



# Advancing Structural Biology: Dynamic Nuclear Polarization's Role in Biomolecular Investigations

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# INTRODUCTION

In the search of structural biology, Nuclear Magnetic Resonance (NMR) spectroscopy shows as a potential tool for investigating the structures and dynamics of biomolecules [1]. However, the sensitivity of NMR spectroscopy, particularly in solid-state NMR experiments, has often been a limiting factor, especially when distributing with large biomolecular complexes or samples with low concentrations. Dynamic Nuclear Polarization (DNP) has shown as a transformative technique to address this limitation by significantly enhancing the sensitivity of solid-state NMR experiments [2]. This article aims to explore the principles of DNP, its application in biomolecular solid-state NMR, recent advancements, and its potential impact on structural biology.

## DESCRIPTION

#### Principles of dynamic nuclear polarization

Dynamic nuclear polarization is a technique that conducts the large difference in the magnetic moments of electrons and nuclei to transfer the high polarization of electron spins to nuclear spins, thereby increasing the NMR signal. This polarization transfer is achieved through the irradiation of the sample with microwaves at or near the Electron Paramagnetic Resonance (EPR) frequency, which causes the electrons to become highly polarized [3]. Subsequently, this polarization is transferred to nearby nuclei through dipolar interactions or *via* a carrier such as biradicals or nitroxide radicals, resulting in enhanced NMR signals.

#### Application in biomolecular solid-state NMR

The application of DNP in biomolecular solid-state NMR has revolutionized the field by enabling the study of complex biomolecular systems with new sensitivity and resolution. One of the fundamental challenges in solid-state NMR is the low sensitivity arising from the inherently low polarization of nuclear spins in the absence of an external magnetic field. DNP overcomes this limitation by providing a means to transfer the highly polarized electron spins to the nuclei, thus significantly enhancing the Signal-to-Noise Ratio (SNR) of NMR spectra [4]. This section elaborates on the diverse applications of DNP-enhanced solid-state NMR across various biomolecular targets and highlights its transformative impact on structural biology.

**Membrane proteins:** Membrane proteins play vital roles in numerous physiological processes and are prime drug targets for various diseases. However, their structural characterization advances significant challenges due to their insolubility in aqueous environments and tendency to aggregate in crystalline forms [5].

**Protein aggregates:** Protein misfolding and aggregation are characteristic features of numerous neurodegenerative disorders, including Alzheimer's, Parkinson's, and Huntington's diseases. Understanding the structural properties of protein aggregates and their intermediates is essential for understanding the underlying mechanisms of disease pathogenesis [6].

**Amyloid fibrils:** Amyloid fibrils represent a characteristic of protein misfolding diseases and are characterized by their cross- $\beta$  sheet-rich secondary structures [7]. Conventional structural biology techniques often struggle to resolve the complex architecture of amyloid fibrils due to their insolubility and polymorphic nature.

**Protein-RNA complexes:** Protein-RNA interactions play essential roles in gene expression, RNA processing, and regulation of cellular functions. However, the structural characterization of protein-RNA complexes gives significant challenges due to the transient nature of their interactions and the dynamic nature of RNA molecules [8].

**Nanoparticles and biomaterials:** In addition to biomolecules, DNP-enhanced solid-state NMR finds applications in studying complex biomaterials, nanoparticles, and supramolecular assemblies. By providing detailed structural information at the

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atomic level, DNP-enhanced solid-state NMR enables the characterization of nanomaterials for various applications, including drug delivery, tissue engineering, and nanomedicine [9].

#### **Recent advancements**

Recent advancements in DNP technology have further expanded its utility in biomolecular solid-state NMR studies. These include the development of novel polarization agents with improved properties, such as enhanced stability and polarization transfer efficiency. Additionally, significant progress has been made in the design of DNP instrumentation, leading to improvements in microwave irradiation techniques, sample handling, and cryogenic cooling systems. These advancements have collectively contributed to the deep adoption of DNPenhanced solid-state NMR as a routine tool in structural biology laboratories.

#### Potential impact on structural biology

The enhanced sensitivity afforded by DNP has opened up new methods for investigating previously inaccessible biomolecular systems, including membrane proteins, protein aggregates, and amyloid fibrils. By providing detailed structural insights into these challenging targets, DNP-enhanced solid-state NMR has the potential to facilitate drug discovery efforts targeting various diseases, including neurodegenerative disorders and cancer [10]. Moreover, DNP can complement other structural biology techniques such as X-ray crystallography and cryo-electron microscopy, offering unique advantages in studying dynamic and heterogeneous biomolecular assemblies.

### CONCLUSION

Dynamic nuclear polarization has emerged as a transformative technique for enhancing the sensitivity of biomolecular solidstate NMR experiments, thereby enabling the structural characterization of complex biomolecular systems with innovative detail. Recent advancements in DNP technology have further expanded its utility and potential impact on structural biology research. By providing enhanced sensitivity and resolution, DNP-enhanced solid-state NMR holds potential for resolving the structures and dynamics of biomolecules that play critical roles in health and disease. As the field continues to evolve, ongoing efforts to optimize DNP methodologies and instrumentation are prepared to further accelerate progress in structural biology and drug discovery.

### REFERENCES

- Eills J, Budker D, Cavagnero S, Chekmenev EY, Elliott SJ, Jannin S, et al. Spin hyperpolarization in modern magnetic resonance. Chem Rev. 2023;123(4):1417-551.
- Nishiyama Y, Hou G, Agarwal V, Su Y, Ramamoorthy A. Ultrafast magic angle spinning solid-state NMR spectroscopy: Advances in methodology and applications. Chem Rev. 2022;123(3):918-88.
- Abhyankar N, Agrawal A, Campbell J, Maly T, Shrestha P, Szalai V. Recent advances in microresonators and supporting instrumentation for electron paramagnetic resonance spectroscopy. Rev Sci Instrum. 2022;93(10).
- Khelifa M, Mounier D, Yaakoubi N. Design of high performance scroll microcoils for nuclear magnetic resonance spectroscopy of nanoliter and subnanoliter samples. Sensors. 2020;21(1):170.
- Muller MP, Jiang T, Sun C, Lihan M, Pant S, Mahinthichaichan P, et al. Characterization of lipid-protein interactions and lipidmediated modulation of membrane protein function through molecular simulation. Chem Rev. 2019;119(9):6086-161.
- Gonçalves PB, Sodero AC, Cordeiro Y. Green tea epigallocatechin-3-gallate (EGCG) targeting protein misfolding in drug discovery for neurodegenerative diseases. Biomolecules. 2021;11(5): 767.
- Sanderson JM. The association of lipids with amyloid fibrils. J Biol Chem. 2022;298(8).
- Deng L, Yang W, Liu H. PredPRBA: Prediction of protein-RNA binding affinity using gradient boosted regression trees. Front Genet. 2019;10:461677.
- Aguilar-Rabiela AE, Leal-Egaña A, Nawaz Q, Boccaccini AR. Integration of mesoporous bioactive glass nanoparticles and curcumin into PHBV microspheres as biocompatible composite for drug delivery applications. Molecules. 2021;26(11):3177.
- Shimada I, Ueda T, Kofuku Y, Eddy MT, Wüthrich K. GPCR drug discovery: Integrating solution NMR data with crystal and cryo-EM structures. Nat Rev Drug Discov. 2019;18(1):59-82.