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A new method of enhancing the sensitivity of tumor-specific extracellular nanovesicle detection in plasma

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Liquid biopsy is an attractive strategy for diagnosing cancer. Recent studies of the nano-components of circulating blood have opened up a new cancer markers class. For example, extracellular nanovesicles (ENV) are secreted by all types of cells into the extracellular space and circulate with plasma. ENVs retain a tissue-specific pattern of surface proteins. The ENVs secreted by malignant cells are thought to reflect specific characteristics of cancer, and their detection represents a promising approach for cancer diagnosis. Standard approaches to analyze tumor-derived ENVs (tu-ENVs) in plasma suppose consequent steps of ENVs isolation followed by immune-based labeling and quantification. However, this technique is not sensitive enough to detect small fraction of tu-ENVs in total population of plasma vesicles. To improve sensitivity of method, we explored 'inverse' technology of tu-ENVs quantification. Firstly, we colored membrane parts of plasma with lipophilic dye (CM-Dil). Secondly, we isolated population of ENVs by size-exclusion chromatography. Thirdly, we quantified tu-ENVs by flow cytometry with immune-beads. This approach allowed us to increase considerably sensitivity of detection of tu-ENVs in plasma. To confirm this statement we mixed plasma with different quantity of colon cancer cell Colo320 derived ENVs. ENVs were isolated by ultracentrifugation, quantified by NTA, and added to the plasma. Three samples of donor's plasma contained 20*10¹⁰, 10*10¹⁰ and 1*10¹⁰ tu-ENVs were processed by proposed method and amount of ENVs positive for specific colon epithelium marker GPA33 was estimated. We obtained gradient of signal (CV,%): 272, 177, 119. Since portion of artificially mixed tu-ENVs could be estimated in a range of 0,1 – 1% of total amount of plasma ENVs, proposed methods provides with exceptional sensitivity for tu-ENVs detection and might be applied for cancer screening.

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