

# Aged Human Skin is More Susceptible than Young Skin to Accumulate Advanced Glycoxidation Products Induced by Sun Exposure

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## Abstract

**Background:** Skin aging is the result of intrinsic and extrinsic phenomena. Among the factors involved in skin aging, the glycation reaction and the formation of advanced glycation end-products (AGEs) is an important element. AGEs such as carboxy-methyl-lysine (CML) and pentosidine, are formed by an oxidative process, and often referred to as glycoxidation products. AGEs have been reported to accumulate during chronological aging but sun exposure has also been shown to contribute to this accumulation likely through induction of an oxidative environment. The purpose of this study was to investigate the accumulation of AGEs in human skin and, more specifically, CML and pentosidine, both of which are generated by oxidative pathways.

**Methods:** CML and pentosidine immunolabelling were investigated in skin samples from both non photo-exposed (sun protected) and photo-exposed (sun exposed) sites from donors of two different age groups (18-25 years and 70-75 years).

**Results:** Results demonstrate CML and pentosidine accumulation in sun-exposed skin especially in the aged group. A vicious circle is envisioned in which the presence of AGEs in a tissue accelerates the formation of additional glycoxidation products following UV-exposure.

**Conclusion:** The damaging effects of UV radiation might be more detrimental in aged skin than in young skin due, in part, to an increased accumulation of AGEs and, in turn, the exacerbation of alterations caused by chronological aging.

**Keywords:** CML; Pentosidine; Glycation; Glycoxidation; Aging skin; Ultraviolet exposure

## Introduction

Skin aging is the result of intrinsic and extrinsic phenomena inducing important alterations in morphological, physical and biological properties of the tissue [1]. Among the factors involved in skin aging, the glycation reaction is an important element. Briefly, this non enzymatic reaction, also known as the Maillard reaction, takes place between free amino groups in proteins (lysine and arginine side groups) and a reducing sugar such as glucose and leads to the formation of advanced glycation end-products (AGEs) [2]. Oxidative environments resulting in the formation of oxoaldehydes, metal-catalyzed auto-oxidation of reducing sugars, and auto-oxidation of Amadori Products induce AGEs formation [3]. AGEs produced by oxidative pathways are referred to as glycoxidation products [4]. Many AGEs have been described and identified in various human tissues including human skin e.g. N-ε-carboxymethyl-lysine (CML) [5], pentosidine [6-9] (Figure 1). Of the many skin proteins found in the skin, dermal proteins making up the extracellular matrix (ECM) with a long half life and a slow turnover, such as collagen, may be particularly affected *in vivo* by this process [10].

Accumulation of AGEs with age is well-known, notably in skin and is believed to modify skin homeostasis [11]. For example, AGEs have been shown to alter fibroblast viability by inducing the apoptotic process [12,13], a senescence phenotype [14] or a cytotoxic effect [15]. Glycation products have also been reported to alter the biology of these dermal cells by disturbing the balance between synthesis of extracellular matrix components and enzymes [16-18] and consequently skin structure. Most of these biological responses could be mediated by Receptor of AGE (RAGE); the AGE-RAGE interaction alters cellular signaling, alters gene expression, enhances the release of pro-inflammatory cytokines and increases intracellular reactive oxygen species (ROS) via

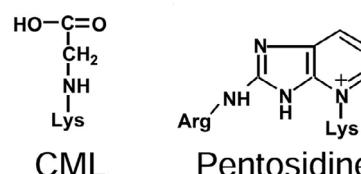


Figure 1: Structures of CarboxyMethyl-Lysine (CML) and pentosidine.

NADPH-oxidase [19]. If biological signature is altered, the mechanical properties are also disrupted [20] and could explain the loss of elasticity observed with age [21].

Skin photo aging is the result of a cumulative process wherein UV-induced damage is superimposed on intrinsic (chronological) skin aging [1]. Particularly, it has been reported that dermal extracellular matrix can be altered by UVA. It has been suggested that cross-linked structures in dermal ECM could act as photo-sensitizers for UVA-induced oxidative damage in skin [22]. Wondrak et al. described endogenous chromophores in human skin as photosensitizers: following absorption

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of solar photons (particularly in the UVA region), these photoexcited components generate ROS, organic free radicals and other toxic photoproducts that mediate skin photo-oxidative stress. Among these components, AGEs have been identified as putative photosensitizers [23]. Following UVA-exposure of pentosidine-rich compounds, hydrogen peroxide ( $H_2O_2$ ) production is increased at a pentosidine concentration-dependent level [24]. Chronic sunlight exposure of the skin induces hyperplasia of elastic tissue in the upper dermis known as actinic elastosis. It has been reported that lesions of actinic elastosis accumulate CML and that ultraviolet-induced oxidation may accelerate CML formation in this case [25]. Jeanmaire et al. illustrated this result by comparing CML accumulation in photo-exposed (glabella) and non photo-exposed (breast) skin from the same donor. They observed higher CML accumulation in the exposed site. Using a 3D-skin model of de-epidermized dermis (DED), they demonstrated *in vitro* that UV-exposure of the DED skin model potentiated glycation [26].

The purpose of our study was to investigate AGEs accumulation, and more specifically CML and pentosidine in human skin. We compared skin specimens from both non photo-exposed and photo-exposed sites from young and old donors.

## Materials and Methods

### Skin samples

Skin samples from two age groups (young=18 -25 years, N=6 and old = 70 -75 years, N=7) of healthy volunteers were analyzed. They were all non-smoking females and Fitzpatrick phototypes between II and IV.

Punch biopsies (3 mm in diameter) were collected from inner (non photo-exposed) and dorsal (photo-exposed) forearms of each subject after approval by an ethics committee and informed written consent.

### Histological staining

Skin biopsies were fixed in neutral formalin and processed for histology. Paraffin sections (5  $\mu$ m) were processed and stained with Hematoxylin Eosin Safron (HES) and Orcein for elastic fibers.

### Immunolabelling

Immunolabelling was performed on the 5  $\mu$ m paraffin sections using mouse monoclonal antibodies against CML (clone CMS-10 from Transgenic reference KH011, dilution 1:50) and pentosidine (clone PEN-12 from Transgenic reference KH012, dilution 1:150). Immunolabelling was performed overnight at room temperature with an amplifactory system Vectastain RTU Universal VECTOR and developed in VIP (kit Vector SK4600).

Immunolabelled tissue sections were observed and imaged under transmission optic microscope (DMR, Leica, Microsystems). Quantitative image analysis was performed using Histolab software (version7.6.0) from Microvision Instruments (Evry, France). Quantification was performed only on the dermal extracellular matrix and cells were excluded. The ratio between the total surface of dermis and the positive surface occupied by the AGEs after immunostaining is reported.

### Statistical methodology

CML and pentosidine expression levels are described by skin type (old versus young) and site (exposed versus non-exposed) using box plots. The bottom and the top of the box represent the first and third quartiles. The horizontal band and the diamond inside the box represent the median and the mean value, respectively. Individual values have

been superposed for completeness. Young vs old donors were compared by zone using exact Wilcoxon tests. Exposed vs unexposed sites were compared according to age group (old vs young) using paired Wilcoxon signed rank tests. All statistical analyses were performed using SAS 9.3 statistical software. The two-sided significance threshold was set at 5%.

## Results

### Skin morphology

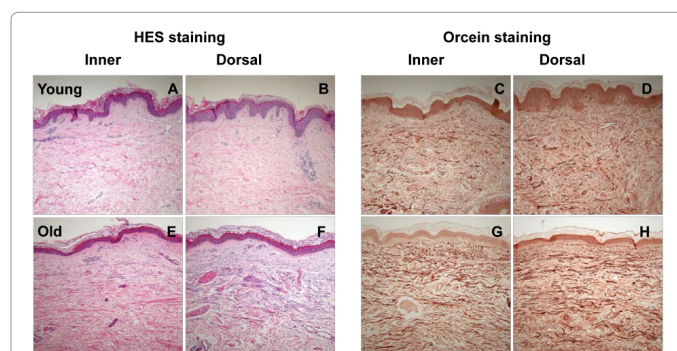
Hematoxylin eosin safron (HES) and orcein staining confirmed the presence of known age-induced skin alterations: decreased epidermal and dermal thickness, flattening of the dermal epidermal junction (DEJ) and elastosis, in particular on the sun-exposed site (Figure 2).

### CML and pentosidine in skin samples

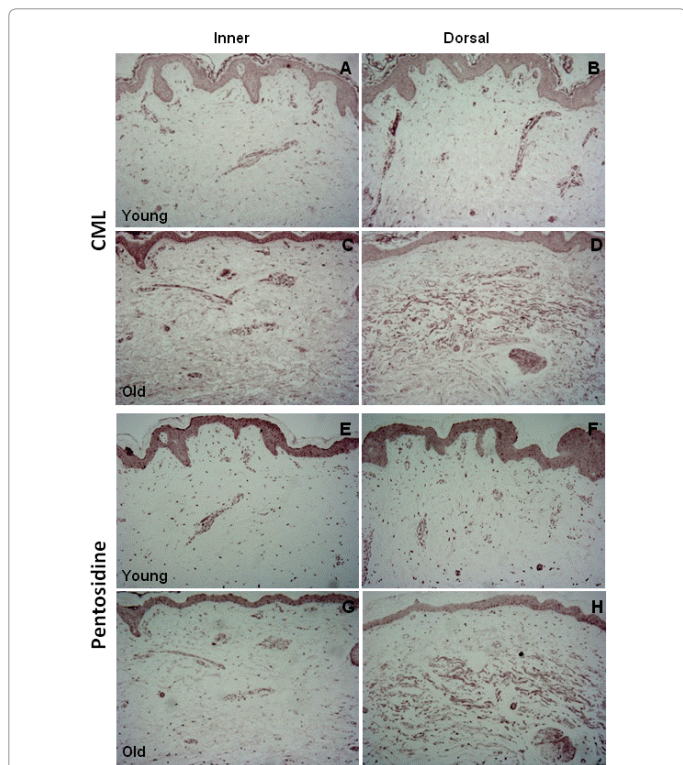
**Difference between non photo-exposed and photo-exposed sites in “young” skin samples:** CML or pentosidine formation in forearm skin of the young group did not vary according to sun exposure (Figure 3A, 3B, 3E and 3F). Indeed, no modifications were observed when comparing inner (Figure 3A and 3E) and dorsal (Figure 3B and 3F) sites of forearm skin. Image analysis showed no statistically significant differences between both areas (Figure 4A and 5A Inner and Dorsal “young” box plot).

**Difference between non photo-exposed and photo-exposed in “old” skin specimens:** In contrast, CML and pentosidine formation in skin of the older group augmented in sun-exposed skin (Figure 3C, 3D, 3G and 3H). Increased positive immunostaining was observed in dorsal (Figure 3D and 3H) compared to inner (Figure 3C and 3G) forearm sites. Image analysis showed a statistically significant difference for pentosidine (Figure 5A Inner and Dorsal “old” box plot,  $p=0.0156$ ) and a tendency ( $p=0.0781$ ) towards increased CML immunostaining (Figure 4A Inner and Dorsal “old” boxplots).

**Comparison of “young” and “old” skin specimens:** An accumulation of AGEs was observed in older skin as compared to young skin (Figure 3). A statistical difference was observed for both pentosidine ( $p=0.0012$ ; Figure 3F and 3H) and CML ( $p=0.0066$ ; Figure 3B and 3D) in dorsal skin as well as for pentosidine ( $p=0.0012$ ; Figure 3E and 3G) in inner skin. A tendency towards a CML (Figure 3A and 3C) increase was observed in inner forearm skin. As noted above, sun exposure in older human skin, increased the accumulation of CML ( $\times 1.5$ , ns  $p=0.0781$ ) and pentosidine ( $\times 2$ ,  $p=0.0156$ ) as showed in the



**Figure 2: Histologic staining of human skin specimens.** Representative skin morphology using Hematoxylin Eosin Safron (HES) staining (A, B, E, F) and elastic fibers using orcein staining (C, D, G, H) of human skin specimens from inner forearm (non photo-exposed) (A, E, C, G) or dorsal forearm (photo-exposed) (B, F, D, H) of “young” (A-D) or “old” (E-H) donors.



**Figure 3: CML and Pentosidine staining**  
 Representative Immunolabelling of CML (A-D) and pentosidine (E-H) of human skin specimens from inner (non photo- exposed) (A, C, E, G) or dorsal forearm (photo-exposed) (B, D, F, H) of “young” (A, B, E, F) or “old” (C, D, G, H) donors. Note that CML and pentosidine were strongly detected in “old” human skin specimens as compared to “young” skin specimens and were potentiated with solar exposure in the former group.

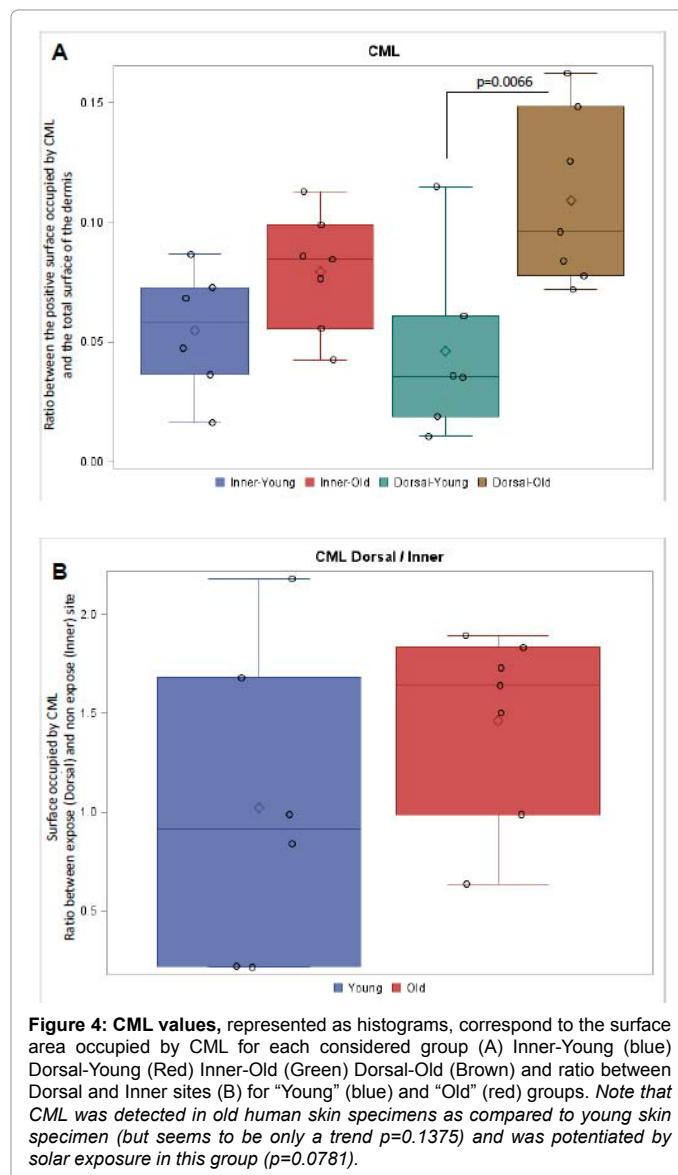
Dorsal/Inner ratio in Figure 4B and 5B whereas no such effect was detected in “young” skin specimens.

## Discussion

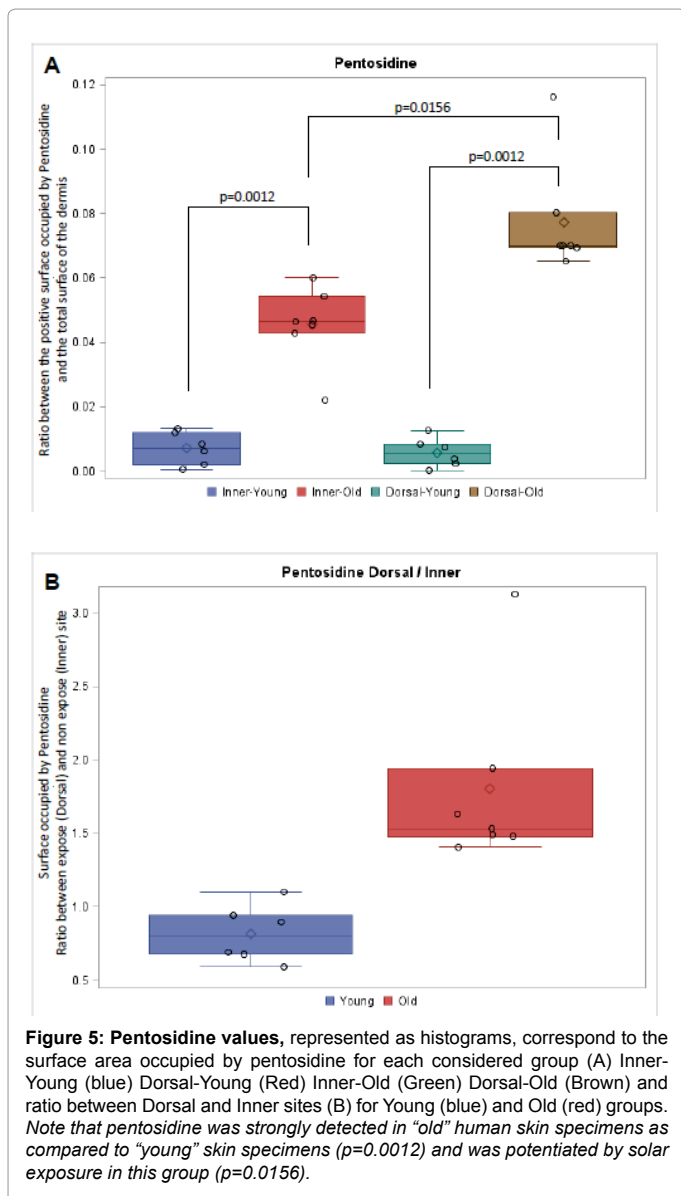
Human skin aging results in decreased elasticity, accumulation of insoluble collagen and impaired wound healing, phenomena which may be explained, in part, by AGEs formation. The protein turn-over rate is important in AGEs accumulation in tissues. In skin, the half-life of collagen has been reported to be fifteen years and consequently this molecule is highly susceptible to AGEs accumulation [5]. Verzijl et al. [10] showed, in normal buttock skin (photo protected) of subjects from 19 to 91 years of age, that CML and pentosidine accumulated linearly with chronological age. Additional reports have also described an increase in these AGEs with solar exposure [25,26]. We show here that sun exposure of the skin increases the abundance of AGEs, specifically CML and pentosidine, and not only in the areas affected by actinic elastosis. We observed age-related AGEs accumulation in the inner face of the forearm and an even higher level in the photo-exposed site (dorsal side of the forearm) of older donors. We believe that AGEs produced by chronological aging may act as photosensitizers [23] and act as an important source of ROS in the dermal matrix facilitating the formation of additional glycoxidation products, such as CML and pentosidine following UV exposure. In support of this theory, Okano et al. demonstrated that pentosidine itself is photo-unstable and produces hydrogen peroxide in the medium after UVA exposure [24]. In addition, oxidative environments have been shown to create a toxic effect on cell

viability in the presence of AGEs as shown by Mazaki on human dermal fibroblasts [27].

The consequences of AGEs accumulation in the skin are likely to be many. Pentosidine cross-linking of proteins chains may be partly responsible for the impairment of skin biomechanical properties [20] and elasticity [21] that has reported with age. CML-collagen has also been reported to induce a time-dependent and dose-dependent increase in fibroblast apoptosis and this process has been shown to be mediated by RAGE [12]. RAGE is highly expressed in skin and up-regulated in sun-exposed skin [28]. The expression of RAGE and AGEs interaction has been reported to increase gene transcription of pro-inflammatory and pro-fibrotic cytokines, chemokines [19] and to be implicated in melanoma process and metastasis [29]. In consequence, AGEs accumulation in the skin increases the possible interaction with RAGE and the following deleterious effects on cells and extracellular matrix. Many studies reported AGEs effects on extracellular matrix maturation and quality through alterations of macromolecules and synthesis of degradation enzymes [11].



**Figure 4: CML values**, represented as histograms, correspond to the surface area occupied by CML for each considered group (A) Inner-Young (blue) Dorsal-Young (Red) Inner-Old (Green) Dorsal-Old (Brown) and ratio between Dorsal and Inner sites (B) for “Young” (blue) and “Old” (red) groups. Note that CML was detected in old human skin specimens as compared to young skin specimen (but seems to be only a trend  $p=0.1375$ ) and was potentiated by solar exposure in this group ( $p=0.0781$ ).



Our results demonstrate the role of solar exposure in CML and pentosidine accumulation with aging. This accumulation may be due to repeated exposure over time but we also believe that the presence of AGEs in older skin specimens may actively promote increased accumulation of glycoxidation products such as CML and pentosidine. A vicious circle is envisioned in which the presence of AGEs in a tissue accelerates the formation of additional AGEs following UV-exposure. We thus speculate that the damaging effects of sun exposure are more adverse in aged skin than in young skin through an enhancement of toxic effect on cells and extracellular matrix and an exacerbation of age-related alterations.

Consequently, the use of molecules with antioxidant activity may help to inhibit or slow down the accumulation of glycoxidation products induced by photo exposure. Previously, we demonstrated the antiglycation effect of a blueberry extract [30] which, indeed, has powerful antioxidant properties due to a rich polyphenols, flavonoids and anthocyanins [31] content. Such ingredients are promising

ingredients for minimizing formation of glycoxidation products and for combating skin aging processes.

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#### References

- Zouboulis CC, Makrantonaki E (2011) Clinical aspects and molecular diagnostics of skin aging. *Clin Dermatol* 29: 3-14.
- Ulrich P, Cerami A (2001) Protein glycation, diabetes, and aging. *Recent Prog Horm Res* 56: 1-21.
- Tessier FJ (2010) The Maillard reaction in the human body. The main discoveries and factors that affect glycation. *Pathol Biol (Paris)* 58: 214-219.
- Dyer DG, Blackledge JA, Katz BM, Hull CJ, Adkisson HD, et al. (1991) The Maillard reaction in vivo. *Z Ernährungswiss* 30: 29-45.
- Dunn JA, Patrick JS, Thorpe SR, Baynes JW (1989) Oxidation of glycated proteins: age-dependent accumulation of N epsilon-(carboxymethyl) lysine in lens proteins. *Biochemistry* 28: 9464-9468.
- Sell DR, Monnier VM (1989) Structure elucidation of a senescence cross-link from human extracellular matrix. Implication of pentoses in the aging process. *J Biol Chem* 264: 21597-21602.
- Sell DR, Nagaraj RH, Grandhee SK, Odetti P, Lapolla A, et al. (1991) Pentosidine: a molecular marker for the cumulative damage to proteins in diabetes, aging, and uremia. *Diabetes Metab Rev* 7: 239-251.
- Dyer DG, Blackledge JA, Thorpe SR, Baynes JW (1991) Formation of pentosidine during nonenzymatic browning of proteins by glucose. Identification of glucose and other carbohydrates as possible precursors of pentosidine in vivo. *J Biol Chem* 266: 11654-11660.
- Dyer DG, Dunn JA, Thorpe SR, Bailie KE, Lyons TJ, et al. (1993) Accumulation of Maillard reaction products in skin collagen in diabetes and aging. *J Clin Invest* 91: 2463-2469.
- Verzijl N, DeGroot J, Thorpe SR, Bank RA, Shaw JN, et al. (2000) Effect of collagen turnover on the accumulation of advanced glycation end products. *J Biol Chem* 275: 39027-39031.
- Pigeon H (2010) Reaction of glycation and human skin: the effects on the skin and its components, reconstructed skin as a model. *Pathol Biol (Paris)* 58: 226-231.
- Alikhani Z, Alikhani M, Boyd CM, Nagao K, Trackman PC, et al. (2005) Advanced glycation end products enhance expression of pro-apoptotic genes and stimulate fibroblast apoptosis through cytoplasmic and mitochondrial pathways. *J Biol Chem* 280:12087-12095.
- Alikhani M, Maclellan CM, Raptis M, Vora S, Trackman PC, et al. (2007) Advanced glycation end products induce apoptosis in fibroblasts through activation of ROS, MAP kinases, and the FOXO1 transcription factor. *Am J Physiol Cell Physiol* 292: C850- C856.
- Ravelojaona V, Robert AM, Robert L (2009) Expression of senescence-associated beta galactosidase (SA-beta-Gal) by human skin fibroblasts, effect of advanced glycation end-products and fucose or rhamnose-rich polysaccharides. *Arch Gerontol Geriatr* 48: 151-154.
- Ravelojaona V, Péterszegi G, Molinari J, Gesztési JL, Robert L (2007) Demonstration of the cytotoxic effect of Advanced Glycation Endproducts (AGE-s). *J Soc Biol* 201: 185-188.
- Okano Y, Masaki H, Sakurai H (2002) Dysfunction of dermal fibroblasts induced by advanced glycation end-products (AGEs) and the contribution of a nonspecific interaction with cell membrane and AGEs. *J Dermatol Sci* 29: 171-180.
- Molinari J, Ruszova E, Velebný V, Robert L (2008) Effect of advanced glycation end products on gene expression profiles of human dermal fibroblasts. *Biogerontology* 9: 177-182.
- Pigeon H, Bakala H, Monnier VM, Asselineau D (2007) Collagen glycation triggers the formation of aged skin in vitro. *Eur J Dermatol* 17: 12-20.
- Barlovic DP, Soro-Paavonen A, Jandeleit-Dahm KA (2011) RAGE biology, atherosclerosis and diabetes. *Clin Sci (Lond)* 121: 43-55.

20. Reihnsner R, Melling M, Pfeiler W, Menzel EJ (2000) Alterations of biochemical and two-dimensional biomechanical properties of human skin in diabetes mellitus as compared to effects of in vitro non-enzymatic glycation. *Clin Biomech (Bristol, Avon)* 15: 379-386.
21. Corstjens H, Dicanio D, Muizzuddin N, Neven A, Sparacio R, et al. (2008) Glycation associated skin autofluorescence and skin elasticity are related to chronological age and body mass index of healthy subjects. *Exp Gerontol* 43: 663-667.
22. Ou-Yang H, Stamatias G, Kollias N (2009) Dermal contributions to UVA-induced oxidative stress in skin. *Photodermatol Photoimmunol Photomed* 25: 65-70.
23. Wondrak GT, Jacobson MK, Jacobson EL (2006) Endogenous UVA-photosensitizers: mediators of skin photodamage and novel targets for skin photoprotection. *Photochem Photobiol Sci* 5: 215-237.
24. Okano Y, Masaki H, Sakurai H (2001) Pentosidine in advanced glycation end-products (AGEs) during UVA irradiation generates active oxygen species and impairs human dermal fibroblasts. *J Dermatol Sci* 27: S11-S18.
25. Mizutani K, Ono T, Ikeda K, Kayashima K, Horiuchi S (1997) Photo-enhanced modification of human skin elastin in actinic elastosis by N(epsilon)-(carboxymethyl)lysine, one of the glycoxidation products of the Maillard reaction. *J Invest Dermatol* 108: 797-802.
26. Jeanmaire C, Danoux L, Pauly G (2001) Glycation during human dermal intrinsic and actinic ageing: an in vivo and in vitro model study. *Br J Dermatol* 145: 10-18.
27. Masaki H, Okano Y, Sakurai H (1999) Generation of active oxygen species from advanced glycation end-products (AGEs) during ultraviolet light A (UVA) irradiation and a possible mechanism for cell damaging. *Biochim Biophys Acta* 1428: 45-56.
28. Lohwasser C, Neureiter D, Weigle B, Kirchner T, Schuppan D (2006) The receptor for advanced glycation end products is highly expressed in the skin and upregulated by advanced glycation end products and tumor necrosis factor-alpha. *J Invest Dermatol* 126: 291-299.
29. Abe R, Shimizu T, Sugawara H, Watanabe H, Nakamura H, et al. (2004) Regulation of human melanoma growth and metastasis by AGE-AGE receptor interactions. *J Invest Dermatol* 122: 461-467.
30. Pageon H, Técher MP, Asselineau D (2008) Reconstructed skin modified by glycation of the dermal equivalent as a model for skin aging and its potential use to evaluate anti-glycation molecules. *Exp Gerontol* 43: 584-588.
31. Faria A, Oliveira J, Neves P, Gameiro P, Santos-Buelga C, et al. (2005) Antioxidant properties of prepared blueberry (*Vaccinium myrtillus*) extracts. *J Agric Food Chem* 53: 6896-6902.