.
43. Afolayana AF, Bolton JJ, Lateganc CA, Smithc PJ, Beukesa DR (2008) Fucoxanthin, Tetraprenylated Toluquinone and Tolhydroquinone Metabolites from *Sargassum heterophyllum* Inhibit the in vitro Growth of the Malaria Parasite *Plasmodium falciparum*. Z Naturforsch C 63: 848-852.
44. Govenkar MB, Wahidulla S (2000) Constituents of *Chondria armata*. Phytochem 54: 979-81.
45. Khanavi M, Toulabi PB, Abai MR, Sadati N, Hadjiakhoondi F, et al. (2011) Larvicidal activity of marine algae, *Sargassum swartzii* and *Chondria dasyphylla*, against malaria vector *Anopheles stephensi*. J Vector Borne Dis 48: 241-244.
46. Afolayan AF, Mann MG, Lategan CA, Smith PJ, Bolton JJ, et al. (2009) Antiplasmodial halogenated monoterpenes from the marine red alga *Plocamium cornutum*. Phytochemistry 70: 597-600.